Chapter 1

1. The hydrogen-bond donors are the $\rm NH$ and $\rm NH_2$ groups.

The hydrogen-bond acceptors are the carbonyl oxygen atoms and those ring nitrogen atoms that are not bonded to hydrogen or to deoxyribose.

2. Interchange the positions of the single and double bonds in the six-membered ring.

- 3. (a) Electrostatic interactions; (b) van der Waals interactions.
- 4. Processes *a* and *b*

5. $\Delta S_{\text{system}} = -661 \text{ J mol}^{-1} \text{ K}^{-1} (-158 \text{ kcal mol}^{-1} \text{ K}^{-1})$

 $\Delta S_{\text{surroundings}} = +842 \text{ J mol}^{-1} \text{ K}^{-1} (+201 \text{ cal mol}^{-1} \text{ K}^{-1})$ 6. (a) 1.0; (b) 13.0; (c) 1.3; (d) 12.7 7. 2.88 8. 1.96 9. 11.83 10. 447; 0.00050 11. 0.00066 M 12. 6.0 13. 5.53 14. 6.48 15. 7.8 16. 100 17. (a) 1.6; (b) 0.51; (c) 0.16.

18. 0.1 M sodium acetate solution: 6.34; 6.03; 5.70; 4.75.

0.01 M sodium acetate solution: 5.90; 4.75; 3.38; 1.40.

19. 90 mM acetic acid; 160 mM sodium acetate, 0.18 moles acetic acid; 0.32 moles sodium acetate; 10.81 g acetic acid; 26.25 g sodium acetate.

20. 0.50 moles of acetic acid; 0.32 moles of NaOH; 30.03 g of acetic acid; 12.80 g of NaOH.

- 21. 250 mM; yes; no, it will also contain 90 mM NaCl.
- 22. 8.63 g Na₂HPO₄; 4.71 g NaH₂PO₄

23. 7.0; this buffer will not be very useful, because the pH value is far from the pK_a value.

- 24. 1.45 kJ mol⁻¹ (0.35 kcal mol⁻¹); 57.9 kJ mol⁻¹ (13.8 kcal mol⁻¹)
- 25. There will be approximately 15 million differences.

Chapter 2

1. (A) Proline, Pro, P; (B) tyrosine, Tyr, Y; (C) leucine, Leu, L; (D) lysine, Lys, K.

- 2. (a) C, B, A; (b) D; (c) D, B; (d) B, D; (e) B.
- 3. (a) 6; (b) 2; (c) 3; (d) 1; (e) 4; (f) 5.
- 4. (a) Ala; (b) Tyr; (c) Ser; (d) His.
- 5. Ser, Glu, Tyr, Thr
- 6. (a) Alanine-glycine-serine; (b) Alanine; (c and d):



At pH 5.5, the net charge is +1

At pH 7.5, the net charge is 0

8. There are 20 choices for each of the 50 amino acids: 50^{20} , or 5×10^{21} .



Aspartame at pH 7

10. The (nitrogen– α carbon–carbonyl carbon) repeating unit. 11. Side chain is the functional group attached to the α -carbon atom of an amino acid.

12. Amino acid composition refers simply to the amino acids that make up the protein. The order is not specified. Amino acid sequence is the same as the primary structure—the sequence of amino acids from the amino terminal to the carboxyl terminal of the protein. Different proteins may have the same amino acid composition, but amino acid sequence identifies a unique protein. 13. (a) Each strand is 35 kd and hence has about 318 residues (the mean residue mass is 110 daltons). Because the rise per residue in an α helix is 1.5 Å, the length is 477 Å. More precisely, for an α -helical coiled coil, the rise per residue is 1.46 Å; so the length is 464 Å. (b) Eighteen residues in each strand (40 minus 4 divided by 2) are in a β -sheet conformation. Because the rise per residue is 3.5 Å, the length is 63 Å.

14. The methyl group attached to the β -carbon atom of isoleucine sterically interferes with α -helix formation. In leucine, this methyl group is attached to the γ -carbon atom, which is farther from the main chain and hence does not interfere.

15. The first mutation destroys activity because valine occupies more space than alanine does, and so the protein must take a different shape, assuming that this residue lies in the closely packed interior. The second mutation restores activity because of a compensatory reduction of volume; glycine is smaller than isoleucine. 16. The native conformation of insulin is not the thermodynamically most stable form, because it contains two separate chains linked by disulfide bonds. Insulin is formed from proinsulin, a single-chain precursor, that is cleaved to form insulin, a 51-residue molecule, after the disulfide bonds have formed.

17. A segment of the main chain of the protease could hydrogen bond to the main chain of the substrate to form an extended parallel or antiparallel pair of β strands.

18. Glycine has the smallest side chain of any amino acid. Its size is often critical in allowing polypeptide chains to make tight turns or to approach one another closely.

19. Glutamate, aspartate, and the terminal carboxylate can form salt bridges with the guanidinium group of arginine. In addition, this group can be a hydrogen-bond donor to the side chains of glutamine, asparagine, serine, threonine, aspartate, tyrosine, and glutamate and to the main-chain carbonyl group. Histidine can form hydrogen bonds with arginine at pH7.

20. Disulfide bonds in hair are broken by adding a thiol-containing reagent and applying gentle heat. The hair is curled, and an oxidizing agent is added to re-form disulfide bonds to stabilize the desired shape.

21. Some proteins that span biological membranes are "the exceptions that prove the rule" because they have the reverse distribution of hydrophobic and hydrophilic amino acids. For example, consider

porins, proteins found in the outer membranes of many bacteria. Membranes are built largely of hydrophobic chains. Thus, porins are covered on the outside largely with hydrophobic residues that interact with the neighboring hydrophobic chains. In contrast, the center of the protein contains many charged and polar amino acids that surround a water-filled channel running through the middle of the protein. Thus, because porins function in hydrophobic environments, they are "inside out" relative to proteins that function in aqueous solution.

22. The amino acids would be hydrophobic in nature. An α helix is especially suited to crossing a membrane because all of the amide hydrogen atoms and carbonyl oxygen atoms of the peptide backbone take part in intrachain hydrogen bonds, thus stabilizing these polar atoms in a hydrophobic environment.

23. This example demonstrates that the pK_a values are affected by the environment. A given amino acid can have a variety of pK_a values, depending on the chemical environment inside the protein. 24. A possible explanation is that the severity of the symptoms corresponds to the degree of structural disruption. Hence, substi-

tution of alanine for glycine might result in mild symptoms, but substitution of the much larger tryptophan might prevent little or no collagen triple-helix formation.

25. The energy barrier that must be crossed to go from the polymerized state to the hydrolyzed state is large even though the reaction is thermodynamically favorable.

26. Using the Henderson–Hasselbalch equation, we find the ratio of alanine-COOH to alanine-COO⁻ at pH 7 to be 10^{-4} . The ratio of alanine-NH₂ to alanine-NH₃⁺, determined in the same fashion, is 10^{-1} . Thus, the ratio of neutral alanine to the zwitterionic species is $10^{-4} \times 10^{-1} = 10^{-5}$.

27. The assignment of absolute configuration requires the assignment of priorities to the four groups connected to a tetrahedral carbon atom. For all amino acids except cysteine, the priorities are: (1) amino group; (2) carbonyl group; (3) side chain; (4) hydrogen. For cysteine, because of the sulfur atom in its side chain, the side chain has a greater priority than does the carbonyl group, leading to the assignment of an R rather than S configuration.

28. ELVISISLIVINGINLASVEGAS

29. No, Pro–X would have the characteristics of any other peptide bond. The steric hindrance in X–Pro arises because the R group of Pro is bonded to the amino group. Hence, in X–Pro, the proline R group is near the R group of X, which would not be the case in Pro–X.

30. A, c; B, e; C, d; D, a; E, b.

31. The reason is that the wrong disulfides formed pairs in urea. There are 105 different ways of pairing eight cysteine molecules to form four disulfides; only one of these combinations is enzymatically active. The 104 wrong pairings have been picturesquely termed "scrambled" ribonuclease.

Chapter 3

1. (a) Phenyl isothiocyanate; (b) urea; β -mercaptoethanol to reduce disulfides; (c) chymotrypsin; (d) CNBr; (e) trypsin. 2. Each amino acid residue, except the carboxyl-terminal residue, gives rise to a hydrazide on reacting with hydrazine. The carboxyl-terminal residue can be identified because it yields a free amino acid.

3. The S-aminoethylcysteine side chain resembles that of lysine. The only difference is a sulfur atom in place of a methylene group. 4. A 1 mg ml⁻¹ solution of myoglobin (17.8 kd; Table 3.2) corresponds to 5.62×10^{-5} M. The absorbance of a 1-cm path length is 0.84, which corresponds to an I_0/I ratio of 6.96. Hence 14.4% of the incident light is transmitted. 5. The sample was diluted 1000-fold. The concentration after dialysis is thus 0.001 M, or 1 mM. You could reduce the salt concentration by dialyzing your sample, now 1 mM, in more buffer free of $(NH_4)_2SO_4$.

6. If the salt concentration becomes too high, the salt ions interact with the water molecules. Eventually, there will not be enough water molecules to interact with the protein, and the protein will precipitate. If there is lack of salt in a protein solution, the proteins may interact with one another—the positive charges on one protein with the negative charges on another or several others. Such an aggregate becomes too large to be solublized by water alone. If salt is added, the salt neutralizes the charges on the proteins, preventing protein—protein interactions.

7. Tropomyosin is rod shaped, whereas hemoglobin is approximately spherical.

8. The frictional coefficient, f, and the mass, m, determine s. Specifically, f is proportional to r (see equation 2 on p. 71). Hence, f is proportional to $m^{1/3}$, and so s is proportional to $m^{2/3}$ (see the equation on p. 76). An 80-kd spherical protein undergoes sedimentation 1.59 times as rapidly as a 40-kd spherical protein.

9. The long hydrophobic tail on the SDS molecule (see p. 72) disrupts the hydrophobic interactions in the interior of the protein. The protein unfolds, with the hydrophobic R groups now interacting with SDS rather than with one another.

10. 50 kd.

11. The protein may be modified. For instance, serine, threonine, and tyrosine may have phosphoryl groups attached.

12. A fluorescence-labeled derivative of a bacterial degradation product (e.g., a formylmethionyl peptide) would bind to cells containing the receptor of interest.

13. (a) Trypsin cleaves after arginine (R) and lysine (K), generating AVGWR, VK, and S. Because they differ in size, these products could be separated by molecular exclusion chromatography. (b) Chymotrypsin, which cleaves after large aliphatic or aromatic R groups, generates two peptides of equal size (AVGW) and (RVKS). Separation based on size would not be effective. The peptide RVKS has two positive charges (R and K), whereas the other peptide is neutral. Therefore, the two products could be separated by ion-exchange chromatography.

14. Antibody molecules bound to a solid support can be used for affinity purification of proteins for which a ligand molecule is not known or unavailable.

15. If the product of the enzyme-catalyzed reaction is highly antigenic, it may be possible to obtain antibodies to this particular molecule. These antibodies can be used to detect the presence of product by ELISA, providing an assay format suitable for the purification of this enzyme.

16. An inhibitor of the enzyme being purified might have been present and subsequently removed by a purification step. This removal would lead to an apparent increase in the total amount of enzyme present.

17. Many proteins have similar masses but different sequences and different patterns when digested with trypsin. The set of masses of tryptic peptides forms a detailed "fingerprint" of a protein that is very unlikely to appear at random in other proteins regardless of size. (A conceivable analogy is: "Just as similarly sized fingers will give different individual fingerprints, so also similarly sized proteins will give different digestion patterns with trypsin.") 18. Isoleucine and leucine are isomers and, hence, have identical masses. Peptide sequencing by mass spectrometry as described in this chapter is incapable of distinguishing these residues. Further analytical techniques are required to differentiate these residues. 19. See the table at the top of the facing page.

A3

Purification procedure	Total protein (mg)	Total activity (units)	Specific activity (units mg ⁻¹)	Purification level	Yield (%)
Crude extract	20,000	4,000,000	200	1	100
(NH ₄) ₂ SO ₄ precipitation	5,000	3,000,000	600	3	75
DEAE-cellulose chromatography	1,500	1,000,000	667	3.3	25
Gel-filtration chromatography	500	750,000	1,500	7.5	19
Affinity chromatography	45	675,000	15,000	75	17

20. Protein crystal formation requires the ordered arrangement of identically positioned molecules. Proteins with flexible linkers can introduce disorder into this arrangement and prevent the formation of suitable crystals. A ligand or binding partner may induce an ordered conformation to this linker and could be included in the solution to facilitate crystal growth. Alternatively, the individual domains separated by the linker may be expressed by recombinant methods and their crystal structures solved separately.

21. Treatment with urea will disrupt noncovalent bonds. Thus the original 60-kd protein must be made of two 30-kd subunits. When these subunits are treated with urea and β -mercaptoethanol, a single 15-kd species results, suggesting that disulfide bonds link the 30-kd subunits.

22. (a) Electrostatic repulsion between positively charged ε-amino groups hinders α-helix formation at pH 7. At pH 10, the side chains become deprotonated, allowing α-helix formation.
(b) Poly-L-glutamate is a random coil at pH 7 and becomes α helical below pH 4.5 because the γ-carboxylate groups become protonated.
23. The difference between the predicted and the observed masses for this fragment equals 28.0, exactly the mass shift that would be expected in a formylated peptide. This peptide is likely formylated at its amino terminus, and corresponds to the most N-terminal

fragment of the protein. 24. Light was used to direct the synthesis of these peptides. Each amino acid added to the solid support contained a photolabile protecting group instead of a *t*-Boc protecting group at its α -amino group. Illumination of selected regions of the solid support led to the release of the protecting group, which exposed the amino groups in these sites to make them reactive. The pattern of masks used in these illuminations and the sequence of reactants define the ultimate products and their locations.

25. Mass spectrometry is highly sensitive and capable of detecting the mass difference between a protein and its deuterated counterpart. Fragmentation techniques can be used to identify the amino acids that retained the isotope label. Alternatively, NMR spectroscopy can be used to detect the isotopically labeled atoms because the deuteron and the proton have very different nuclear-spin properties. 26. First amino acid: A

Last amino acid: R (not cleaved by carboxypeptidase). Sequence of N-terminal tryptic peptide: AVR (tryptic peptide ends in K) Sequence of N-terminal chymotryptic peptide: AVRY (chymotryptic peptide ends in Y) Sequence: AVRYSR

27. First amino acid: S

Last amino acid: L Gyanogen bromide cleavage: M is 10th position, C-terminal residues are: (2S,L,W) Amino-terminal residues: (G,K,S,Y), tryptic peptide, ends in K Amino-terminal sequence: SYGK Chymotryptic peptide order: (S,Y), (G,K,L), (F,I,S), (M,T), (S,W), (S,L) Sequence: SYGKLSIFTMSWSL 28. If the protein did not contain any disulfide bonds, then the electrophoretic mobility of the trypsin fragments would be the same before and after performic acid treatment: all the fragments would lie along the diagonal of the paper. If one disulfide bond were present, the disulfide-linked trypsin fragments would run as a single peak in the first direction, then would run as two separate peaks after performic acid treatment. The result would be two peaks appearing off the diagonal:



These fragments could then be isolated from the chromatography paper and analyzed by mass spectrometry to determine their amino acid composition and thus identify the cysteines participating in the disulfide bond.

Chapter 4

1. A nucleoside is a base attached to a ribose sugar. A nucleotide is a nucleoside with one or more phosphoryl groups attached to the ribose. 2. Hydrogen-bond pairing between the base A and the base T as well as hydrogen-bond pairing between the base G and the base C in DNA.

3. T is always equal to A, and so these two nucleotides constitute 40% of the bases. G is always equal to C, and so the remaining 60% must be 30% G and 30% C.

4. Nothing, because the base-pair rules do not apply to single-stranded nucleic acids.

5. (a) TTGATC; (b) GTTCGA; (c) ACGCGT; (d) ATGGTA. 6. (a) [T] + [C] = 0.46. (b) [T] = 0.30, [C] = 0.24, and [A] + [G] = 0.46.

7. Stable hydrogen bonding occurs only between GC and AT pairs. Moreover, two purines are too large to fit inside the double helix, and two pyrimidines are too small to form base pairs with each other. 8. The thermal energy causes the chains to wiggle about, which disrupts the hydrogen bonds between base pairs and the stacking forces between bases and thereby causes the strands to separate. 9. The probability that any sequence will appear is 4^n , where 4 is the number of nucleotides and *n* is the length of the sequence. The probability of any 15-base sequence appearing is $1/4^{15}$, or 1/1,073,741,824. Thus, a 15-nucleotide sequence would be likely to appear approximately three times (3 billion \times probability of appear-ance). The probability of a 16-base sequence appearing is $1/4^{16}$, which is equal to 1/4,294,967,296. Such a sequence will be unlikely to appear more than once. 10. One end of a nucleic acid polymer ends with a free 5'-hydroxyl group (or a phosphoryl group esterified to the hydroxyl group), and the other end has a free 3'-hydroxyl group. Thus, the ends are different. Two chains of DNA can form a double helix only if the chains are running in different directions—that is, have opposite polarity. 11. Although the individual bonds are weak, the population of thousands to millions of such bonds provides much stability. There is strength in numbers.

12. There would be too much charge repulsion from the negative charges on the phosphoryl groups. These charges must be countered by the addition of cations.

13. The three forms are the A-DNA, the B-DNA and the Z-DNA, with B-DNA being the most common. There are many differences (see Table 4.2). Some key differences are: A-DNA and B-DNA are right-handed, whereas Z-DNA is left-handed. A-DNA forms in less-hydrated conditions than does B-DNA. The A form is shorter and wider than the B form.

14. 5.88×10^3 base pairs

15. In conservative replication, after 1.0 generation, half of the molecules would be ${}^{15}N$.¹⁵N, the other half ${}^{14}N$.¹⁴N. After 2.0 generations, one-quarter of the molecules would be ${}^{15}N$.¹⁵N, the other three-quarters ${}^{14}N$.¹⁴N. Hybrid ${}^{14}N$.¹⁵N molecules would not be observed in conservative replication.

16. (a) Tritiated thymine or tritiated thymidine. (b) dATP, dGTP, dCTP, and TTP labeled with ³²P in the innermost (α) phosphorus atom. 17. Molecules in parts *a* and *b* would not lead to DNA synthesis, because they lack a 3'-OH group (a primer). The molecule in part d has a free 3'-OH group at one end of each strand but no template strand beyond. Only the molecule in part *c* would lead to DNA synthesis. 18. A retrovirus is a virus that has RNA as its genetic material. However, for the information to be expressed, it must first be converted into DNA, a reaction catalyzed by the enzyme reverse transcriptase. Thus, at least initially, information flow is opposite that of a normal cell: $RNA \rightarrow$ DNA rather than DNA \rightarrow RNA. 19. A thymidylate oligonucleotide should

be used as the primer. The poly(A) template specifies the incorporation of T; hence, radioactive thymidine triphosphate (labeled in the α phosphoryl group) should be used in the assay. 20. The ribonuclease serves to degrade the RNA strand, a necessary step in forming duplex DNA from the RNA–DNA hybrid. 21. Treat one aliquot of the sample with ribonuclease and another with deoxyribonuclease. Test these nuclease-treated samples for infectivity. 22. Deamination changes the original G \cdot C base pair into a G \cdot U pair. After one round of rep-

lication, one daughter duplex will contain a $G \cdot C$ pair and the other duplex will contain an $A \cdot U$ pair. After two rounds of replication, there will be two $G \cdot C$ pairs, one $A \cdot U$ pair, and one $A \cdot T$ pair. 23. (a) $4^8 = 65$, 536. In computer terminology, there are 64K 8-mers of DNA.

(b) A bit specifies two bases (say, A and C) and a second bit specifies the other two (G and T). Hence, two bits are needed to specify a single nucleotide (base pair) in DNA. For example, 00, 01, 10, and 11 could encode A, C, G, and T. An 8-mer stores 16 bits ($2^{16} = 65,536$), the *E. coli* genome (4.6×10^6 bp) stores 9.2×10^6 bits, and the human genome (3.0×10^9 bases) stores 6.0×10^9 bits of genetic information.

(c) A standard CD can hold about 700 megabytes, which is equal to 5.6×10^9 bits. A large number of 8-mer sequences could be stored on such a CD. The DNA sequence of *E. coli*, could be written on a single CD with room to spare for a lot of music. One CD would not be quite enough to record the entire human genome.

24. (a) Deoxyribonucleoside triphosphates versus ribonucleoside triphosphates.

(b) $5' \rightarrow 3'$ for both.

(c) Semiconserved for DNA polymerase I; conserved for RNA polymerase.

(d) DNA polymerase I needs a primer, whereas RNA polymerase does not.

25. Messenger RNA encodes the information that, on translation, yields a protein. Ribosomal RNA is the catalytic component of ribosomes, the molecular complexes that synthesize proteins. Transfer RNA is an adaptor molecule, capable of binding a specific amino acid and recognizing a corresponding codon. Transfer RNAs with attached amino acids are substrates for the ribosome. 26. (a) 5'-UAACGGUACGAU-3'

(b) Leu-Pro-Ser-Asp-Trp-Met

(c) Poly(Leu-Leu-Thr-Tyr)

27. The 2'-OH group in RNA acts as an intramolecular nucleophile. In the alkaline hydrolysis of RNA, it forms a 2'-3' cyclic intermediate. 28.



29. Gene expression is the process of expressing the information of a gene in its functional molecular form. For many genes, the functional information is a protein molecule. Thus, gene expression includes transcription and translation.

30. A nucleotide sequence whose bases represent the most-common, but not necessarily the only, members of the sequence. A consensus sequence can be thought of as the average of many similar sequences.31. Cordycepin terminates RNA synthesis. An RNA chain containing cordycepin lacks a 3'-OH group.

32. Only single-stranded RNA can serve as a template for protein synthesis.

33. Degeneracy of the code refers to the fact that most amino acids are encoded by more than one codon.

A 5

34. If only 20 of the 64 possible codons encoded amino acids, then a mutation that changed a codon would likely result in a nonsense codon, leading to termination of protein synthesis. With degeneracy, a nucleotide change might yield a synonym or a codon for an amino acid with similar chemical properties.

35. (a) 2, 4, 8; (b) 1, 6, 10; (c) 3, 5, 7, 9.

36. (a) 3; (b) 6; (c) 2; (d) 5; (e) 7; (f) 1; (g) 4.

37. Incubation with RNA polymerase and only UTP, ATP, and CTP led to the synthesis of only poly(UAC). Only poly(GUA) was formed when GTP was used in place of CTP.

38. A peptide terminating with Lys (UGA is a stop codon), another containing -Asn-Glu-, and a third containing -Met-Arg-. 39. Highly abundant amino acid residues have the most codons (e.g., Leu and Ser each have six), whereas the least-abundant amino acids have the fewest (Met and Trp each have only one). Degeneracy (1) allows variation in base composition and (2) decreases the likelihood that a substitution for a base will change the encoded amino acid. If the degeneracy were equally distributed, each of the 20 amino acids would have three codons. Both benefits (1 and 2) are maximized by the assignment of more codons to prevalent amino acids than to less frequently used ones.

40. Phe-Cys-His-Val-Ala-Ala

41. Exon shuffling is a molecular process that can lead to the generation of new proteins by the rearrangement of exons within genes. Because many exons encode functional protein domains, exon shuffling is a rapid and efficient means of generating new genes.

42. It shows that the genetic code and the biochemical means of interpreting the code are common to even very distantly related life forms. It also testifies to the unity of life; that all life arose from a common ancestor.

43. (a) A codon for lysine cannot be changed to one for aspartate by the mutation of a single nucleotide. (b) Arg, Asn, Gln, Glu, Ile, Met, or Thr.

44. The genetic code is degenerate. Of the 20 amino acids, 18 are specified by more than one codon. Hence, many nucleotide changes (especially in the third base of a codon) do not alter the nature of the encoded amino acid. Mutations leading to an altered amino acid are usually more deleterious than those that do not and hence are subject to more stringent selection.

45. GC base pairs have three hydrogen bonds compared with two for AT base pairs. Thus, the higher content of GC means more hydrogen bonds and greater helix stability.

46. C_0 t value essentially corresponds to the complexity of the DNA sequence—in other words, how long it will take for a sequence of DNA to find its complementary strand to form a double helix. The more complex the DNA, the slower it reassociates to make the double-stranded form.

Chapter 5

1. (a) 5'-GGCATAC-3'

(b) The Sanger dideoxy method of sequencing would give the gel pattern shown here.



Ovalbumin cDNA should be used. *E. coli* lacks the machinery to splice the primary transcript arising from genomic DNA.
 Consistent with its planar, aromatic structure, ethidium bromide is a DNA intercalator: it aligns itself between the paired bases in a DNA duplex.

4. The presence of the *AluI* sequence would, on average, be $(1/4)^4$, or 1/256, because the likelihood of any base being at any position is one-fourth and there are four positions. By the same reasoning, the presence of the *NotI* sequence would be $(1/4)^8$, or 1/65,536. Thus, the average product of digestion by *AluI* would be 250 base pairs (0.25 kb) in length, whereas that by *NotI* would be 66,000 base pairs (66 kb) in length.

No, because most human genes are much longer than 4 kb. A fragment would contain only a small part of a complete gene.
 Southern blotting of an *Mst*II digest would distinguish between the normal and the mutant genes. The loss of a restriction site would lead to the replacement of two fragments on the Southern blot by a single longer fragment. Such a finding would not prove that GTG replaced GAG; other sequence changes at the restriction site could yield the same result.

7. Although the two enzymes cleave the same recognition site, they each break different bonds within the 6-bp sequence. Cleavage by KpnI yields an overhang on the 3' strand, whereas cleavage by Acc65I produces an overhang on the 5' strand. These sticky ends do not overlap.



8. A simple strategy for generating many mutants is to synthesize a degenerate set of cassettes by using a mixture of activated nucleosides in particular rounds of oligonucleotide synthesis. Suppose that the 30-bp coding region begins with GTT, which encodes valine. If a mixture of all four nucleotides is used in the first and second rounds of synthesis, the resulting oligonucleotides will begin with the sequence XYT (where X and Y denote A, C, G, or T). These 16 different versions of the cassette will encode proteins containing either Phe, Leu, Ile, Val, Ser, Pro, Thr, Ala, Tyr, His, Asn, Asp, Cys, Arg, or Gly at the first position. Likewise, degenerate cassettes can be made in which two or more codons are simultaneously varied. 9. Because PCR can amplify as little as one molecule of DNA, statements claiming the isolation of ancient DNA need to be greeted with some skepticism. The DNA would need to be sequenced. Is it similar to human, bacterial, or fungal DNA? If so, contamination is the likely source of the amplified DNA. Is it similar to that of birds or crocodiles? This sequence similarity would strengthen the case that it is dinosaur DNA because these species are evolutionarily close to dinosaurs.

10. PCR amplification is greatly hindered by the presence of G–C-rich regions within the template. Owing to their high melting temperatures, these templates do not denature easily, preventing the initiation of an amplification cycle. In addition, rigid secondary

structures prevent the progress of DNA polymerase along the template strand during elongation.

11. At high temperatures of hybridization, only very close matches between primer and target would be stable because all (or most) of the bases would need to find partners to stabilize the primertarget helix. As the temperature is lowered, more mismatches would be tolerated; so the amplification is likely to yield genes with less sequence similarity. In regard to the yeast gene, synthesize primers corresponding to the ends of the gene, and then use these primers and human DNA as the target. If nothing is amplified at 54°C, the human gene differs from the yeast gene, but a counterpart may still be present. Repeat the experiment at a lower temperature of hybridization.

12. Digest genomic DNA with a restriction enzyme, and select the fragment that contains the known sequence. Circularize this fragment. Then carry out PCR with the use of a pair of primers that serve as templates for the synthesis of DNA away from the known sequence.

13. The encoded protein contains four repeats of a specific sequence.

14. Use chemical synthesis or the polymerase chain reaction to prepare hybridization probes that are complementary to both ends of the known (previously isolated) DNA fragment. Challenge clones representing the library of DNA fragments with both of the hybridization probes. Select clones that hybridize to one of the probes but not the other; such clones are likely to represent DNA fragments that contain one end of the known fragment along with the adjacent region of the particular chromosome.

15. The codon(s) for each amino acid can be used to determine the number of possible nucleotide sequences that encode each peptide sequence (see Table 4.5):

Ala-Met-Ser-Leu-Pro-Trp: $4 \times 1 \times 6 \times 6 \times 4 \times 1 = 576$ total sequences Gly-Trp-Asp-Met-His-Lys: $4 \times 1 \times 2 \times 1 \times 2 \times 2 = 32$ total sequences

Cys–Val–Trp–Asn–Lys–Ile: $2 \times 4 \times 1 \times 2 \times 2 \times 3 = 96$ total sequences

Arg–Ser–Met–Leu–Gln–Asn: $6 \times 6 \times 1 \times 6 \times 2 \times 2 = 864$ total sequences

The set of DNA sequences encoding the peptide Gly-Trp-Asp-Met-His-Lys would be most ideal for probe design because it encompasses only 32 total oligonucleotides.

16. Within a single species, individual dogs show enormous variation in body size and substantial diversity in other physical characteristics. Therefore, genomic analysis of individual dogs would provide valuable clues concerning the genes responsible for the diversity within the species.

17. On the basis of the comparative genome map shown in Figure 5.27, the region of greatest overlap with human chromosome 20 can be found on mouse chromosome 2.

18. $T_{\rm m}$ is the melting temperature of a double-stranded nucleic acid. If the melting temperatures of the primers are too different, the extent of hybridization with the target DNA will differ during the annealing phase, which would result in differential replications of the strands.

19. Careful comparison of the sequences reveals that there is a 7-bp region of complementarity at the 3' ends of these two primers:

In a PCR experiment, these primers would likely anneal to one another, preventing their interaction with the template DNA. During DNA synthesis by the polymerase, each primer would act as a template for the other primer, leading to the amplification of a 25-bp sequence corresponding to the overlapped primers. 20. A mutation in person B has altered one of the alleles for gene X, leaving the other intact. The fact that the mutated allele is smaller suggests that a deletion has occurred in one copy of the gene. The one functioning copy is transcribed and translated and apparently produces enough protein to render the person asymptomatic.

Person C has only the smaller version of the gene. This gene is neither transcribed (negative northern blot) nor translated (negative western blot).

Person D has a normal-size copy of the gene but no corresponding RNA or protein. There may be a mutation in the promoter region of the gene that prevents transcription.

Person E has a normal-size copy of the gene that is transcribed, but no protein is made, which suggests that a mutation prevents translation. There are a number of possible explanations, including a mutation that introduced a premature stop codon in the mRNA.

Person F has a normal amount of protein but still displays the metabolic problem. This finding suggests that the mutation affects the activity of the protein—for instance, a mutation that compromises the active site of enzyme Y.

21. Chongqing: residue 2, $L \rightarrow R$, CTG \rightarrow CGG Karachi: residue 5, $A \rightarrow P$, GCC \rightarrow CCC Swan River: residue 6, $D \rightarrow G$, GAC \rightarrow GGC

22. This particular person is heterozygous for this particular mutation: one allele is wild type, whereas the other carries a point mutation at this position. Both alleles are PCR amplified in this experiment, yielding the "dual peak" appearance on the sequencing chromatogram.

Chapter 6

1. There are 26 identities and two gaps for a score of 210. The two sequences are approximately 26% identical. This level of homology is likely to be statistically significant.

2. They are likely related by divergent evolution, because three-dimensional structure is more conserved than is sequence identity.
3. (a) Identity score = -25; Blosum score = 14; (b) identity score = 15; Blosum score = 3.

4. U



5. There are 4^{40} , or 1.2×10^{24} , different molecules. Each molecule has a mass of 2.2×10^{-20} , because 1 mol of polymer has a mass of 330 g mol⁻¹ × 40, and there are 6.02×10^{23} molecules per mole. Therefore, 26.4 kg of RNA would be required.

6. Because three-dimensional structure is much more closely associated with function than is sequence, tertiary structure is more evolutionarily conserved than is primary structure. In other words, protein function is the most important characteristic, and protein function is determined by structure. Thus, the structure must be conserved but not necessarily a specific amino acid sequence. 7. Alignment score of sequences (1) and (2) is $6 \times 10 = 60$. Many answers are possible, depending on the randomly reordered sequence. A possible result is

Shuffled sequence: (2) **TKADKAGEYL**

Alignment:	(1) ASNFLDKAGK
	(2) $\texttt{TKADKAGEYL}$

Alignment score is $4 \times 10 = 40$.

8. (a) Almost certainly diverged from a common ancestor.
(b) Almost certainly diverged from a common ancestor. (c) May have diverged from a common ancestor, but the sequence alignment may not provide supporting evidence. (d) May have diverged from a common ancestor, but the sequence alignment is unlikely to provide supporting evidence.

9. Replacement of cysteine, glycine, and proline never yields a positive score. Each of these residues exhibits features unlike those of its other 19 counterparts: cysteine is the only amino acid capable of forming disulfide bonds, glycine is the only amino acid without a side-chain and is highly flexible, and proline is the only amino acid that is highly constrained through the bonding of its side chain to its amine nitrogen.

10. Protein A is clearly homologous to protein B, given 65% sequence identity, and so A and B are expected to have quite similar three-dimensional structures. Likewise, proteins B and C are clearly homologous, given 55% sequence identity, and so B and C are expected to have quite similar three-dimensional structures. Thus, proteins A and C are likely to have similar three-dimensional structures, even though they are only 15% identical in sequence. 11. The likely secondary structure is



12. To detect pairs of residues with correlated mutations, there must be variability in these sequences. If the alignment is over-represented by closely related organisms, there may not be enough changes in their sequences to allow the identification of potential base-pairing patterns.

13. After RNA molecules have been selected and reverse transcribed, PCR is performed to introduce additional mutations into these strands. The use of this error-prone, thermostable polymerase in the amplification step would enhance the efficiency of this random mutagenesis.

14. The initial pool of RNA molecules used in a molecular-evolution experiment is typically much smaller than the total number of possible sequences. Hence, the best possible RNA sequences will likely not be represented in the initial set of oligonucleotides. Mutagenesis of the initial selected RNA molecules allows for the iterative improvement of these sequences for the desired property.

15. 107 or 108 identities (depending on which annotated human sequence is chosen).

Chapter 7

1. The whale swims long distances between breaths. A high concentration of myoglobin in the whale muscle maintains a ready supply of oxygen for the muscle between breathing episodes. 2. (a) 2.96×10^{-11} g

(b)
$$2.74 \times 10^8$$
 molecule

(c) No. There would be 3.17×10^8 hemoglobin molecules in a red cell if they were packed in a cubic crystalline array. Hence, the actual packing density is about 84% of the maximum possible. 3. 2.65 g (or 4.75×10^{-2} mol) of Fe

4. (a) In human beings, 1.44×10^{-2} g (4.49×10^{-4} mol) of O₂ per kilogram of muscle. In sperm whale, 0.144 g (4.49×10^{-3} mol) of O₂ per kilogram.

(b) 128

5. The pK_a is (a) lowered; (b) raised; and (c) raised.

6. Deoxy Hb A contains a complementary site, and so it can add on to a fiber of deoxy Hb S. The fiber cannot then grow further, because the terminal deoxy Hb A molecule lacks a sticky patch. 7. 62.7% oxygen-carrying capacity

8. A higher concentration of BPG would shift the oxygen-binding curve to the right, causing an increase in P_{50} . The larger value of P_{50} would promote dissociation of oxygen in the tissues and would thereby increase the percentage of oxygen delivered to the tissues. 9. Oxygen binding appears to cause the copper ions and their associated histidine ligands to move closer to one another, thereby also moving the helices to which the histidines are attached (in similar fashion to the conformational change in hemoglobin). 10. The modified hemoglobin should not show cooperativity. Although the imidazole in solution will bind to the heme iron (in place of histidine) and will facilitate oxygen binding, the imidazole lacks the crucial connection to the particular α helix that must move so as to transmit the change in conformation.

11. Inositol pentaphosphate (part c) is highly anionic, much like 2,3-bisphosphoglycerate.

12.



13. Release of acid will lower the pH. A lower pH promotes oxygen dissociation in the tissues. However, the enhanced release of oxygen in the tissues will increase the concentration of deoxy-Hb, thereby increasing the likelihood that the cells will sickle. 14. (a) Y = 0.5 when $pO_2 = 10$ torr. The plot of Y versus pO_2 appears to indicate little or no cooperativity.

(b) The Hill plot shows slight cooperativity with $n \cdot 1.3$ in the central region.

(c) Deoxy dimers of lamprey hemoglobin could have lower affinity for oxygen than do the monomers. If the binding of the first oxygen atom to a dimer causes dissociation of the dimer to give two monomers, then the process would be cooperative. In this mechanism, oxygen binding to each monomer would be easier than binding the first oxygen atom to a deoxy dimer.

15. (a) 2; (b) 4; (c) 2; (d) 1.

16. The electrostatic interactions between BPG and hemoglobin would be weakened by competition with water molecules. The T state would not be stabilized.

Chapter 8

- 1. Rate enhancement and substrate specificity
- 2. A cofactor
- 3. Coenzymes and metals
- 4. Vitamins are converted into coenzymes.
- 5. Enzymes facilitate the formation of the transition state.

6. The intricate three-dimensional structure of proteins allows the construction of active sites that will recognize only specific substrates.

7. The energy required to reach the transition state (the activation energy) is returned when the transition state proceeds to product

8. Protein hydrolysis has a large activation energy. Protein synthesis must require energy to proceed.

9. The enzymes help protect the fluid that surrounds eyes from bacterial infection.

10. Transition states are very unstable. Consequently, molecules that resemble transition states are themselves likely to be unstable and, hence, difficult to synthesize.

11. (a) 0; (b) 28.53; (c) -22.84; (d) -11.42; (e) 5.69. 12. (a) $\Delta G^{\circ\prime} = -RT \ln K'_{\star}$

12. (a)
$$\Delta G$$
 (1.00 × 1.0⁻³)

+1.8 =
$$-(1.98 \times 10^{-3} \text{ kcal}^{-1} \text{ K}^{-1} \text{ mol}^{-1})$$
 (298 K)
(ln[G1P]/[G6P])
-3.05 = ln [G1P]/[G6P]
+3.05 = ln [G6P]/[G1P]
 $K'_{eq}{}^{-1} = 21$ or $K'_{eq} = 4.8 \times 10^{-2}$

Because
$$[G6P]/[G1P] = 21$$
, there is 1 molecule of G1P for every 21 molecules of G6P. Because we started with 0.1 M, the [G1P] is $1/22(0.1 \text{ M}) = 0.0045 \text{ M}$ and [G6P] must be $21/22(0.1 \text{ M})$ or

) or 0.096 M. Consequently, the reaction does not proceed as written to a significant extent.

(b) Supply G6P at a high rate and remove G1P at a high rate by other reactions. In other words, make sure that the [G6P]/[G1P] is kept large.

13. $K_{\text{eq}} = 19, \Delta G^{\circ\prime} = -7.41 \text{ kJ mol}^{-1} (-1.77 \text{ kcal mol}^{-1})$ 14. The three-dimensional structure of an enzyme is stabi-

lized by interactions with the substrate, reaction intermediates, and products. This stabilization minimizes thermal denaturation.

15. At substrate concentrations near the $K_{\rm M}$, the enzyme displays significant catalysis yet is sensitive to changes in substrate concentration.

16. A + S = 10 $K_{\rm M}$, $V_0 = 0.91 V_{\rm max}$. I + S = 20 $K_{\rm M}$, $V_0 =$ $0.91 V_{\rm max}$.

So any Michaelis-Menten curves showing that the enzyme actually attains V_{max} are pernicious lies.

17. (a) 31.1 μ mol; (b) 0.05 μ mol; (c) 622 s⁻¹, a midrange value for enzymes (see Table 8.5).

18. (a) Yes, $K_{\rm M} = 5.2 \times 10^{-6}$ M; (b) $V_{\rm max} = 6.8 \times 10^{-10}$ mol minute⁻¹; (c) 337 s⁻¹.

19. Penicillinase, like glycopeptide transpeptidase, forms an acylenzyme intermediate with its substrate but transfers the intermediate to water rather than to the terminal glycine residue of the pentaglycine bridge.

20. (a) V_{max} is 9.5 μ mol minute⁻¹. K_{M} is 1.1 \times 10⁻⁵ M, the same as without inhibitor.

(b) Noncompetitive

(c) 2.5×10^{-5} M

(d) $f_{\rm ES} = 0.73$, in the presence or absence of this noncompetitive inhibitor.

21. (a) $V = V_{\text{max}} - (V/[S]) K_{\text{M}}$. (b) Slope = $-K_{\rm M}$, y-intercept = $V_{\rm max}$, x-intercept = $V_{\rm max}/K_{\rm M}$. (c) An Eadie-Hofstee plot



22. The rates of utilization of substrates A and B are given by

$$V_{\rm A} = \left(\frac{k_{\rm cat}}{K_{\rm M}}\right)_{\rm A} [\rm E][\rm A]$$

$$V_{\rm A} = \left(\frac{k_{\rm cat}}{K_{\rm M}}\right)_{\rm B} [\rm E][\rm B]$$

Hence, the ratio of these rates is

and

$$V_{\rm A}/V_{\rm B} = \left(\frac{k_{\rm cat}}{K_{\rm M}}\right)_{\rm B} [{\rm A}] / \left(\frac{k_{\rm cat}}{K_{\rm M}}\right)_{\rm A} [{\rm B}]$$

Thus, an enzyme discriminates between competing substrates on the basis of their values of k_{cat}/K_{M} rather than of K_{M} alone. 23. The mutation slows the reaction by a factor of 100 because the activation free energy is increased by 53.22 kJ mol⁻¹ (12.72 kcal mol^{-1}). Strong binding of the substrate relative to the transition state slows catalysis.

24. 1.1 μ mol minute⁻¹

25. (a) This piece of information is necessary for determining the correct dosage of succinvlcholine to administer.

(b) The duration of the paralysis depends on the ability of the serum cholinesterase to clear the drug. If there were one-eighth the amount of enzyme activity, paralysis could last eight times as long, which is undesirable for two reasons. First, the respirator might break from extended use, which would not be good for the patient on the respirator; second, the doctors might miss their golf game. (c) $K_{\rm M}$ is the concentration needed by the enzyme to reach $^{1}/_{2}V_{\rm max}$. Consequently, for a given concentration of substrate, the reaction catalyzed by the enzyme with the lower $K_{\rm M}$ will have the higher rate. The mutant patient with the higher $K_{\rm M}$ will clear the drug at a much lower rate.

26. If the total amount of enzyme (E_T) is increased, V_{max} will increase, because $V_{\text{max}} = k_2[E_T]$. But $K_M = (k_{-1} + k_2)/k_1$; that is, it is independent of substrate concentration. The middle graph describes this situation. 27. (a)



(b) This behavior is substrate inhibition: at high concentrations, the substrate forms unproductive complexes at the active site. The adjoining drawing shows what might happen. Substrate normally binds in a defined orientation, shown in the drawing

as red to red and blue to blue. At high concentrations, the substrate may bind at the active site such that the proper orientation is met for each end of the molecule, but two different substrate molecules are binding.



28. The first step will be the rate-limiting step. Enzymes E_B and E_C are operating at $\frac{1}{2} V_{max}$, whereas the K_M for enzyme E_A is greater than the substrate concentration. E_A would be operating at approximately $10^{-2}V_{max}$.

29. The fluorescence spectroscopy reveals the existence of an enzyme–serine complex and of an enzyme–serine–indole complex. 30. (a) When [S⁺] is much greater than the value of $K_{\rm M}$, pH will have a negligible effect on the enzyme because S⁺ will interact with E⁻ as soon as the enzyme becomes available.



(b) When [S⁺] is much less than the value of $K_{\rm M}$, the plot of V_0 versus pH becomes essentially a titration curve for the ionizable groups, with enzyme activity being the titration marker. At low pH, the high concentration of H⁺ will keep the enzyme in the EH form and inactive. As the pH rises, more and more of the enzyme will be in the E⁻ form and active. At high pH (low H⁺), all of the enzyme is E⁻.



(c) The midpoint on this curve will be the pK_a of the ionizable group, which is stated to be pH 6.

31. (a) Incubating the enzyme at 37° C leads to a denaturation of enzyme structure and a loss of activity. For this reason, most enzymes must be kept cool if they are not actively catalyzing their reactions.

(b) The coenzyme apparently helps to stabilize enzyme structure, because enzyme from PLP-deficient cells denatures faster. Cofactors often help stabilize enzyme structure.

Chapter 9

1. For the amide substrate, the formation of the acyl-enzyme intermediate is slower than the hydrolysis of the acyl-enzyme intermediate, and so no burst is observed. A burst is observed for ester substrates; the formation of the acyl-enzyme intermediate is faster, leading to the observed burst.

2. The histidine residue in the substrate can substitute to some extent for the missing histidine residue of the catalytic triad of the mutant enzyme.

3. No. The catalytic triad works as a unit. After this unit has been made ineffective by the mutation of histidine to alanine, the further mutation of serine to alanine should have only a small effect.

4. The substitution corresponds to one of the key differences between trypsin and chymotrypsin, and so trypsinlike specificity (cleavage after lysine and arginine) might be predicted. In fact, additional changes are required to effect this specificity change.

5. Imidazole is apparently small enough to reach the active site of carbonic anhydrase. Buffers with large molecular components cannot do so, and the effects of the mutation are more evident. 6. No. The odds of such a sequence being present are approximately 1 in $4^{10} = 1,048,576$. Because a typical viral genome has only 50,000 bp, the target sequence would be unlikely to be present. 7. No, because the enzyme would destroy the host DNA before protective methylation could take place.

8. No. The bacteria receiving the enzyme would have their own DNA destroyed because they would likely lack the appropriate protective methylase.

9. EDTA will bind to Zn^{2+} and remove the ion, which is required for enzyme activity, from the enzyme.

10. (a) The aldehyde reacts with the active-site serine. (b) A hemiacetal is formed.

11. Trypsin

12. The reaction is expected to be slower by a factor of 10 because the rate depends on the p K_a of the zinc-bound water. $k_{cat} = 60,000 \text{ s}^{-1}$.

13. EDTA binds the magnesium necessary for the reaction.

ATP hydrolysis is reversible within the active site. ATP hydrolysis takes place within the active site with the incorporation of ¹⁸O, ATP is re-formed, and the ATP is released back into solution.
 If the aspartate is mutated, the protease is inactive and the

virus will not be viable.

16. Water substitutes for the hydroxyl group of serine 236 in mediating proton transfer from the attacking water and the γ -phosphoryl group.

17. (a) Cysteine protease: The same as Figure 9.8, except that cysteine replaces serine in the active site and no aspartate is present.(b) Aspartyl protease:



(c) Metalloprotease:



Chapter 10

1. The enzyme catalyzes the first step in the synthesis of pyrimidines. It facilitates the condensation of carbamoyl phosphate and aspartate to form N-carbamoylaspartate and inorganic phosphate. 2. The protonated form of histidine probably stabilizes the negatively charged carbonyl oxygen atom of the scissile bond in the transition state. Deprotonation would lead to a loss of activity. Hence, the rate is expected to be half maximal at a pH of about 6.5 (the pK of an unperturbed histidine side chain in a protein) and to decrease as the pH is raised.

3. The inhibition of an allosteric enzyme by the end product of the pathway controlled by the enzyme. It prevents the production of too much end product and the consumption of substrates when product is not required.

4. High concentrations of ATP might signal two overlapping situations. The high levels of ATP might suggest that some *nucleotides* are available for nucleic acid synthesis, and consequently, CTP should be synthesized. The high levels of ATP indicate that *energy* is available for nucleic acid synthesis, and so CTP should be produced.

5. All of the enzyme would be in the R form all of the time. There would be no cooperativity. The kinetics would look like that of a Michaelis–Menten enzyme.

6. The enzyme would show simple Michaelis–Menten kinetics because it is essentially always in the R state.

7. CTP is formed by the addition of an amino group to UTP. Evidence indicates the UTP is also capable of inhibiting ATCase. 8. Homotropic effectors are the substrates of allosteric enzymes. Heterotropic effectors are the regulators of allosteric enzymes. Homotropic effectors account for the sigmoidal nature of the velocity versus substrate concentration curve, whereas heterotropic effectors alter the midpoint of $K_{\rm M}$ of the curve. Ultimately, both types of effectors work by altering the T/R ratio.

 9. The reconstitution shows that the complex quaternary structure and the resulting catalytic and regulatory properties are ultimately encoded in the primary structure of individual components.
 10. If substrates had been used, the enzyme would catalyze the reaction. Intermediates would not accumulate on the enzyme. Consequently, any enzyme that crystallized would have been free of substrates or products.

11. (a) 100. The change in the [R]/[T] ratio on binding one substrate molecule must be the same as the ratio of the substrate affinities of the two forms.

(b) 10. The binding of four substrate molecules changes the [R]/[T] by a factor of $100^4 = 10^8$. The ratio in the absence of substrate is 10^{-7} . Hence, the ratio in the fully liganded molecule is $10^8 \times 10^{-7} = 10$. 12. The fraction of molecules in the R form is 10^{-5} , 0.004, 0.615, 0.998, and 1 when 0, 1, 2, 3, and 4 ligands, respectively, are bound. 13. The sequential model can account for negative cooperativity, whereas the concerted model cannot.

14. The binding of PALA switches ATCase from the T to the R state because PALA acts as a substrate analog. An enzyme

molecule containing bound PALA has fewer free catalytic sites than does an unoccupied enzyme molecule. However, the PALA-containing enzyme will be in the R state and, hence, have a higher affinity for the substrates. The dependence of the degree of activation on the concentration of PALA is a complex function of the allosteric constant L_0 and of the binding affinities of the R and T states for the analog and substrates.

15. The net outcome of the two reactions is the hydrolysis of ATP to ADP and P_i, which has a ΔG of -50 kJ mol⁻¹ (-12 kcal mol⁻¹) under cellular conditions.

16. Isozymes are homologous enzymes that catalyze the same reaction but have different kinetic or regulatory properties.

17. Although the same reaction may be required in a variety of different tissues, the biochemical properties of tissues will differ according to their biological function. Isozymes allow the fine-tuning of catalytic and regulatory properties to meet the specific needs of the tissue. 18. (a) 7; (b) 8; (c) 11; (d) 6; (e) 1; (f) 12; (g) 3; (h) 4; (i) 5; (j) 2;

(k) 10; (l) 9.19. When phosphorylation takes place at the expense of ATP, sufficient energy is expended to dramatically alter the structure and hence activity of a protein. Moreover, because ATP is the cellular energy

activity of a protein. Moreover, because A I P is the cellular energy currency, protein modification is linked to the energy status of the cell. 20. Covalent modification is reversible, whereas proteolytic cleavage is irreversible.

21. Activation is independent of zymogen concentration because the reaction is intramolecular.

22. Although quite rare, cases of enteropeptidase deficiency have been reported. The affected person has diarrhea and fails to thrive because digestion is inadequate. In particular, protein digestion is impaired.

23. Add blood from the second patient to a sample from the first. If the mixture clots, the second patient has a defect different from that of the first. This type of assay is called a complementation test. 24. Activated factor X remains bound to blood-platelet membranes, which accelerates its activation of prothrombin.

25. Antithrombin III is a very slowly hydrolyzed substrate of thrombin. Hence, its interaction with thrombin requires a fully formed active site on the enzyme.

26. Residues *a* and *d* are located in the interior of an α -helical coiled coil, near the axis of the superhelix. Hydrophobic interactions between these side chains contribute to the stability of the coiled coil. 27. Leucine would be a good choice. It is resistant to oxidation and has nearly the same volume and degree of hydrophobicity as methionine has.

28. Inappropriate clot formation could block arteries in the brain, causing a stroke, or the heart, causing a heart attack.

29. Tissue-type plasminogen activator, or TPA, is a serine protease that leads to the dissolution of blood clots. TPA activates plasminogen that is bound to a fibrin clot, converting it into active plasmin, which then hydrolyzes the fibrin of the clot.

30. A mature clot is stabilized by amide linkages between the side chains of lysine and glutamine that are absent in a soft clot. The linkages are formed by transglutaminase.

31. The simple sequential model predicts that the fraction of catalytic chains in the R state, $f_{\rm R}$, is equal to the fraction containing bound substrate, Y. The concerted model, in contrast, predicts that $f_{\rm R}$ increases more rapidly than Y as the substrate concentration is increased. The change in $f_{\rm R}$ leads to the change in Y on addition of substrate, as predicted by the concerted model.

32. The binding of succinate to the functional catalytic sites of the native c_3 moiety changed the visible absorption spectrum of nitrotyrosine residues in the *other* c_3 moiety of the hybrid enzyme. Thus, the binding of substrate analog to the active sites of one trimer altered the structure of the other trimer.

33. According to the concerted model, an allosteric activator shifts the conformational equilibrium of all subunits toward the R state, whereas an allosteric inhibitor shifts it toward the T state. Thus, ATP (an allosteric activator) shifted the equilibrium to the R form, resulting in an absorption change similar to that obtained when substrate is bound. CTP had a different effect. Hence, this allosteric inhibitor shifted the equilibrium to the T form. Thus, the concerted model accounts for the ATP-induced and CTP-induced (heterotropic), as well as for the substrate-induced (homotropic), allosteric interactions of ATCase.

34. In the R state, ATCase expands and becomes less dense. This decrease in density results in a decrease in the sedimentation value (see the formula on p. 76).

35. The interaction between trypsin and the inhibitor is so stable that the transition state is rarely formed. Recall that maximal binding energy is released when an enzyme binds to the transition state. If the substrate-enzyme interaction is too stable, the transition state rarely forms.

36.











 2^{-} 0 0 $-^{-}$ 0 $-^{-}$ 0 $-^{-$

Chapter 11

1. Carbohydrates were originally regarded as *hydrates* of *carbon* because the empirical formula of many of them is $(CH_2O)_n$.

2. Three amino acids can be linked by peptide bonds in only six different ways. However, three different monosaccharides can be linked in a plethora of ways. The monosaccharides can be linked in a linear or branched manner, with α or β linkages, with bonds between C-1 and C-3, between C-1 and C-4, between C-1 and C-6, and so forth. In fact, the three monosaccharides can form 12.288 different trisaccharides.

3. (a) aldose-ketose; (b) epimers; (c) aldose-ketose (d) anomers; (e) aldose-ketose; (f) epimers.

4. Erythrose: tetrose aldose; Ribose: pentose aldose; Glyceraldehyde: triose aldose; Dihydroxyacetone: triose ketose; Erythrulose: tetrose ketose; Ribulose: pentose ketose; Fructose: hexose ketose.



6. The proportion of the α anomer is 0.36, and that of the β anomer is 0.64.

7. Glucose is reactive because of the presence of an aldehyde group in its open-chain form. The aldehyde group slowly condenses with amino groups to form aldimine products of a type called Schiff-base adducts.

8. A pyranoside reacts with two molecules of periodate; formate is one of the products. A furanoside reacts with only one molecule of periodate; formate is not formed.

9. From methanol

10 (a) β -D-Mannose; (b) β -D-galactose; (c) β -D-fructose;

(d) β -D-glucosamine.

11. The trisaccharide itself should be a competitive inhibitor of cell adhesion if the trisaccharide unit of the glycoprotein is critical for the interaction.

12. Reducing ends would form 1,2,3,6-tetramethylglucose. The branch points would yield 2,3-dimethylglucose. The remainder of the molecule would yield 2,3,6-trimethylglucose.

13. (a) not a reducing sugar; no open-chain forms are possible. (b) D-Galactose, D-glucose, D-fructose. (c) D-Galactose and sucrose (glucose + fructose).



The hemiketal linkage of the α anomer is broken to form the open form. Rotation about the C-1 and C-2 bonds allows the formation of the β anomer, and a mixture of isomers results. 15. Heating converts the very sweet pyranose form into the more-stable but less-sweet

furanose form. Consequently, the sweetness of the preparation is difficult to accurately control, which also accounts for why honey loses sweetness with time. See Figure 11.5 for structures. 16. (a) Each glycogen molecule has one reducing end, whereas the number of nonreducing ends is determined by the number of branches, or α -1,6 linkages. (b) Because the number of nonreducing ends greatly exceeds the number of reducing ends in a collection of glycogen molecules, all of the degradation and synthesis of glycogen takes place at the nonreducing ends, thus maximizing the rate of degradation and synthesis.

17. No, sucrose is not a reducing sugar. The anomeric carbon atom acts as the reducing agent in both glucose and fructose but, in sucrose, the anomeric carbon atoms of fructose and glucose are joined by a covalent bond and are thus not available to react. 18. Glycogen is polymer of glucose linked by α -1,4-glycosidic bonds with branches formed approximately every 10 glucose units by α -1,6-glycosidic bonds. Starch consists of two polymers of glucose. Amylose is a straight-chain polymer formed by α -1,4-glycosidic bonds. Amylopectin is similar to glycogen but amylopectin has fewer branches, one branch per 30 or so glucose units.

19. Cellulose is a linear polymer of glucose joined by β -1,4 linkages. Glycogen is a branched polymer with the main chain being formed by α -1,4-glycosidic bonds. The β -1,4 linkages allow the formation of a linear polymer ideal for structural roles. The α -1,4 linkages of glycogen form a helical structure, which allows the storage of many glucose moieties in a small space.

20. Simple glycoproteins are often secreted proteins and thus play a variety of roles. For example, the hormone EPO is a glycoprotein. Usually, the protein component constitutes the bulk of the glycoprotein by mass. In contrast, proteoglycans and mucoproteins are predominantly carbohydrates. Proteoglycans have glycosaminoglycans attached, and play structural roles as in cartilage and the extracellular matrix. Mucoproteins often serve as lubricants and have multiple carbohydrates attached through an *N*-acetylgalactosamine moiety. 21. The attachment of the carbohydrate allows the EPO to stay in circulation longer and thus to function for longer periods of time than would a carbohydrate-free EPO.

22. The glycosaminoglycan, because it is heavily charged, binds many water molecules. When cartilage is stressed, such as when your heel hits the ground, the water is released, thus cushioning the impact. When you lift your heel, the water rebinds.

23. The lectin that binds the mannose 6-phosphate might be defective and not recognize a correctly addressed protein.

24. Different molecular forms of a glycoprotein that differ in the amount of carbohydrate attached or the location of attachment or both.

25. The total collection of carbohydrates synthesized by a cell at particular times and under particular environmental conditions.

26. The genome comprises all of the genes present in an organism. The proteome includes all of the possible protein products and modified proteins that a cell expresses under any particular set of circumstances. The glycome consists of all of the carbohydrates synthesized by the cell under any particular set of circumstances. Because the genome is static, but any given protein can be variously expressed and modified, the proteome is more complex than the genome. The glycome, which includes not only glycoforms of proteins, but also many possible carbohydrate structures, must be even more complex. 27. It suggests that carbohydrates are on the cell surfaces of all

organisms for the purpose of recognition by other organisms or the environment.

28. A glycoprotein is a protein that is decorated with carbohydrates. A lectin is a protein that specifically recognizes carbohydrates. A lectin can also be a glycoprotein.

29. Each site either is or is not glycosylated, and so there are $2^6 = 64$ possible proteins.

30. As discussed in Chapter 9, many enzymes display stereochemical specificity. Clearly, the enzymes of sucrose synthesis are able to distinguish between the isomers of the substrates and link only the correct pair.

31. If the carbohydrate specificity of the lectin is known, an affinity column with the appropriate carbohydrate attached could be prepared. The protein preparation containing the lectin of interest could be passed over the column. The use of this method was indeed how the glucose-binding lectin concanavalin A was purified.

32. (a) Aggrecan is heavily decorated with glycosaminoglycans. If glycosaminoglycans are released into the media, aggrecan must be undergoing degradation.

(b) Another enzyme might be present that cleaves glycosaminoglycans from aggrecan without degrading aggrecan. Other experiments not shown established that glycosaminoglycan release is an accurate measure of aggrecan destruction.

(c) The control provides a baseline of "background" degradation inherent in the assay.

(d) Aggrecan degradation is greatly enhanced.

(e) Aggrecan degradation is reduced to the background system.

(f) It is an in vitro system in which not all the factors contributing to cartilage stabilization in vivo are present.

Chapter 12

1. 2.86×10^6 molecules, because each leaflet of the bilayer contains 1.43×10^6 molecules.

2. Essentially an "inside-out" membrane. The hydrophilic groups would come together on the interior of the structure, away from the solvent, whereas the hydrocarbon chains would interact with the solvent.

3. 2×10^{-7} cm, 6×10^{-6} cm, and 2×10^{-4} cm.

4. The radius of this molecule is 3.1×10^{-7} cm, and its diffusion coefficient is 7.4×10^{-9} cm² s⁻¹. The average distances traversed are 1.7×10^{-7} cm in 1 µs, 5.4×10^{-6} cm in 1 ms, and 1.7×10^{-4} cm in 1 s.

5. The membrane underwent a phase transition from a highly fluid to a nearly frozen state when the temperature was lowered. A carrier can shuttle ions across a membrane only when the bilayer is highly fluid. A channel, in contrast, allows ions to traverse its pore even when the bilayer is quite rigid.

6. The presence of a cis double bond introduces a kink in the fatty acid chain that prevents tight packing and reduces the number of atoms in van der Waals contact. The kink lowers the melting point compared with that of a saturated fatty acid. Trans fatty acids do not have the kink, and so their melting temperatures are higher, more similar to those of saturated fatty acids.

Because trans fatty acids have no structural effect, they are rarely observed.

7. Palmitic acid is shorter than stearic acid. Thus, when the chains pack together, there is less opportunity for van der Waals interaction and the melting point is thus lower than that of the longer stearic acid. 8. Hibernators selectively feed on plants that have a high proportion of polyunsaturated fatty acids with lower melting temperature. 9. The initial decrease in fluorescence with the first addition of sodium dithionite results from the quenching of NBD-PS molecules in the outer leaflet of the bilayer. Sodium dithionite does not traverse the membrane under these experimental conditions; hence, it does not quench the labeled phospholipids in the inner leaflet. A second addition of sodium dithionite has no effect, as the NBD-PS molecules in the outer leaflet remain quenched. However, after a 6.5 hour incubation, about half the NBD-PS has flipped over to the outer leaflet of the bilayer, resulting in the 50% decrease in fluorescence when sodium dithionite is added.

10. The addition of the carbohydrate introduces a significant energy barrier to the flip-flop because a hydrophilic carbohydrate moiety would need to be moved through a hydrophobic environment. This energetic barrier enhances membrane asymmetry.

11. The C_{16} alkyl chain is attached by an ether linkage. The C-2 carbon atom of glycerol has only an acetyl group attached by an ester linkage instead of a fatty acid, as is the case with most phospholipids.

12. In a hydrophobic environment, the formation of intrachain hydrogen bonds stabilizes the amide hydrogen atoms and carbonyl oxygen atoms of the polypeptide chain, and so an α helix forms. In an aqueous environment, these groups are stabilized by interaction with water, and so there is no energetic reason to form an α helix. Thus, the α helix would be more likely to form in a hydrophobic environment.

13. The protein may contain an α helix that passes through the hydrophobic core of the protein. This helix is likely to feature a stretch of hydrophobic amino acids similar to those observed in transmembrane helices.

14. The shift to the lower temperature would decrease fluidity by enhancing the packing of the hydrophobic chains by van der Waals interactions. To prevent this packing, new phospholipids having shorter chains and a greater number of cis double bonds would be synthesized. The shorter chains would reduce the number of van der Waals interactions, and the cis double bonds, which cause the kink in structure, would prevent the packing of the fatty acid tails of the phospholipids.

15. Each of the 21 v-SNARE proteins could interact with each of 7 t-SNARE partners. Multiplication gives the total number of different interacting pairs: $7 \times 21 = 147$ different v-SNARE-t-SNARE pairs.

16. (a) The graph shows that, as temperature increases, the phospholipid bilayer becomes more fluid. $T_{\rm m}$ is the temperature of the transition from the predominantly less fluid state to the predominantly more fluid state. Cholesterol broadens the transition from the less-fluid to the more-fluid state. In essence, cholesterol makes membrane fluidity less sensitive to temperature changes.

(b) This effect is important because the presence of cholesterol tends to stabilize membrane fluidity by preventing sharp transitions. Because protein function depends on the proper fluidity of the membrane, cholesterol maintains the proper environment for membrane-protein function.

17. The protein plotted in part c is a transmembrane protein from *C. elegans*. It spans the membrane with four α helices that are prominently displayed as hydrophobic peaks in the hydropathy

plot. Interestingly, the protein plotted in part *a* also is a membrane protein, a porin. This protein is made primarily of β strands, which lack the prominent hydrophobic window of membrane helices. This example shows that, although hydropathy plots are useful, they are not infallible.

18. To purify any protein, the protein must first be solubilized. For a membrane protein, solubilization usually requires a detergent hydrophobic molecules that bind to the protein and thus replace the lipid environment of the membrane. If the detergent is removed, the protein aggregates and precipitates from solution. Often, the steps in purification, such as ion-exchange chromatography, are difficult to perform in the presence of sufficient detergent to solubilize the protein. Crystals of appropriate protein–detergent complexes must be generated.

Chapter 13

1. In simple diffusion, the substance in question can diffuse down its concentration gradient through the membrane. In facilitated diffusion, the substance is not lipophilic and cannot directly diffuse through the membrane. A channel or carrier is required to facilitate movement down the gradient.

 The two forms are (1) ATP hydrolysis and (2) the movement of one molecule down its concentration gradient coupled with the movement of another molecule up its concentration gradient.
 The three types of carriers are symporters, antiporters, and uniporters. Symporters and antiporters can mediate secondary active transport.

4. The free-energy cost is 32 kJ mol^{-1} (7.6 kcal mol⁻¹). The chemical work performed is 20.4 kJ mol^{-1} (4.9 kcal mol⁻¹), and the electrical work performed is 11.5 kJ mol^{-1} (2.8 kcal mol⁻¹). 5. For chloride, z = -1; for calcium z = +2. At the concentrations given, the equilibrium potential for chloride is -97 mV and the equilibrium potential for calcium is +122 mV.

6. The concentration of glucose inside the cell is 66 times as great as that outside the cell $[(c_2/c_1) = 66]$ when the free-energy input is 10.8 kJ mol⁻¹ (2.6 kcal mol⁻¹).

7. By analogy with the Ca^{2+} ATPase, with three Na⁺ ions binding from inside the cell to the E_1 conformation and with two K⁺ ions binding from outside the cell to the E_2 conformation, a plausible mechanism is as follows:

(i) A catalytic cycle could begin with the enzyme in its unphosphorylated state (E_1) with three sodium ions bound.

(ii) The E_1 conformation binds ATP. A conformational change traps sodium ions inside the enzyme.

(iii) The phosphoryl group is transferred from ATP to an aspartyl residue.

(iv) On ADP release, the enzyme changes its overall conformation, including the membrane domain. This new conformation (E₂) releases the sodium ions to the side of the membrane opposite that at which they entered and binds two potassium ions from the side where sodium ions are released.

(v) The phosphorylaspartate residue is hydrolyzed to release inorganic phosphate. With the release of phosphate, the interactions stabilizing E_2 are lost, and the enzyme everts to the E_1 conformation. Potassium ions are released to the cytoplasmic side of the membrane. The binding of three sodium ions from the cytoplasmic side of the membrane completes the cycle.

8. Establish a lactose gradient across vesicle membranes that contain properly oriented lactose permease. Initially, the pH should be the same on both sides of the membrane and the lactose concentration should be higher on the "exit" side of lactose

permease. As the lactose flows "in reverse" through the permease, down its concentration gradient, it can be tested whether or not a pH gradient becomes established as the lactose gradient is dissipated.

9. Ligand-gated channels open in response to the binding of a molecule by the channel, whereas voltage-gated channels open in response to changes in the membrane potential.

10. An ion channel must transport ions in either direction at the same rate. The net flow of ions is determined only by the composition of the solutions on either side of the membrane.

11. Uniporters act as enzymes do; their transport cycles include large conformational changes, and only a few molecules interact with the protein per transport cycle. In contrast, channels, after having opened, provide a pore in the membrane through which many ions may pass. As such, channels mediate transport at a much higher rate than do uniporters.

12. FCCP effectively creates a pore in the bacterial membrane through which protons can pass rapidly. Protons that are pumped out of the bacteria will pass through this pore preferentially (the "path of least resistance"), rather than participate in H^+ /lactose symport.

13. Cardiac muscle must contract in a highly coordinated manner in order to pump blood effectively. Gap junctions mediate the orderly cell-to-cell propagation of the action potential through the heart during each beat.

14. The positively charged guanidinium group resembles Na⁺ and binds to negatively charged carboxylate groups in the mouth of the channel.

15. SERCA, a P-type ATPase, uses a mechanism by which a covalent phosphorylated intermediate (at an aspartate residue) is formed. At steady state, a subset of the SERCA molecules are trapped in the E_2 -P state and, as a result, radiolabeled. The MDR protein is an ABC transporter and does not operate through a phosphorylated intermediate. Hence, a radiolabeled band would not be observed for MDR.

16. The blockage of ion channels inhibits action potentials, leading to loss of nervous function. Like tetrodotoxin, these toxin molecules are useful for isolating and specifically inhibiting particular ion channels.

 After repolarization, the ball domains of the ion channels engage the channel pore, rendering them inactive for a short period of time. During this time, the channels cannot be reopened until the ball domains disengage and the channel returns to the "closed" state.
 Because sodium ions are charged and because sodium channels carry only sodium ions (but not anions), the accumulation of excess positive charge on one side of the membrane dominates the chemical gradients.

19. A mutation that impairs the ability of the sodium channel to inactivate would prolong the duration of the depolarizing sodium current, thus lengthening the cardiac action potential.

20. No. Channels will likely open or close in response to an external stimulus, but the unit conductance of the open channel will be influenced very little.

21. The ratio of closed to open forms of the channel is 10^5 , 5000, 250, 12.5, and 0.625 when zero, one, two, three, and four ligands, respectively, are bound. Hence, the fraction of open channels is 1.0 $\times 10^{-5}$, 2.0 $\times 10^{-4}$, 4.0 $\times 10^{-3}$, 7.4 $\times 10^{-2}$, and 0.62.

22. These organic phosphates inhibit acetylcholinesterase by reacting with the active-site serine residue to form a stable phosphorylated derivative. They cause respiratory paralysis by blocking synaptic transmission at cholinergic synapses.

23. (a) The binding of the first acetylcholine molecule increases the open-to-closed ratio by a factor of 240, and the binding of

the second increases it by a factor of 11,700. (b) The free-energy contributions are 14 kJ mol⁻¹ (3.3 kcal mol⁻¹) and 23 kJ mol⁻¹ (5.6 kcal mol⁻¹), respectively. (c) No; the MWC model predicts that the binding of each ligand will have the same effect on the open-to-closed ratio.

24. Batrachotoxin blocks the transition from the open to the closed state.

25. (a) Chloride ions flow into the cell. (b) Chloride flux is inhibitory because it hyperpolarizes the membrane. (c) The channel consists of five subunits.

26. After the addition of ATP and calcium, SERCA will pump Ca^{2+} ions into the vesicle. However, the accumulation of Ca^{2+} ions inside the vesicle will rapidly lead to the formation of an electrical gradient that cannot be overcome by ATP hydrolysis. The addition of calcimycin will allow the pumped Ca^{2+} ions to flow back out of the vesicle, dissipating the charge buildup, and enabling the pump to operate continuously.

27. The catalytic prowess of acetylcholinesterase ensures that the duration of the nerve stimulus will be short.

28. See reaction below.



29. (a) Only ASIC1a is inhibited by the toxin. (b) Yes; when the toxin was removed, the activity of the acid-sensing channel began to be restored. (c) 0.9 nM.

30. This mutation is one of a class of mutations that result in slow-channel syndrome (SCS). The results suggest a defect in channel closing; so the channel remains open for prolonged periods. Alternatively, the channel may have a higher affinity for acetyl-choline than does the control channel.

31. The mutation reduces the affinity of acetylcholine for the receptor. The recordings would show the channel opening only infrequently.

32. Glucose displays a transport curve that suggests the participation of a carrier because the initial rate is high but then levels off at higher concentrations, consistent with saturation of the carrier, which is reminiscent of Michaelis–Menten enzymes (Section 8.4). Indole shows no such saturation phenomenon, which implies that the molecule is lipophilic and simply diffuses across the membrane. Ouabain is a specific inhibitor the Na^+-K^+ pump. If ouabain were to inhibit glucose transport, then a Na^+ -glucose cotransporter would be assisting in transport.

Chapter 14

1. The negatively charged glutamate residues mimic the negatively charged phosphoserine or phosphothreonine residues and stabilize the active conformation of the enzyme.

2. No. Phosphoserine and phosphothreonine are considerably shorter than phosphotyrosine.

3. The GTPase activity terminates the signal. Without such activity, after a pathway has been activated, it remains activated and is unresponsive to changes in the initial signal. If the GTPase activity were more efficient, the lifetime of the GTP-bound G_{α} subunit would be too short to achieve downstream signaling.

4. Two identical receptor molecules must recognize different aspects of the same signal molecule.

5. Growth-factor receptors can be activated by dimerization. If an antibody causes a receptor to dimerize, the signal-transduction pathway in a cell will be activated.

6. The mutated α subunit will always be in the GTP form and, hence, in the active form, which would stimulate its signaling pathway.

7. A G protein is a component of the signal-transduction pathway. GTP γ S is not hydrolyzed by the G_{α} subunit, leading to prolonged activation.

 Calcium ions diffuse slowly because they bind to many protein surfaces within a cell, impeding their free motion. Cyclic AMP does not bind as frequently, and so it diffuses more rapidly.
 Fura-2 is a highly negatively charged molecule, with five carboxylate groups. Its charge prevents it from effectively crossing the hydrophobic region of the plasma membrane.

10. $G_{\alpha s}$ stimulates adenylate cyclase, leading to the generation of cAMP. This signal then leads to glucose mobilization (see Chapter 21). If cAMP phosphodiesterase were inhibited, then cAMP levels would remain high even after the termination of the epinephrine signal, and glucose mobilization would continue. 11. If the two kinase domains are forced to be within close proximity of each other, the activation loop of one kinase, in its inactivating conformation, can be displaced by the activation loop of the neighboring kinase, which acts as a substrate for phosphorylation. 12. The full network of pathways initiated by insulin includes a large number of proteins and is substantially more elaborate than indicated in Figure 14.25. Furthermore, many additional proteins take part in the termination of insulin signaling. A defect in any of the proteins in the insulin signaling pathways or in the subsequent termination of the insulin response could potentially cause problems. Therefore, it is not surprising that many different gene defects can cause type 2 diabetes.

13. The binding of growth hormone causes its monomeric receptor to dimerize. The dimeric receptor can then activate a separate tyrosine kinase to which the receptor binds. The signaling pathway can then continue in similar fashion to the pathways that are activated by the insulin receptor or other mammalian EGF receptors.

14. The truncated receptor will dimerize with the full-length monomers on EGF-binding, but cross-phosphorylation cannot take place, because the truncated receptor possesses neither the substrate for the neighboring kinase domain nor its own kinase domain to

phosphorylate the C-terminal tail of the other monomer. Hence, these mutant receptors will block normal EGF signaling. 15. Insulin would elicit the response that is normally caused by EGF. Insulin binding will likely stimulate dimerization and phosphorylation of the chimeric receptor and thereby signal the downstream events that are normally triggered by EGF binding. Exposure of these cells to EGF would have no effect. 16. 10^5

17. The formation of diacylglycerol implies the participation of phospholipase C. A simple pathway would entail receptor activation by cross-phosphorylation, followed by the binding of phospholipase C γ (through its SH2 domains). The participation of phospholipase C indicates that IP₃ would be formed and, hence, calcium concentrations would increase.

18. Other potential drug targets within the EGF signaling cascade include, but are not limited to, the kinase active sites of the EGF receptor, Raf, MEK, or ERK.

19. In the reaction catalyzed by adenylate cyclase, the 3'-OH group nucleophilically attacks the α -phosphorus atom attached to the 5'-OH group, leading to displacement of pyrophosphate. The reaction catalyzed by DNA polymerase is similar except that the 3'-OH group is on a different nucleotide.

20. ATP-competitive inhibitors are likely to act on multiple kinases because every kinase domain contains an ATP-binding site. Hence, these drugs may not be selective for the desired kinase target.

21. (a) $X \approx 10^{-7}$ M; $Y \approx 5 \times 10^{-6}$ M; $Z \approx 10^{-3}$ M. (b) Because much less X is required to fill half of the sites, X displays the highest affinity. (c) The binding affinity almost perfectly matches the ability to stimulate adenylate cyclase, suggesting that the hormone–receptor complex leads to the stimulation of adenylate cyclase. (d) Try performing the experiment in the presence of antibodies to $G_{\alpha s}$. 22. (a) The total binding does not distinguish binding to a specific

receptor from binding to different receptors or from nonspecific binding to the membrane.

(b) The rationale is that the receptor will have a high affinity for the ligand. Thus, in the presence of excess nonradioactive ligand, the receptor will bind to nonradioactive ligand. Therefore, any binding of the radioactive ligand must be nonspecific.

(c) The plateau suggests that the number of receptor-binding sites in the cell membrane is limited.

23. Number of receptors per cell =

$$\frac{10^{4} \text{ cpm}}{\text{mg of membrane protein}} \times \frac{\text{mg of membrane protein}}{10^{10} \text{ cells}} \times \frac{\text{mmol}}{10^{12} \text{ cpm}} \times \frac{6.023 \times 10^{20} \text{ molecules}}{\text{mmol}} = 600$$

Chapter 15

1. The highly integrated biochemical reactions that take place inside the cell.

2. Anabolism is the set of biochemical reactions that use energy to build new molecules and ultimately new cells. Catabolism is the set of biochemical reactions that extract energy from fuel sources or breakdown biomolecules.

3. Cellular movements and the performance of mechanical work; active transport; biosynthetic reactions.

4. 1. f; 2. h; 3. i; 4. a; 5. g; 6. b; 7. c; 8. e; 9. j; 10. d.

5. Charge repulsion, resonance stabilization, and stabilization by hydration.

6. Trick question. The answer is not known. Adenine appears to form more readily under prebiotic conditions; so ATP may have predominated initially.

7. Having only one nucleotide represent the available energy allows the cell to better monitor its energy status.

8. Increasing the concentration of ATP or decreasing the concentration cellular ADP or P_i (by rapid removal by other reactions, for instance) would make the reaction more exergonic. Likewise, altering the Mg^{2+} concentration could raise or lower the ΔG of the reaction.

9. The free-energy changes of the individual steps in a pathway are summed to determine the overall free-energy change of the entire pathway. Consequently, a reaction with a positive freeenergy value can be powered to take place if coupled to a sufficiently exergonic reaction.

10. Reactions in parts a and c, to the left; reactions in parts b and d. to the right.

11. None whatsoever

12. (a) $\Delta G^{\circ} = 31.4 \text{ kJ mol}^{-1} (7.5 \text{ kcal mol}^{-1}) \text{ and } K'_{eq} = 3.06 \times 10^{-6}$; (b) 3.28×10^{4} .

13. $\Delta G^{\circ\prime} = 7.1 \text{ kJ mol}^{-1} (1.7 \text{ kcal mol}^{-1})$. The equilibrium ratio is 17.8.

14. (a) Acetate + CoA + H⁺ goes to acetyl CoA + H₂O, $\Delta G^{\circ \prime}$ = -31.4 kJ mol⁻¹ (-7.5 kcal mol⁻¹). ATP hydrolysis to AMP and PP_i, $\Delta G^{\circ \prime} = -45.6 \text{ kJ mol}^{-1} (-10.9 \text{ kcal mol}^{-1})$. Overall reaction, $\Delta G^{\circ'} = -14.2 \text{ kJ mol}^{-1} (-3.4 \text{ kcal mol}^{-1})$.

(b) With pyrophosphate hydrolysis, $\Delta G^{\circ\prime} = -33.4 \text{ kJ mol}^{-1}$ $(-7.98 \text{ kcal mol}^{-1})$. Pyrophosphate hydrolysis makes the overall reaction even more exergonic.

15. (a) For an acid AH,

$$AH \Longrightarrow A^{-1} + H^{+} \quad K = \frac{[A^{-}][H^{+}]}{[AH]}$$

The pK is defined as $pK = -\log_{10} K$. $\Delta G^{\circ\prime}$ is the standard freeenergy change at pH 7. Thus, $\Delta G^{\circ\prime} = -RT \ln K = -2.303 RT$ log₁₀ K = +2.303 RT pK. (b) $\Delta G^{\circ \prime} = 27.32 \text{ kJ mol}^{-1} (6.53 \text{ kcal mol}^{-1})$.

16. Arginine phosphate in invertebrate muscle, like creatine phosphate in vertebrate muscle, serves as a reservoir of high-potential phosphoryl groups. Arginine phosphate maintains a high level of ATP in muscular exertion.

17. An ADP unit

18. (a) The rationale behind creatine supplementation is that it would be converted into creatine phosphate and thus serve as a rapid means of replenishing ATP after muscle contraction. (b) If creatine supplementation is beneficial, it would affect activities that depend on short bursts of activity; any sustained activity would require ATP generation by fuel metabolism, which, as Figure 15.7 shows, requires more time.

19. Under standard conditions, $\Delta G^{\circ\prime} = -RT \ln [\text{products}]/[\text{reactants}]$. Substituting 23.8 kJ mol⁻¹ (5.7 kcal mol⁻¹) for $\Delta G^{\circ\prime}$ and solving for [products]/[reactants] yields 7 × 10⁻⁵. In other words, the forward reaction does not take place to a significant extent. Under intracellular conditions, ΔG is -1.3 kJ mol⁻ $(-0.3 \text{ kcal mol}^{-1})$. Using the equation $\Delta G = \Delta G^{\circ} + RT \ln I$ [products]/[reactants] and solving for [products]/[reactants] gives a ratio of 3.7×10^{-5} . Thus, a reaction that is endergonic under standard conditions can be converted into an exergonic reaction by maintaining the [products]/[reactants] ratio below the equilibrium value. This conversion is usually attained by using the products in another coupled reaction as soon as they are formed. 20. Under standard conditions,

$$K'_{\rm eq} = \frac{[B]_{\rm eq}}{[A]_{\rm eq}} \times \frac{[ADP]_{\rm eq} [P_{\rm i}]_{\rm eq}}{[ATP]_{\rm eq}} = 10^{3.3/1.36} = 2.67 \times 10^2$$

At equilibrium, the ratio of [B] to [A] is given by

$$\frac{[B]_{eq}}{[A]_{eq}} = K'_{eq} \frac{[ATP]_{eq}}{[ADP]_{eq}[P_i]_{eq}}$$

The ATP-generating system of cells maintains the [ATP]/[ADP] $[P_i]$ ratio at a high level, typically about 500 M⁻¹. For this ratio,

$$\frac{[B]_{eq}}{[A]_{eq}} = 2.67 \times 10^2 \times 500 = 1.34 \times 10^5$$

This equilibrium ratio is strikingly different from the value of 1.15×10^{-3} for the reaction A \rightarrow B in the absence of ATP hydrolysis. In other words, coupling the hydrolysis of ATP with the conversion of A into B has changed the equilibrium ratio of B to A by a factor of about 10^8 .

21. Liver: $-45.2 \text{ kJ mol}^{-1}$ (-10.8 kcal mol⁻¹); muscle: -48.1 kJ mol⁻¹ (-11.5 kcal mol⁻¹); brain: -48.5 kJ mol⁻¹ (-11.6 kcal mol⁻¹). The ΔG is most negative in brain cells.

22. (a) Ethanol; (b) lactate; (c) succinate; (d) isocitrate; (e) malate; (f) 2-phosphoglycerate.

23. Recall that $\Delta G = \Delta G^{\circ'} + RT \ln [\text{products}]/[\text{reactants}]$. Altering the ratio of products to reactants will cause ΔG to vary. In glycolysis, the concentrations of the components of the pathway result in a value of ΔG greater than that of $\Delta G^{\circ'}$.

24. Unless the ingested food is converted into molecules capable of being absorbed by the intestine, no energy can ever be extracted by the body.

25. NADH and FADH₂ are electron carriers for catabolism; NADPH is the carrier for anabolism.

26. The electrons of the C–O bond cannot form resonance structures with the C-S bond that are as stable as those that they can form with the C-O bond. Thus, the thioester is not stabilized by resonance to the same degree as an oxygen ester is stabilized.

27. Oxidation-reduction reactions; ligation reactions; isomerization reactions; group-transfer reactions; hydrolytic reactions; the addition of functional groups to double bonds to form single bonds or the removal of functional groups to form double bonds.

28. Controlling the amount of enzymes; controlling enzyme activity; controlling the availability of substrates.

29. Although the reaction is thermodynamically favorable, the reactants are kinetically stable because of the large activation energy. Enzymes lower the activation energy so that reactions take place on time scales required by the cell.

30. The activated form of sulfate in most organisms is 3'-phosphoadenosine-5'-phosphosulfate.

31. (a) As the Mg²⁺ concentration falls, the ΔG of hydrolysis rises. Note that pMg is a logarithmic plot, and so each number on the *x*-axis represents a 10-fold change in $[Mg^{2+}]$.

(b) Mg²⁺ would bind to the phosphates of ATP and help to mitigate charge repulsion. As the $[Mg^{2+}]$ falls, charge stabilization of ATP would be less, leading to greater charge repulsion and an increase in ΔG on hydrolysis.

Chapter 16

1. Two molecules of ATP are produced per molecule of glyceraldehyde 3-phosphate and, because two molecules of GAP are produced per molecule of glucose, the total ATP yield is four. However, two molecules of ATP are required to convert glucose into fructose 1,6-bisphosphate. Thus, the net yield is only two molecules of ATP.

2. In both cases, the electron donor is glyceraldehyde 3-phosphate. In lactic acid fermentation, the electron acceptor is pyruvate, converting it into lactate. In alcoholic fermentation, acetaldehyde is the electron acceptor, forming ethanol.

3. (a) 3 ATP; (b) 2 ATP; (c) 2 ATP; (d) 2 ATP; (e) 4 ATP.

4. Glucokinase enables the liver to remove glucose from the blood when hexokinase is saturated, ensuring that glucose is captured for later use.

5. Glycolysis is a component of alcoholic fermentation, the pathway that produces alcohol for beer and wine. The belief was that understanding the biochemical basis of alcohol production might lead to a more-efficient means of producing beer.

6. The conversion of glyceraldehyde 3-phosphate into 1,3bisphosphoglycerate would be impaired. Glycolysis would be less effective.

7. Glucose 6-phosphate must have other fates. Indeed, it can be converted into glycogen (Chapter 21) or be processed to yield reducing power for biosynthesis (Chapter 20).

8. The energy needs of a muscle cell vary widely, from rest to intense exercise. Consequently, the regulation of phosphofructo-kinase by energy charge is vital. In other tissues, such as the liver, ATP concentration is less likely to fluctuate and will not be a key regulator of phosphofructokinase.

9. The $\Delta G^{\circ\prime}$ for the reverse of glycolysis is +96 kJ mol⁻¹ (+23 kcal mol⁻¹), far too endergonic to take place.

10. The conversion of glucose into glucose 6-phosphate by

hexokinase; the conversion of fructose 6-phosphate into fructose 1,6-bisphosphate by phosphofructokinase; the formation of pyruvate from phosphoenolpyruvate by pyruvate kinase.

11. Lactic acid is a strong acid. If it remained in the cell, the pH of the cell would fall, which could lead to the denaturation of muscle protein and result in muscle damage.

12. GLUT2 transports glucose only when the blood concentration of glucose is high, which is precisely the condition in which the β cells of the pancreas secrete insulin.

13. Fructose + ATP \longrightarrow fructose 1-phosphate + ADP:

Fructokinase

Fructose 1-phosphate \longrightarrow dihydroxyacetone phosphate + glyceraldehyde: Fructose 1-phosphate aldolase

Glyceraldehyde + ATP \longrightarrow glyceraldehyde 3-phosphate + ADP: Triose kinase

The primary controlling step of glycolysis catalyzed by phosphofructokinase is bypassed by the preceding reactions. Glycolysis will proceed in an unregulated fashion.

14. Without triose isomerase, only one of the two three-carbon molecules generated by aldolase could be used to generate ATP. Only two molecules of ATP would result from the metabolism of each glucose. But two molecules of ATP would still be required to form fructose 1,6-bisphosphate, the substrate for aldolase. The net yield of ATP would be zero, a yield incompatible with life. 15. Glucose is reactive because its open-chain form contains an aldehyde group.

16. (a) The label is in the methyl carbon atom of pyruvate. (b) 5 mCi mM^{-1} . The specific activity is halved because the number of moles of product (pyruvate) is twice that of the labeled substrate (glucose).

17. (a) Glucose + 2
$$P_i$$
 + 2 ADP \rightarrow 2 lactate + 2 ATP.

(b)
$$\Delta G = -114 \text{ kJ mol}^{-1} (-27.2 \text{ kcal mol}^{-1}).$$

18. 3.06×10^{-5}

19. The equilibrium concentrations of fructose 1,6-bisphosphate, dihydroxyacetone phosphate, and glyceraldehyde 3-phosphate

are 7.8×10^{-4} M, 2.2×10^{-4} M, and 2.2×10^{-4} M, respectively.

20. All three carbon atoms of 2,3-BPG are 14 C labeled. The phosphorus atom attached to the C-2 hydroxyl group is 32 P labeled.

21. Hexokinase has a low ATPase activity in the absence of a sugar because it is in a catalytically inactive conformation. The addition of xylose closes the cleft between the two lobes of the enzyme. However, xylose lacks a hydroxymethyl group, and so it cannot be phosphorylated. Instead, a water molecule at the site normally occupied by the C-6 hydroxymethyl group acts as the acceptor of the phosphoryl group from ATP.

22. (a) The fructose 1-phosphate pathway forms glyceraldehyde 3-phosphate.

(b) Phosphofructokinase, a key control enzyme, is bypassed.Furthermore, fructose 1-phosphate stimulates pyruvate kinase.23. The reverse of glycolysis is highly endergonic under cellular conditions. The expenditure of six NTP molecules in gluconeogenesis renders gluconeogenesis exergonic.

24. Lactic acid is capable of being further oxidized and is thus useful energy. The conversion of this acid into glucose saves the carbon atoms for future combustion.

25. In glycolysis, the formation of pyruvate and ATP by pyruvate kinase is irreversible. This step is bypassed by two reactions in gluconeogenesis: (1) the formation of oxaloacetate from pyruvate and CO_2 by pyruvate carboxylase and (2) the formation of phosphoenolpyruvate from oxaloacetate and GTP by phosphoenolpyruvate carboxykinase. The formation of fructose 1,6-bisphosphate by phosphofructokinase is bypassed by fructose 1,6-bisphosphates in gluconeogenesis, which catalyzes the conversion of fructose 1,6-bisphosphate into fructose 6-phosphate. Finally, the hexokinase-catalyzed formation of glucose 6-phosphate in glycolysis is bypassed by glucose 6-phosphatase, but only in the liver.

26. Reciprocal regulation at the key allosteric enzymes in the two pathways. For instance, PFK is stimulated by fructose 2,6-bisphosphate and AMP. The effect of these signals is opposite that of fructose 1,6-bisphosphatase. If both pathways were operating simultaneously, a futile cycle would result. ATP would be hydrolyzed, yielding only heat.

27. Muscle is likely to produce lactic acid during contraction. Lactic acid is a strong acid and cannot accumulate in muscle or blood. Liver removes the lactic acid from the blood and converts it into glucose. The glucose can be released into the blood or stored as glycogen for later use.

28. Glucose produced by the liver could not be released into the blood. Tissues that rely on glucose as an energy source would not function as well unless glucose was provided in the diet.

29. Glucose is an important energy source for both tissues and is essentially the only energy source for the brain. Consequently, these tissues should never release glucose. Glucose release is prevented by the absence of glucose 6-phosphatase.

30. 6 NTP (4 ATP and 2 GTP); 2 NADH.

31. (a) None; (b) none; (c) 4 (2 ATP and 2 GTP); (d) none.

32. If the amino groups are removed from alanine and aspartate, the ketoacids pyruvate and oxaloacetate are formed. Both of these molecules are components of the gluconeogenic pathway.

33. (a) Increased; (b) increased; (c) increased; (d) decreased.

34. Fructose 2,6-bisphosphate, present at high concentration when glucose is abundant, normally inhibits gluconeogenesis by blocking fructose 1,6-bisphosphatase. In this genetic disorder, the phosphatase is active irrespective of the glucose level. Hence, substrate cycling is increased. The level of fructose 1,6-bisphosphate is

consequently lower than normal. Less pyruvate is formed and thus less ATP is generated.

35. Reactions in parts *b* and *e* would be blocked.

36. There will be no labeled carbons. The CO₂ added to pyruvate (formed from the lactate) to form oxaloacetate is lost with the conversion of oxaloacetate into phosphoenolpyruvate.
37. The net reaction in the presence of arsenate is

Glyceraldehyde 3-phosphate + $NAD^+ + H_2O \longrightarrow$ 3-phosphoglycerate + $NADH + 2 H^+$

Glycolysis proceeds in the presence of arsenate, but the ATP normally formed in the conversion of 1,3-bisphosphoglycerate into 3-phosphoglycerate is lost. Thus, arsenate uncouples oxidation and phosphorylation by forming a highly labile acyl arsenate.

38. This example illustrates the difference between the *stoichio-metric* and the *catalytic* use of a molecule. If cells used NAD⁺ stoichiometrically, a new molecule of NAD⁺ would be required each time a molecule of lactate was produced. As we will see, the synthesis of NAD⁺ requires ATP. On the other hand, if the NAD⁺ that is converted into NADH could be recycled and reused, a small amount of the molecule could regenerate a vast amount of lactate, which is the case in the cell. NAD⁺ is regenerated by the oxidation of NADH and reused. NAD⁺ is thus used catalytically.

39. Consider the equilibrium equation of adenylate kinase:

 $K_{eq} = [ATP] [AMP] / [ADP]^{2} (1)$

or

$$AMP = K_{eq} [ADP_2] / [ATP] (2)$$

Recall that [ATP] > [ADP] > [AMP] in the cell. As ATP is utilized, a small decrease in its concentration will result in a larger percentage increase in [ADP] because its concentration is greater than that of ADP. This larger percentage increase in [ADP] will result in an even greater percentage increase in [AMP] because the concentration of AMP is related to the square of [ADP]. In essence, equation 2 shows that monitoring the energy status with AMP magnifies small changes in [ATP], leading to tighter control.

40. The synthesis of glucose during intense exercise provides a good example of interorgan cooperation in higher organisms. When muscle is actively contracting, lactate is produced from glucose by glycolysis. The lactate is released into the blood and absorbed by the liver, where it is converted by gluconeogenesis into glucose. The newly synthesized glucose is then released and taken up by the muscle for energy generation.

41. The input of four additional high-phosphoryl-transfer-potential molecules in gluconeogenesis changes the equilibrium constant by a factor of 10^{32} , which makes the conversion of pyruvate into glucose thermodynamically feasible. Without this energetic input, gluconeogenesis would not take place.

42. The mechanism is analogous to that for triose phosphate isomerase (Figure 16.5). It proceeds through an enediol intermediate. The active site would be expected to have a general base (analogous to Glu 165 in TPI) and a general acid (analogous to His 95 in TPI).

43. Galactose is a component of glycoproteins. Possibly, the absence of galactose leads to the improper formation or function of glycoproteins required in the central nervous system. More generally, the fact that the symptoms arise in the absence of galactose suggests that galactose is required in some fashion.

44. Fructose 2,6-bisphosphate stabilizes the R state of the enzyme.

45. (a) Curiously, the enzyme uses ADP as the phosphoryl donor rather than ATP.

(b) Both AMP and ATP behave as competitive inhibitors of ADP, the phosphoryl donor. Apparently, the *P. furiosus* enzyme is not allosterically inhibited by ATP.

46. (a) If both enzymes operated simultaneously, the following reactions would take place:



The net result would be simply:

$$ATP + H_2O \longrightarrow ADP + P_i$$

The energy of ATP hydrolysis would be released as heat. (b) Not really. For the cycle to generate heat, both enzymes must be functional at the same time in the same cell.

(c) The species *B. terrestris* and *B. rufocinctus* might show some futile cycling because both enzymes are active to a substantial degree.

(d) No. These results simply suggest that simultaneous activity of phosphofructokinase and fructose 1,6-bisphosphatase is unlikely to be employed to generate heat in the species shown.

Chapter 17

1. Pyruvate dehydrogenase catalyzes the decarboxylation of pyruvate and the formation of acetyllipoamide. Dihydrolipoyl transacetylase catalyzes the formation of acetyl CoA. Dihydrolipoyl dehydrogenase catalyzes the reduction of the oxidized lipoic acid. The kinase associated with the complex phosphorylates and inactivates the complex, whereas the phosphatase dephosphorylates and activates the complex.

2. Thiamine pyrophosphate plays a role in the decarboxylation of pyruvate. Lipoic acid (as lipoamide) transfers the acetyl group. Coenzyme A accepts the acetyl group from lipoic acid to form acetyl CoA. FAD accepts the electrons and hydrogen ions when reduced lipoic acid is oxidized. NAD⁺ accepts electrons from FADH₂.

3. Catalytic coenzymes (TPP, lipoic acid, and FAD) are modified but regenerated in each reaction cycle. Thus, they can play a role in the processing of many molecules of pyruvate. Stoichiometric coenzymes (coenzyme A and NAD⁺) are used in only one reaction because they are the components of products of the reaction.

4. The advantages are as follows:

The reaction is facilitated by having the active sites in proximity. The reactants do not leave the enzyme until the final product is formed.

Constraining the reactants minimizes loss due to diffusion and minimizes side reactions.

All of the enzymes are present in the correct amounts. Regulation is more efficient because the regulatory enzymes—the kinase and phosphatase—are part of the complex.

5. (a) After one round of the citric acid cycle, the label emerges in C-2 and C-3 of oxaloacetate. (b) The label emerges in CO_2 in the formation of acetyl CoA from pyruvate. (c) After one round of the citric acid cycle, the label emerges in C-1 and C-4 of oxaloacetate. (d) and (e) Same fate as that in part *a*.

6. (a) Isocitrate lyase and malate synthase are required in addition to the enzymes of the citric acid cycle.

(b) 2 Acetyl CoA + 2 NAD⁺ + FAD + 3 $H_2O \rightarrow$ oxaloacetate + 2 CoA + 2 NADH + FADH₂ + 3 H⁺.

(c) No. Hence, mammals cannot carry out the net synthesis of oxaloacetate from acetyl CoA.

7. $-41.0 \text{ kJ mol}^{-1} (-9.8 \text{ kcal mol}^{-1})$

8. Enzymes or enzyme complexes are biological catalysts. Recall that a catalyst facilitates a chemical reaction without the catalyst itself being permanently altered. Oxaloacetate can be thought of as a catalyst because it binds to an acetyl group, leads to the oxidative decarboxylation of the two carbon atoms, and is regenerated at the completion of a cycle. In essence, oxaloacetate (and any cycle intermediate) acts as a catalyst.

9. Thiamine thiazolone pyrophosphate is a transition-state analog. The sulfur-containing ring of this analog is uncharged, and so it closely resembles the transition state of the normal coenzyme in thiamine-catalyzed reactions (e.g., the uncharged resonance form of hydroxyethyl-TPP).

10. A decrease in the amount of O_2 will necessitate an increase in anaerobic glycolysis for energy production, leading to the generation of a large amount of lactic acid. Under conditions of shock, the kinase inhibitor is administered to ensure that pyruvate dehydrogenase is operating maximally.

11. Acetyllipoamide and acetyl CoA

12. In muscle, the acetyl CoA generated by the complex is used for energy generation. Consequently, signals that indicate an energy-rich state (high ratios of ATP/ADP and NADH/NAD⁺) inhibit the complex, whereas the reverse conditions stimulate the enzyme. Calcium as the signal for muscle contraction (and, hence, energy need) also stimulates the enzyme. In liver, acetyl CoA derived from pyruvate is used for biosynthetic purposes, such as fatty acid synthesis. Insulin, the hormone denoting the fed state, stimulates the complex.

13. (a) Enhanced kinase activity will result in a decrease in the activity of the PDH complex because phosphorylation by the kinase inhibits the complex.

(b) Phosphatase activates the complex by removing a phosphate. If the phosphatase activity is diminished, the activity of the PDH complex also will decrease.

14. She might have been ingesting, in some fashion, the arsenite from the peeling paint or the wallpaper. Also, she might have been breathing arsine gas from the wallpaper, which would be oxidized to arsenite in her body. In any of these circumstances, the arsenite inhibited enzymes that require lipoic acid—notably, the PDH complex.

15. The TCA cycle depends on a steady supply of NAD⁺, which is typically generated from NADH by reaction of the NADH with oxygen. If there is no oxygen to accept the electrons, the citric acid cycle will cease to operate.

16. (a) The steady-state concentrations of the products are low compared with those of the substrates. (b) The ratio of malate to oxaloacetate must be greater than 1.57×10^4 for oxaloacetate to be formed.

17.

 $\begin{array}{c} Pyruvate\\ dehydrogenase\\ Pyruvate + CoA + NAD^{+} & \underbrace{-complex}_{acetyl} CoA + CO_{2} + NADH \end{array}$

Pyruvate
Pyruvate +
$$CO_2$$
 + ATP + H_2O $\xrightarrow{carboxylase}$
oxaloacetate + ADP + P_i + H^+

Citrate

 $Oxaloacetate + acetyl CoA + H_2O \xrightarrow{synthase} citrate + CoA + H^+$

Citrate $\xrightarrow{\text{Aconitase}}$ isocitrate

Isocitrate + NAD⁺ $\xrightarrow{\text{dehydrogenase}} \alpha$ -ketoglutarate + CO₂ + NADH

Net: 2 Pyruvate + 2 NAD⁺ + ATP + H₂O \longrightarrow α -ketoglutarate + CO₂ + ADP + P_i + 2 NADH + 3 H⁺

18. Succinate will increase in concentration, followed by α -ketoglutarate and the other intermediates "upstream" of the site of inhibition. Succinate has two methylene groups that are required for the dehydrogenation, whereas malonate has but one. 19. Pyruvate carboxylase should be active only when the acetyl CoA concentration is high. Acetyl CoA might accumulate if the energy needs of the cell are not being met, because of a deficiency of oxaloacetate. Under these conditions the pyruvate carboxylase catalyzes an anapleurotic reaction. Alternatively, acetyl CoA might accumulate because the energy needs of the cell have been met. In this circumstance, pyruvate will be converted back into glucose, and the first step in this conversion is the formation of oxaloacetate. 20. The energy released when succinate is reduced to fumarate is not sufficient to power the synthesis of NADH but is sufficient to reduce FAD.

21. Citrate is a tertiary alcohol that cannot be oxidized, because oxidation requires a hydrogen atom to be removed from the alcohol and a hydrogen atom to be removed from the carbon atom bonded to the alcohol. No such hydrogen exists in citrate. The isomerization converts the tertiary alcohol into isocitrate, which is a secondary alcohol that can be oxidized.

22. Because the enzyme nucleoside diphosphokinase transfers a phosphoryl group from GTP (or any nucleoside triphosphate) to ADP according to the reversible reaction:

 $GTP + ADP \Longrightarrow GDP + ATP$

23. The reaction is powered by the hydrolysis of a thioester. Acetyl CoA provides the thioester that is converted into citryl CoA. When this thioester is hydrolyzed, citrate is formed in an irreversible reaction.

24. We cannot get the net conversion of fats into glucose, because the only means to get the carbon atoms from fats into oxaloacetate, the precursor of glucose, is through the citric acid cycle. However, although two carbon atoms enter the cycle as acetyl CoA, two carbon atoms are lost as CO_2 before oxaloacetate is formed. Thus, although some carbon atoms from fats may end up as carbon atoms in glucose, we cannot obtain a *net* synthesis of glucose from fats. 25. Acetyl CoA will inhibit the complex. Glucose metabolism to pyruvate will be slowed because acetyl CoA is being derived from an alternative source.

26. The enol intermediate of acetyl CoA attacks the carbonyl carbon atom of glyoxylate to form a C–C bond. This reaction is like the condensation of oxaloacetate with the enol intermediate of acetyl CoA in the reaction catalyzed by citrate synthase. Glyoxylate contains a hydrogen atom in place of the $-CH_2COO^-$ group of oxaloacetate; the reactions are otherwise nearly identical. 27. Citrate is a symmetric molecule. Consequently, the investigators assumed that the two $-CH_2COO^-$ groups in it would react identically. Thus, for every citrate molecule undergoing the reactions shown in path 1, they thought that another citrate

molecule would react as shown in path 2. If so, then only *half* the label should have emerged in the CO_2 .



28. Call one hydrogen atom A and the other B. Now suppose that an enzyme binds three groups of this substrate—X, Y, and H—at three complementary sites. The adjoining diagram shows X, Y, and H_A bound to three points on the enzyme. In contrast, X, Y, and H_B cannot be bound to this active site; two of these three groups can be bound, but not all three. Thus, H_A and H_B will have different fates.



Sterically nonequivalent groups such as H_A and H_B will almost always be distinguished in enzymatic reactions. The essence of the differentiation of these groups is that the enzyme holds the substrate in a specific orientation. Attachment at three points, as depicted in the diagram, is a readily visualized way of achieving a particular orientation of the substrate, but it is not the only means of doing so. 29. (a) The complete oxidation of citrate requires 4.5 μ mol of O₂ for every micromole of citrate.

$$C_6H_8O_7 + 4.5O_2 \longrightarrow 6CO_2 + 4H_2O$$

Thus, 13.5 μ mol of O₂ would be consumed by 3 μ mol of citrate. (b) Citrate led to the consumption of far more O₂ than can be accounted for simply by the oxidation of citrate itself. Citrate thus facilitated O₂ consumption. 30. (a) In the absence of arsenite, the amount of citrate remained constant. In its presence, the concentration of citrate fell, suggesting that it was being metabolized.

(b) The action of arsenite is not altered. Citrate still disappears.(c) Arsenite is preventing the regeneration of citrate. Recall (pp. 517–518) that arsenite inhibits the pyruvate dehydrogenase complex.31. (a) The initial infection is unaffected by the absence of isoci-

trate lyase, but the absence of this enzyme inhibits the latent phase of the infection. (b) Yes

(c) A critic could say that, in the process of deleting the isocitrate lyase gene, some other gene was damaged, and it is the absence of this other gene that prevents latent infection. Reinserting the isocitrate lyase gene into the bacteria from which it had been removed renders the criticism less valid.

(d) Isocitrate lyase enables the bacteria to synthesize carbohydrates that are necessary for survival, including carbohydrate components of the cell membrane.

Chapter 18

1. In fermentations, organic compounds are both the donors and the acceptors of electrons. In respiration, the electron donor is usually an organic compound, whereas the electron acceptor is an inorganic molecule, such as oxygen.

2. Biochemists use E'_0 , the value at pH 7, whereas chemists use E_0 , the value in 1 M H⁺. The prime denotes that pH 7 is the standard state.

3. The reduction potential of $FADH_2$ is less than that of NADH (see Table 18.1). Consequently, when those electrons are passed along to oxygen, less energy is released. The consequence of the difference is that electron flow from $FADH_2$ to O_2 pumps fewer protons than do the electrons from NADH.

4. The $\Delta G^{o'}$ for the reduction of oxygen by FADH₂ is $-200 \text{ kJ mol}^{-1}(-48 \text{ kcal mol}^{-1})$.

5. $\Delta G^{\circ'}$ is + 67 kJ mol⁻¹ (+16.1 kcal mol⁻¹) for oxidation by NAD⁺ and -3.8 kJ mol⁻¹ (-0.92 kcal mol⁻¹) for oxidation by FAD. The oxidation of succinate by NAD⁺ is not thermodynamically feasible.

6. Pyruvate accepts electrons and is thus the oxidant. NADH gives up electrons and is the reductant.

7. $\Delta G^{\circ\prime} = -nF\Delta E'_0$

8. The $\Delta E'_0$ value of iron can be altered by changing the environment of the ion.

9. c, e, b, a, d.

10. (a) 4; (b) 3; (c) 1; (d) 5; (e) 2.

11. The 10 isoprene units render coenzyme Q soluble in the hydrophobic environment of the inner mitochondrial membrane. The two oxygen atoms can reversibly bind two electrons and two protons as the molecule transitions between the quinone form and quinol form. 12. Rotenone: NADH, NADH-Q oxidoreductase will be reduced. The remainder will be oxidized. Antimycin A: NADH, NADH-Q oxidoreductase and coenzyme Q will be reduced. The remainder will be oxidized. Cyanide: All will be reduced.

13. Complex I would be reduced, whereas Complexes II, III, and IV would be oxidized. The citric acid cycle would become reduced because it has no way to oxidize NADH.

14. The respirasome is another example of the use of supramolecular complexes in biochemistry. Having the three complexes that are proton pumps associated with one another will enhance the efficiency of electron flow from complex to complex, which in turn will cause more-efficient proton pumping.

15. Hydroxyl radical ($OH \cdot$), hydrogen peroxide (H_2O_2), superoxide ion (O_2^-), and peroxide (O_2^{2-}). These small molecules react with a host of macromolecules—including proteins, nucleotides, and membranes—to disrupt cell structure and function.

16. The ATP is recycled by ATP-generating processes, most notably oxidative phosphorylation.

17. (a) 12.5; (b) 14; (c) 32; (d) 13.5; (e) 30; (f) 16.

18. (a) It blocks electron transport and proton pumping at Complex IV. (b) It blocks electron transport and ATP synthesis by inhibiting the exchange of ATP and ADP across the inner mitochondrial membrane. (c) It blocks electron transport and proton pumping at Complex I. (d) It blocks ATP synthesis without inhibiting electron transport by dissipating the proton gradient. (e) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex IV. (f) It blocks electron transport and proton pumping at Complex

19. If the proton gradient is not dissipated by the influx of protons into a mitochondrion with the generation of ATP, eventually the outside of the mitochondrion develops such a large positive charge that the electron-transport chain can no longer pump protons against the gradient.

20. The subunits are jostled by background thermal energy (Brownian motion). The proton gradient makes clockwise rotation more likely because that direction results in protons flowing down their concentration gradient.

21. In the presence of poorly functioning mitochondria, the only means of generating ATP is by anaerobic glycolysis, which will lead to an accumulation of lactic acid in blood.

22. If ADP cannot get into mitochondria, the electron-transport chain will cease to function because there will be no acceptor for the energy. NADH will build up in the matrix. Recall that NADH inhibits some citric acid cycle enzymes and that NAD⁺ is required by several citric acid cycle enzymes. Glycolysis will stop functioning aerobically but will switch to anaerobic glycolysis so that the NADH can be reoxidized to NAD⁺ by lactate dehydrogenase. 23. (a) No effect; mitochondria cannot metabolize glucose.

(b) No effect; no fuel is present to power the synthesis of ATP.
(c) The [O₂] falls because citrate is a fuel and ATP can be formed from ADP and P_i.

(d) Oxygen consumption stops because oligomycin inhibits ATP synthesis, which is coupled to the activity of the electron-transport chain.

(e) No effect, for the reasons given in part *d*.

(f) $[O_2]$ falls rapidly because the system is uncoupled and does not require ATP synthesis to lower the proton-motive force.

(g) $[O_2]$ falls, though at a lower rate. Rotenone inhibits Complex I, but the presence of succinate will enable electrons to enter at Complex II.

(h) Oxygen consumption ceases because Complex IV is inhibited and the entire chain backs up.

24. (a) The P : O ratio is equal to the product of $(H^+/2 e^-)$ and (P/H^+) . Note that the P : O ratio is identical with the P : 2 e^- ratio. (b) 2.5 and 1.5, respectively.

25. Cyanide can be lethal because it binds to the ferric form of cytochrome oxidase and thereby inhibits oxidative phosphorylation. Nitrite converts ferrohemoglobin into ferrihemoglobin, which also binds cyanide. Thus, ferrihemoglobin competes with cytochrome oxidase for cyanide. This competition is therapeutically effective because the amount of ferrihemoglobin that can be formed without impairing oxygen transport is much greater than the amount of cytochrome oxidase.

26. Such a defect (called Luft syndrome) was found in a 38-yearold woman who was incapable of performing prolonged physical work. Her basal metabolic rate was more than twice normal, but her thyroid function was normal. A muscle biopsy showed that her mitochondria were highly variable and atypical in structure. Biochemical studies then revealed that oxidation and phosphorylation were not tightly coupled in these mitochondria. In this patient, much of the energy of fuel molecules was converted into heat rather than ATP.

27. Triose phosphate isomerase converts dihydroxyacetone phosphate (a potential dead end) into glyceraldehyde 3-phosphate (a mainstream glycolytic intermediate).

28. This inhibitor (like antimycin A) blocks the reduction of cytochrome c_1 by QH₂, the crossover point.

29. If oxidative phosphorylation were uncoupled, no ATP could be produced. In a futile attempt to generate ATP, much fuel would be consumed. The danger lies in the dose. Too much uncoupling would lead to tissue damage in highly aerobic organs such as the brain and heart, which would have severe consequences for the organism as a whole. The energy that is normally transformed into ATP would be released as heat. To maintain body temperature, sweating might increase, although the very process of sweating itself depends on ATP.

30. If ATP and ADP cannot exchange between the matrix and the mitochondria, ATP synthase will cease to function because its substrate ADP is absent. The proton gradient will eventually become so large that the energy released by the electron-transport chain will not be great enough to pump protons against the larger-thannormal gradient.

31. Add the inhibitor with and without an uncoupler, and monitor the rate of O_2 consumption. If the O_2 consumption increases again in the presence of inhibitor and uncoupler, the inhibitor must be inhibiting ATP synthase. If the uncoupler has no effect on the inhibition, the inhibitor is inhibiting the electron-transport chain.

32. Presumably, because the muscle has greater energy needs, especially during exercise, it will require more ATP. This requirement means that more sites of oxidative phosphorylation are called for, and these sites can be provided by an increase in the amount of cristae.33. The arginine residue, with its positive charge, will facilitate proton release from aspartic acid by stabilizing the negatively charged aspartate.

34. 4; 4.7

35. The ATP synthase would pump protons at the expense of ATP hydrolysis, thus maintaining the proton-motive force. The synthase would function as an ATPase. There is some evidence that damaged mitochondria use this tactic to maintain, at least temporarily, the proton-motive force.

36. It suggests that malfunctioning mitochondria may play a role in the development of Parkinson disease. Specifically, it implicates Complex I.

37. The extra negative charge on ATP relative to that on ADP accounts for ATP's more-rapid translocation out of the mitochondrial matrix. If the charge differences between ATP and ADP were lessened by the binding of Mg^{2+} , ADP might more readily compete with ATP for transport to the cytoplasm.

38. When all of the available ADP has been converted into ATP, ATP synthase can no longer function. The proton gradient becomes large enough that the energy of the electron-transport chain is not enough to pump against the gradient, and electron transport and, hence, oxygen consumption falls.

39. The effect on the proton gradient is the same in each case.

40. ATP export from the matrix. Phosphate import into the matrix. 41. Recall from the discussion of enzyme-catalyzed reactions that the direction of a reaction is determined by the ΔG difference between substrate and products. An enzyme speeds up the rate of both the forward and the backward reactions. The hydrolysis of ATP is exergonic, and so ATP synthase will enhance the hydrolytic reaction.

42. The cytoplasmic kinases thereby obtaining preferential access to the exported ATP.

43. The organic acids in the blood are indications that the mice are deriving a large part of their energy needs through anaerobic gly-colysis. Lactate is the end product of anaerobic glycolysis. Alanine is an aminated transport form of lactate. Alanine formation plays a role in succinate formation, which is caused by the reduced state of the mitochondria.



The electron-transport chain is slowed because the inner mitochondrial membrane is hyperpolarized. Without ADP to accept the energy of the proton-motive force, the membrane becomes polarized to such an extent that protons can no longer be pumped. The excess H_2O_2 is probably due to the fact that the superoxide radical is present in higher concentration because the oxygen can no longer be effectively reduced.

$$O_2 \cdot \overline{} + O_2 \cdot \overline{} + 2 H^+ \longrightarrow O_2 + H_2O_2$$

Indeed, these mice display evidence of such oxidative damage. 44. (a) Vitamins C and E.

(b) Exercise induces superoxide dismutase, which converts ROS in hydrogen peroxide and oxygen.

(c) The answer to this question is not fully established. Two possibilities are (1) the suppression of ROS by vitamins prevents the expression of more superoxide dismutase and (2) some ROS may be signal molecules required to stimulate insulin-sensitivity pathways.
45. (a) Succinate is oxidized by Complex II, and the electrons are used to establish a proton-motive force that powers ATP synthesis.
(b) The ability to synthesize ATP is greatly reduced.

(c) Because the goal was to measure ATP hydrolysis. If succinate had been added in the presence of ATP, no reaction would have taken place, because of respiratory control.

(d) The mutation has little effect on the ability of the enzyme to catalyze the hydrolysis of ATP.

(e) They suggest two things: (1) the mutation did not affect the catalytic site on the enzyme, because ATP synthase is still capable of catalyzing the reverse reaction, and (2) the mutation did not affect the amount of enzyme present, given that the controls and patients had similar amounts of activity.

46. The absolute configuration of thiophosphate indicates that inversion at phosphorus has taken place in the reaction catalyzed by ATP synthase. This result is consistent with an in-line phosphoryl-transfer reaction taking place in a single step. The retention of configuration in the Ca^{2+} -ATPase reaction points to two phosphoryl-transfer reactions—inversion by the first and a return to the starting configuration by the second. The Ca^{2+} -ATPase reaction proceeds by a phosphorylated enzyme intermediate.

Chapter 19

1. Photosystem I generates ferredoxin, which reduces $NADP^+$ to NADPH, a biosynthetic reducing power. Photosystem II activates the manganese complex, an oxidant capable of oxidizing water, generating electrons for photosynthesis, and generating protons to form a proton gradient and to reduce $NADP^+$ and O_2 .

2. The light reactions take place on thylakoid membranes. Increasing the membrane surface increases the number of ATPand NADH-generating sites.

3. These complexes absorb more light than can a reaction center alone. The light-harvesting complexes funnel light to the reaction centers.

4. NADP⁺ is the acceptor. H_2O is the donor. Light energy.

5. The charge gradient, a component of the proton-motive force in mitochondria, is neutralized by the influx of Mg^{2+} into the lumen of the thylakoid membranes.

6. Chlorophyll is readily inserted into the hydrophobic interior of the thylakoid membranes.

7. Protons released by the oxidation of water; protons pumped into the lumen by the cytochrome *bf* complex; protons removed from the stroma by the reduction of NADP⁺ and plastoquinone. 8. 700-nm photons have an energy content of 172 kJ mol⁻¹. The absorption of light by photosystem I results in a $\Delta E'_0$ of -1.0 V. Recall that $\Delta G'_0 = -nF \Delta E'_0$, where F = 96.48 kJ mol⁻¹ V⁻¹. Under standard conditions, the energy change for the electrons is 96.5 kJ. Thus, the efficiency is 96.5/172 = 56%.

9. The electron flow from PS II to PS I is uphill, or exergonic. For this uphill flow, ATP would need to be consumed, defeating the purpose of photosynthesis.

10. $\Delta E'_0 = 10.11$ V, and $\Delta G^{\circ\prime} = -21.3$ kJ mol⁻¹ (-5.1 kcal mol⁻¹). 11. (a) All ecosystems require an energy source from outside the system, because the chemical-energy sources will ultimately be limited. The photosynthetic conversion of sunlight is one example of such a conversion.

(b) Not at all. Spock would point out that chemicals other than water can donate electrons and protons.

DCMU inhibits electron transfer in the link between photosystems II and I. O₂ can evolve in the presence of DCMU if an artificial electron acceptor such as ferricyanide can accept electrons from Q.
 DCMU will have no effect, because it blocks photosystem II, and cyclic photophosphorylation uses photosystem I and the cytochrome *bf* complex.

14. (a) 120 kJ einstein⁻¹ (28.7 kcal einstein⁻¹)

(b) 1.24 V

(c) One 1000-nm photon has the free energy content of 2.4 molecules of ATP. A minimum of 0.42 photon is needed to drive the synthesis of a molecule of ATP.

15. At this distance, the expected rate is one electron per second.16. The distance doubles, and so the rate should decrease by a factor of 64 to 640 ps.

17. The cristae.

18. In eukaryotes, both processes take place in specialized organelles. Both depend on high-energy electrons to generate ATP. In oxidative phosphorylation, the high-energy electrons originate in fuels and are extracted as reducing power in the form of NADH. In photosynthesis, the high-energy electrons are generated by light and are captured as reducing power in the form of NADPH. Both processes use redox reactions to generate a proton gradient, and the enzymes that convert the proton gradient into ATP are very similar in both processes. In both systems, electron transport takes place in membranes inside organelles.

19. We need to factor in the NADPH because it is an energy-rich molecule. Recall from Chapter 18, that NADH is worth 2.5 ATP if oxidized by the electron-transport chain. 12 NADPH = 30 ATP. Eighteen molecules of ATP are used directly, and so the equivalent of 48 molecules of ATP is required for the synthesis of glucose. 20. Both photosynthesis and cellular respiration are powered by high-energy electrons flowing toward a more-stable state. In cellular respiration, the high-energy electrons are derived from the

oxidation of carbon fuels as NADH and FADH₂. They release their energy as they reduce oxygen. In photosynthesis, high-energy electrons are generated by absorbing light energy, and they find stability in photosystem I and ferridoxin.

21. The electrons flow through photosystem II directly to ferricyanide. No other steps are required.

22. (a) Thioredoxin

(b) The control enzyme is unaffected, but the mitochondrial

enzyme with part of the chloroplast γ subunit increases activity as the concentration of DTT increases.

(c) The increase was even larger when thioredoxin was present. Thioredoxin is the natural reductant for the chloroplast enzyme, and so it presumably operates more efficiently than would DTT, which probably functions to keep the thioredoxin reduced.(d) They seem to have done so.

(e) The enzyme is susceptible to control by the redox state. In plant cells, reduced thioredoxin is generated by photosystem I. Thus, the enzyme is active when photosynthesis is taking place. (f) Cysteine

(g) Group-specific modification or site-specific mutagenesis.

Chapter 20

1. The Calvin cycle is the primary means of converting gaseous $\rm CO_2$ into organic matter—that is, biomolecules. Essentially, every carbon atom in your body passed through rubisco and the Calvin cycle at some time in the past.

Krebs cycle
Matrix
Carbon chemistry for oxidative phosphorylation
Releases CO_2
Generates high-energy electrons (NADPH)
Regenerates starting compound (oxaloacetate)
Generates ATP or GTP
Simple stoichiometry

3. (a) 3-Phosphoglycerate. (b) The other members of the Calvin cycle.

4. Stage 1 is the fixation of CO_2 with ribulose 1,5-bisphosphate and the subsequent formation of 3-phosphoglycerate. Stage 2 is the

conversion of some of the 3-phosphoglycerate into hexose. Stage 3 is the regeneration of ribulose 1,5-bisphosphate.

5. It catalyzes a crucial reaction, but it is highly inefficient. Consequently, it is required in large amounts to overcome its slow catalysis.

6. Because carbamate forms only in the presence of CO_2 , this property prevents rubisco from catalyzing the oxygenase reaction exclusively when CO_2 is absent.

7. Because NADPH is generated in the chloroplasts by the light reactions.

 8. The concentration of 3-phosphoglycerate would increase, whereas that of ribulose 1,5-bisphosphate would decrease.
 9. The concentration of 3-phosphoglycerate would decrease,

whereas that of ribulose 1,5-bisphosphate would increase. 10. Aspartate + glyoxylate → oxaloacetate + glycine 11. The oxygenase activity of rubisco increases with temperature.

Crabgrass is a C_4 plant, whereas most grasses lack this capability. Consequently, the crabgrass will thrive at the hottest part of the summer because the C_4 pathway provides an ample supply of CO_2 . 12. The C_4 pathway allows the CO_2 concentration to increase at the site of carbon fixation. High concentrations of CO_2 inhibit the oxygenase reaction of rubisco. This inhibition is important for tropical plants because the oxygenase activity increases more rapidly with temperature than does the carboxylase activity.

13. ATP is required to form phosphoenolpyruvate (PEP) from pyruvate. The PEP combines with CO_2 to form oxaloacetate and, subsequently, malate. Two ATP molecules are required because a second ATP molecule is required to phosphorylate AMP to ADP.

14. Photorespiration is the consumption of oxygen by plants with the production of CO_2 , but it does not generate energy. Photorespiration is due to the oxygenase activity of rubisco. It is wasteful because, instead of fixing CO_2 for conversion into hexoses, rubisco is generating CO_2 .

15. As global warming progresses, C_4 plants will invade the higher latitudes, and C_3 plants will retreat to cooler regions.

16. The light reactions lead to an increase in the stromal concentrations of NADPH, reduced ferredoxin, and Mg^{2+} , as well as an increase in pH.

17. The enzymes catalyze the transformation of the five-carbon sugar formed by the oxidative phase of the pentose phosphate pathway into fructose 6-phosphate and glyceraldehyde 3-phosphate, intermediates in glycolysis (and gluconeogenesis).

18. The label emerges at C-5 of ribulose 5-phosphate.

19. Oxidative decarboxylation of isocitrate to α -ketoglutarate. A

 β -ketoacid intermediate is formed in both reactions.

20. (a) 5 Glucose 6-phosphate + ATP \rightarrow

6 ribose 5-phosphate + ADP + H⁺.

(b) Glucose 6-phosphate + 12 NADP⁺ + 7 $H_2O \rightarrow$ 6 CO₂ + 12 NADPH + 12 H⁺ + P_i.

21. The nonoxidative phase of the pentose phosphate pathway can be used to convert three molecules of ribose 5-phosphate into two molecules of fructose 6-phosphate and one molecule of glyceraldehyde 3-phosphate. These molecules are components of the glycolytic pathway.

22. The conversion of fructose 6-phosphate into fructose 1,6bisphosphate by phosphofructokinase requires ATP.

23. When much NADPH is required. The oxidative phase of the pentose phosphate pathway is followed by the nonoxidative phase. The resulting fructose 6-phosphate and glyceraldehyde 3-phosphate are used to generate glucose 6-phosphate through gluconeogenesis, and the cycle is repeated until the equivalent of one glucose molecule is oxidized to CO₂.

24. Fava beans contain pamaquine, a purine glycoside that can lead to the generation of peroxides—reactive oxygen species that can damage membranes as well as other biomolecules. Glutathione is used to detoxify the ROS. The regeneration of glutathione depends on an adequate supply of NADPH, which is synthesized by the oxidative phase of the pentose phosphate pathway. People with low levels of the dehydrogenase are especially susceptible to pamaquine toxicity.

25. Because red blood cells do not have mitochondria and the only means to obtain NADPH is through the pentose phosphate pathway. There are biochemical means to convert mitochondrial NADH into cytoplasmic NADPH.

26. Reactive peroxides are a type of reactive oxygen species. The enzyme glutathione peroxidase uses reduced glutathione to neutralize peroxides by converting them into alcohols while generating oxidized glutathione. Reduced glutathione is regenerated by glutathione reductase with the use of NADPH, the product of the oxidative phase of the pentose phosphate pathway.

27. $\Delta E'_0$ for the reduction of glutathione by NADPH is + 0.09 V. Hence, $\Delta G^{\circ'}$ is -17.4 kJ mol⁻¹ (- 4.2 kcal mol⁻¹), which corresponds to an equilibrium constant of 1126. The required [NADPH]/[NADP⁺] ratio is 8.9 × 10⁻⁵. 28.



30. Incubate an aliquot of a tissue homogenate with glucose labeled with ¹⁴C at C-1, and incubate another with glucose labeled with ¹⁴C at C-6. Compare the radioactivity of the CO_2 produced by the two samples. The rationale of this experiment is that only C-1 is decarboxylated by the pentose phosphate pathway, whereas C-1 and C-6 are decarboxylated equally when glucose is metabolized by the glycolytic pathway, the pyruvate dehydrogenase complex, and the citric acid cycle. The reason for the equivalence of C-1 and C-6 in the latter set of reactions is that glyceraldehyde 3-phosphate and dihydroxyacetone phosphate are rapidly interconverted by triose phosphate isomerase.

31. The reduction of each mole of CO_2 to the level of a hexose requires two moles of NADPH. The reduction of NADP⁺ is a two-electron process. Hence, the formation of two moles of NADPH requires the pumping of four moles of electrons by photosystem I. The electrons given up by photosystem I are replenished by photosystem II, which needs to absorb an equal number of photons. Hence, eight photons are needed to generate the required NADPH. The energy input of eight moles of photons is 1594 kJ (381 kcal). Thus, the overall efficiency of photosynthesis under standard conditions is at least 477/1594, or 30%. 32. It is neither a violation nor a miracle. The equation on page 580 requires not only 18 ATP, but also 12 NADPH. These electrons, if transferred to NAD⁺ and used in the electron-transport chain, would yield 30 ATP. Thus, the synthesis of glucose requires the equivalent of 48 ATP.

33. (a) The curve on the right in graph A was generated by the C_4 plant. Recall that the oxygenase activity of rubisco increases with temperature more rapidly than does the carboxylase activity. Consequently, at higher temperatures, the C_3 plants would fix less carbon. Because C_4 plants can maintain a higher CO_2 concentration, the rise in temperature is less deleterious.

(b) The oxygenase activity will predominate. Additionally, when the temperature rise is very high, the evaporation of water might become a problem. The higher temperatures can begin to damage protein structures as well.

(c) The C_4 pathway is a very effective active-transport system for concentrating CO_2 , even when environmental concentrations are very low.

(d) With the assumption that the plants have approximately the same capability to fix CO_2 , the C_4 pathway is apparently the rate-limiting step in C_4 plants.

Chapter 21

1. Glycogen is an important fuel reserve for several reasons. The controlled breakdown of glycogen and release of glucose increase the amount of glucose that is available between meals. Hence, glycogen serves as a buffer to maintain blood-glucose levels. Glycogen's role in maintaining blood-glucose levels is especially important because glucose is virtually the only fuel used by the brain, except during prolonged starvation. Moreover, the glucose from glycogen is readily mobilized and is therefore a good source of energy for sudden, strenuous activity. Unlike fatty acids, the released glucose can provide energy in the absence of oxygen and can thus supply energy for anaerobic activity.

2. As an unbranched polymer, α -amylose has only one nonreducing end. Therefore, only one glycogen phosphorylase molecule could degrade each α -amylose molecule. Because glycogen is highly branched, there are many nonreducing ends per molecule. Consequently, many phosphorylase molecules can release many glucose molecules per glycogen molecule.

3. The patient has a deficiency of the branching enzyme.

4. In muscle, the *b* form of phosphorylase is activated by AMP. In the liver, the *a* form is inhibited by glucose. The difference corresponds to the difference in the metabolic role of glycogen in each tissue. Muscle uses glycogen as a fuel for contraction, whereas the liver uses glycogen to maintain blood-glucose levels.

 Cells maintain the [Pi]/[glucose 1-phosphate] ratio at greater than 100, substantially favoring phosphorolysis. We see here an example of how the cell can alter the free-energy change to favor a reaction taking place by altering the ratio of substrate and product.
 The high level of glucose 6-phosphate in von Gierke disease, resulting from the absence of glucose 6-phosphatase or the transporter, shifts the allosteric equilibrium of phosphorylated glycogen synthase toward the active form.

7. The phosphoryl donor is glucose 1,6-bisphosphate, which is formed from glucose 1-phosphate and ATP in a reaction catalyzed by phosphoglucokinase.

8. The different manifestations correspond to the different roles of the liver and muscle. Liver glycogen phosphorylase plays a crucial role in the maintenance of blood-glucose levels. Recall that glucose is the primary fuel for the brain. Muscle glycogen phosphorylase provides glucose only for the muscle and, even then, only when the energy needs of the muscle are high, as during exercise. The fact that there are two different diseases suggests that there are two different isozymic forms of the glycogen phosphorylase—a liverspecific isozyme and a muscle-specific isozyme.

9. Water is excluded from the active site to prevent hydrolysis. The entry of water could lead to the formation of glucose rather than glucose 1-phosphate. A site-specific mutagenesis experiment is revealing in this regard. In phosphorylase, Tyr 573 is hydrogen bonded to the 2'-OH group of a glucose residue. The ratio of glucose 1-phosphate to glucose product is 9000 : 1 for the wild-type enzyme, and 500 : 1 for the Phe 573 mutant. Model building suggests that a water molecule occupies the site normally filled by the phenolic OH group of tyrosine and occasionally attacks the oxocarbonium ion intermediate to form glucose.

10. The amylase activity was necessary to remove all of the glycogen from the glycogenin. Recall that glycogenin synthesizes oligosaccharides of about eight glucose units, and then activity stops. Consequently, if the glucose residues are not removed by extensive amylase treatment, glycogenin will not function.

11. The substrate can be handed directly from the transferase site to the debranching site.

12. During exercise, [ATP] falls and [AMP] rises. Recall that AMP is an allosteric activator of glycogen phosphorylase *b*. Thus, even in the absence of covalent modification by phosphorylase kinase, glycogen is degraded.

13. Although glucose 1-phosphate is the actual product of the phosphorylase reaction, glucose 6-phosphate is a more versatile molecule with respect to metabolism. Among other fates, glucose-6-phosphate can be processed to yield energy or building blocks. In the liver, glucose 6-phosphate can be converted into glucose and released into the blood.

14. Epinephrine binds to its G-protein-coupled receptor. The resulting structural changes activate a G_{α} protein, which in turn activates adenyl cyclase. Adenyl cyclase synthesizes cAMP, which activates protein kinase A. Protein kinase A partly activates phosphoryl kinase, which phosphorylates and activates glycogen phosphorylase. The calcium released during muscle contraction further activates the phosphorylase kinase, leading to further stimulation of glycogen phosphorylase.

15. First, the signal-transduction pathway is shut down when the initiating hormone is no longer present. Second, the inherent GTPase activity of the G protein converts the bound GTP into inactive GDP. Third, phosphodiesterases convert cyclic AMP into AMP. Fourth, PP1 removes the phosphoryl group from glycogen phosphorylase, converting the enzyme into the usually inactive *b* form.

16. It prevents both from operating simultaneously, which would lead to a useless expenditure of energy. See the answer to Problem 24.17. All these symptoms suggest central nervous system problems.

If exercise is exhaustive enough or the athlete has not prepared well enough or both, liver glycogen also can be depleted. The brain depends on glucose derived from liver glycogen. The symptoms suggest that the brain is not getting enough fuel.

18. Liver phosphorylase *a* is inhibited by glucose, which facilitates the $R \rightarrow T$ transition. This transition releases PP1, which inactivates glycogen breakdown and stimulates glycogen synthesis. Muscle phosphorylase is insensitive to glucose.

19. The presence of high concentrations of glucose 6-phosphate indicates that glucose is abundant and that it is not being used by glycolysis. Therefore, this valuable resource is saved by incorporation into glycogen.

20. Free glucose must be phosphorylated at the expense of a molecule of ATP. Glucose 6-phosphate derived from glycogen is formed by phosphorolytic cleavage, thus sparing one molecule of ATP. Thus, the net yield of ATP when glycogen-derived glucose is processed to pyruvate is three molecules of ATP compared with two molecules of ATP from free glucose.

21. Breakdown: Phosphoglucomutase converts glucose 1-phosphate, liberated from glycogen breakdown, into glucose 6-phosphate, which can be released as free glucose (liver) or processed in glycolysis (muscle and liver). Synthesis: Converts glucose 6-phophosphate into glucose 1-phosphate, which reacts with UTP to form UDP-glucose, the substrate for glycogen synthase. 22. Glycogen_n + $P_i \longrightarrow glycoge_{n-1} + glucose$ 6-phosphate

Glucose 6-phosphate \longrightarrow glucose 1-phosphate

UTP + glucose 1-phosphate \longrightarrow UDP-glucose + 2 P_i

 $Glycogen_{n-1} + UDP$ -glucose \longrightarrow glycogen_n + UDP

Sum: Glycogen_n + UTP \longrightarrow glycogen_n + UDP + P_i

23. In principle, having glycogen be the only primer for the further synthesis of glycogen should be a successful strategy. However, if the glycogen granules were not evenly divided between daughter cells, glycogen stores for future generations of cells might be compromised. Glycogenin synthesizes the primer for glycogen synthase. 24. Insulin binds to its receptor and activates the tyrosine kinase activity of the receptor, which in turn triggers a pathway that activates protein kinases. The kinases phosphorylate and inactivate glycogen synthase kinase. Protein phosphatase 1 then removes the phosphate from glycogen synthase and thereby activates the synthase. 25.



α-1,6-Glucosidase reaction

A 2 6

ANSWERS TO PROBLEMS

26. Galactose + ATP + UTP + H_2O + $glycogen_n \rightarrow$

 $glycogen_{n+1} + ADP + UDP + 2P_i + H^+$

27. Phosphorylase, transferase, glucosidase, phosphoglucomutase, and glucose 6-phosphatase.

28. Glucose is an allosteric inhibitor of phosphorylase *a*. Hence, crystals grown in its presence are in the T state. The addition of glucose 1-phosphate, a substrate, shifts the R-to-T equilibrium toward the R state. The conformational differences between these states are sufficiently large that the crystal shatters unless it is stabilized by chemical cross-links.

29. Galactose is converted into UDP-galactose to eventually form glucose 6-phosphate.

30. This disease can also be produced by a mutation in the gene that encodes the glucose 6-phosphate transporter. Recall that glucose 6-phosphate must be transported into the lumen of the endoplasmic reticulum to be hydrolyzed by phosphatase. Mutations in the other three essential proteins of this system can likewise lead to von Gierke disease.

31. (a) Glycogen was too large to enter the gel and, because analysis was by western blot with the use of an antibody specific to glycogenin, we would not expect to see background proteins.

(b) $\alpha\text{-}Amylase$ degrades glycogen, releasing the protein glycogenin, which can be visualized by a western blot.

(c) Glycogen phosphorylase, glycogen synthase, and protein phosphatase 1. These proteins might be visible if the gel were stained for protein, but a western analysis reveals the presence of glycogenin only.

32. (a) The smear was due to molecules of glycogenin with increasingly large amounts of glycogen attached to them.(b) In the absence of glucose in the medium, glycogen is metabolized, resulting in a loss of the high-molecular-weight material.

(c) Glycogen could have been resynthesized and added to the glycogenin when the cells were fed glucose again.

(d) No difference between lanes 3 and 4 suggests that, by 1 hour, the glycogen molecules had attained maximum size in this cell line. Prolonged incubation does not apparently increase the amount of glycogen.

(e) $\alpha\text{-}Amylase$ removes essentially all of the glycogen, and so only the glycogenin remains.

Chapter 22

1. Glycerol + 2 NAD⁺ + P_i + ADP \longrightarrow pyruvate + ATP + H₂O + 2 NADH + H⁺

Glycerol kinase and glycerol phosphate dehydrogenase

2. The ready reversibility is due to the high-energy nature of the thioester in the acyl CoA.

3. To return the AMP to a form that can be phosphorylated by oxidative phosphorylation or substrate-level phosphorylation, another molecule of ATP must be expended in the reaction:

$$ATP + AMP \implies 2 ADP$$

4. b, c, a, g, h, d, e, f.

5. The citric acid cycle. The reactions that take succinate to oxaloacetate, or the reverse, are similar to those of fatty acid metabolism (Section 17.2).

6. The next-to-last degradation product, acetoacetyl CoA, yields two molecules of acetyl CoA with the thiolysis by only one molecule of CoA.

7. Palmitic acid yields 106 molecules of ATP. Palmitoleic acid has a double bond between carbons C-9 and C-10. When palmitoleic acid is processed in β oxidation, one of the oxidation steps

(to introduce a double bond before the addition of water) will not take place, because a double bond already exists. Thus, $FADH_2$ will not be generated, and palmitoleic acid will yield 1.5 fewer molecules of ATP than palmitic acid, for a total of 104.5 molecules of ATP.

Activation fee to form the acyl CoA	-2 ATP
Seven rounds of yield: 7 acetyl CoA at 10 ATP/acetyl CoA 7 NADH at 2.5 ATP/NADH	+ 70 ATP + 17.5 ATP
7 FADH ₂ at 1.5 ATP/FADH ₂ Propionyl CoA, which requires an ATP to be converted into succinyl CoA	+ 10.5 ATP - 1 ATP
Succinyl CoA \rightarrow succinate Succinate \rightarrow fumarate + FADH ₂ FADH ₂ at 1.5 ATP/FADH ₂ Fumarate \rightarrow malate	+ 1.5 ATP
Malate → oxaloacetate + NADH NADH at 2.5 ATP/NADH	+ 2.5 ATP
Total	120 ATP

9. You might hate yourself in the morning, but at least you won't have to worry about energy. To form stearoyl CoA requires the equivalent of 2 molecules of ATP.

Stearoyl CoA + 8 FAD + 8 NAD⁺ + 8 CoA + 8 H₂O \longrightarrow 9 acetyl CoA + 8 FADH₂ + 8 NADH + 8 H⁺

9 acetyl CoA at 10 ATP/acetyl CoA	+ 90 ATP
8 NADH at 2.5 ATP/NADH	+ 20 ATP
8 FADH_2 at 1.5 ATP/FADH $_2$	+ 12 ATP
Activation fee	-2.0
Total	122 ATP

10. Keep in mind that, in the citric acid cycle, 1 molecule of $FADH_2$ yields 1.5 ATP, 1 molecule of NADH yields 2.5 ATP, and 1 molecule of acetyl CoA yields 10 ATP. Two molecules of ATP are produced when glucose is degraded to 2 molecules of pyruvate. Two molecules of NADH also are produced, but the electrons are transferred to FADH₂ to enter the mitochondria. Each molecule of FADH₂ can generate 1.5 ATP. Each molecule of pyruvate will produce 1 molecule of NADH. Each molecule of acetyl CoA generates 3 molecules of NADH, 1 molecule of FADH₂, and 1 molecule of ATP. So, we have a total of 10 ATP per acetyl CoA, or 20 for the 2 molecules of acetyl CoA. The total for glucose is 30 ATP. Now, what about hexanoic acid? Caprioic acid is activated to caprioic CoA at the expense of 2 ATP, and so we are 2 ATP in the hole. The first cycle of β oxidation generates 1 FADH₂, 1 NADH, and 1 acetyl CoA. After the acetyl CoA has been run through the citric acid cycle, this step will have generated a total of 14 ATP. The second cycle of β oxidation generates 1 FADH₂ and 1 NADH but 2 acetyl CoA. After the acetyl CoA has been run through the citric acid cycle, this step will have generated a total of 24 ATP. The total is 36 ATP. Thus, the foul-smelling caprioic acid has a net yield of 36 ATP. So on a per carbon basis, this fat yields 20% more ATP than does glucose, a manifestation of the fact that fats are more reduced than carbohydrates.

11. Stearate + ATP + 13.5 H_2O + 8 FAD + 8 NAD⁺ \rightarrow 4.5 acetoacetate + 14.5 H⁺ + 8 FADH₂ + 8 NADH + AMP + 2 P_i. 12. Palmitate is activated and then processed by β oxidation according to the following reactions. Palmitate + CoA + ATP \longrightarrow palmitoyl CoA + AMP + 2 P_i

Palmitoyl CoA + 7 FAD + 7 NAD + 7 CoASH + $H_2O \longrightarrow$ 8 acetyl CoA + 7 FAD H_2 + 7 NADH + 7 H^+

The eight molecules of acetyl CoA combine to form four molecules of acetoacetate for release into the blood, and so they do not contribute to the energy yield in the liver. However, the $FADH_2$ and NADH generated in the preparation of acetyl CoA can be processed by oxidative phosphorylation to yield ATP.

$$1.5 \text{ ATP}/\text{FADH}_2 \times 7 = 10.5 \text{ ATP}$$

$$2.5 \text{ ATP/NADH} \times 7 = 17.5 \text{ ATP}$$

The equivalent of 2 ATP were used to form palmitoyl CoA. Thus, 26 ATP were generated for use by the liver.

13. NADH produced with the oxidation to acetoacetate = 2.5 ATP. Acetoacetate is converted into acetoacetyl CoA.

Two molecules of acetyl CoA result from the hydrolysis of acetoacetyl CoA, each worth 10 ATP when processed by the citric acid cycle. Total ATP yield is 22.5.

14. Because a molecule of succinyl CoA is used to form acetoacetyl CoA. Succinyl CoA could be used to generate one molecule of ATP (GTP), and so someone could argue that the yield is 21.5.

15. For fats to be combusted, not only must they be converted into acetyl CoA, but the acetyl CoA must be processed by the citric acid cycle. In order for acetyl CoA to enter the citric acid cycle, there must be a supply of oxaloacetate. Oxaloacetate can be formed by the metabolism of glucose to pyruvate and the subsequent carboxylation of pyruvate to form oxaloacetate. 16. (a)

The problem with phytanic acid is that, as it undergoes β oxidation, we encounter the dreaded pentavalent carbon atom. Because the pentavalent carbon atom doesn't exist, β oxidation cannot take place and phytanic acid accumulates.



(b) Removing methyl groups, though theoretically possible, would be time consuming and, lacking in elegance. What would we do with the methyl groups? Our livers solve the problem by inventing α oxidation.



One round of α oxidation rather than β oxidation converts phytanic acid into a β -oxidation substrate.

17. The first oxidation removes two tritium atoms. The hydration adds nonradioactive H and OH. The second oxidation removes another tritium atom from the β -carbon atom. Thiolysis removes an acetyl CoA with only one tritium atom; so the tritium-to-carbon ratio is 1/2. This ratio will be the same for two of the acetates. The last one, however, does not undergo oxidation, and so all tritium remains. The ratio for this acetate is 3/2. The ratio for the entire molecule is then 5/6.

18. In the absence of insulin, lipid mobilization will take place to an extent that it overwhelms the ability of the liver to convert the lipids into ketone bodies.

19. (a) Oxidation in mitochondria; synthesis in the cytoplasm.
(b) Coenzyme A in oxidation; acyl carrier protein for synthesis.
(c) FAD and NAD⁺ in oxidation; NADPH for synthesis. (d) the L isomer of 3-hydroxyacyl CoA in oxidation; the D isomer in synthesis.
(e) From carboxyl to methyl in oxidation; from methyl to carboxyl in synthesis. (f) The enzymes of fatty acid synthesis, but not those of

oxidation, are organized in a multienzyme complex.

20. 7 acetyl CoA + 6 ATP + 12 NADPH + 12 $\text{H}^+ \rightarrow$ myristate + 7 CoA + 6 ADP + 6 Pi + 12 NADP⁺ + 5H₂O.

21. We will need six acetyl CoA units. One acetyl CoA

unit will be used directly to become the two carbon atoms farthest from the acid end. The other five units must be converted into malonyl CoA. The synthesis of each malonyl CoA molecule costs a molecule of ATP; so 5 molecules of ATP will be required. Each round of elongation requires 2 molecules of NADPH, 1 molecule to reduce the keto group to an alcohol and 1 molecule to reduce the double bond. As a result, 10 molecules of NADPH will be required. Therefore, 5 molecules of ATP and 10 molecules of NADPH are required to synthesize lauric acid. 22. e, b, d, a, c.

23. Such a mutation would inhibit fatty acid synthesis because the enzyme cleaves cytoplasmic citrate to yield acetyl CoA for fatty acid synthesis.

24. (a) False. Biotin is required for acetyl CoA carboxylase activity. (b) True.

(c) False. ATP is required to synthesize malonyl CoA.

(d) True.

(e) True.

(f) False. Fatty acid synthase is a dimer.

(g) True.

(h) False. Acetyl CoA carboxylase is stimulated by citrate, which is cleaved to yield its substrate acetyl CoA.

25. Fatty acids with odd numbers of carbon atoms are synthesized starting with propionyl ACP (instead of acetyl ACP), which is formed from propionyl CoA by acetyl transacetylase.

26. All of the labeled carbon atoms will be retained. Because we need 8 acetyl CoA molecules and only 1 carbon atom is labeled in the acetyl group, we will have 8 labeled carbon atoms. The only acetyl CoA used

directly will retain 3 tritium atoms. The 7 acetyl CoA molecules used to make malonyl CoA will lose 1 tritium atom on addition of the CO_2 and another one at the dehydration step. Each of the 7 malonyl CoA molecules will retain 1 tritium atom. Therefore, the total retained tritium is 10 atoms. The ratio of tritium to carbon is 1.25.

27. With a diet rich in raw eggs, avidin will inhibit fatty acid synthesis by reducing the amount of biotin required by acetyl CoA carboxylase. Cooking the eggs will denature avidin, and so it will no longer bind biotin.

28. The only acetyl CoA used directly, not in the form of malonyl CoA, provides the two carbon atoms at the ω end of the fatty acid chain. Because palmitic acid is a C₁₆ fatty acid, acetyl CoA will have provided carbons 15 and 16.

29. HCO_3^- is attached to acetyl CoA to form malonyl CoA. When malonyl CoA condenses with acetyl CoA to form the four-carbon keto acyl CoA, the HCO_3^- is lost as CO_2 .

30. Phosphofructokinase controls the flux down the glycolytic pathway. Glycolysis functions to generate ATP or building blocks for biosynthesis, depending on the tissue. The presence of citrate in the cytoplasm indicates that those needs are met, and there is no need to metabolize glucose.

31. C-1 is more radioactive.

32. The mutant enzyme will be persistently active because it cannot be inhibited by phosphorylation. Fatty acid synthesis will be abnormally active. Such a mutation might lead to obesity.

33. (a) Palmitoleate; (b) linoleate; (c) linoleate; (d) oleate; (e) oleate; (f) linolenate.

34. Decarboxylation drives the condensation of malonyl ACP and acetyl ACP. In contrast, the condensation of two molecules of acetyl ACP is energetically unfavorable. In gluconeogenesis, decarboxylation drives the formation of phosphoenolpyruvate from oxaloacetate.

35. Fat mobilization in adipocytes is activated by phosphorylation. Hence, overproduction of the cAMP-activated kinase will lead to an accelerated breakdown of triacylglycerols and a depletion of fat stores.

36. Carnitine translocase deficiency and glucose 6-phosphate transporter deficiency.

37. In the fifth round of β oxidation, $cis - \Delta^2$ -enoyl CoA is formed. Dehydration by the classic hydratase yields D-3-hydroxyacyl CoA, the wrong isomer for the next enzyme in β oxidation. This dead end is circumvented by a second hydratase that removes water to give *trans*- Δ^2 -enoyl CoA. The addition of water by the classic hydratase then yields L-3-hydroxyacyl CoA, the appropriate isomer. Thus, hydratases of opposite stereospecificities serve to *epimerize* (invert the configuration of) the 3-hydroxyl group of the acyl CoA intermediate.

38. The probability of synthesizing an error-free polypeptide chain decreases as the length of the chain increases. A single mistake can make the entire polypeptide ineffective. In contrast, a defective subunit can be spurned in the formation of a noncovalent multienzyme complex; the good subunits are not wasted.

39. The absence of ketone bodies is due to the fact that the liver, the source of ketone bodies in the blood, cannot oxidize fatty acids to produce acetyl CoA. Moreover, because of the impaired fatty acid oxidation, the liver becomes more dependent on glucose as an energy source. This dependency results in a decrease in gluconeogenesis and a drop in blood-glucose levels, which is exacerbated by the lack of fatty acid oxidation in muscle and a subsequent increase in glucose uptake from the blood.

40. Peroxisomes enhance the degradation of fatty acids. Consequently, increasing the activity of peroxisomes could help to lower levels of blood triglycerides. In fact, clofibrate is rarely used because of serious side effects.

41. Citrate works by facilitating the formation of active filaments from inactive monomers. In essence, it increases the number of active sites available, or the concentration of enzyme. Consequently, its effect is visible as an increase in the value of $V_{\rm max}$. Allosteric enzymes that alter their $V_{\rm max}$ values in response to regulators are sometimes called V-class enzymes. The more common type of allosteric enzyme, in which $K_{\rm m}$ is altered, comprises K-class enzymes. Palmitoyl CoA causes depolymerization and thus inactivation.

42. The thiolate anion of CoA attacks the 3-keto group to form a tetrahedral intermediate. This intermediate collapses to form acyl CoA and the enolate anion of acetyl CoA. Protonation of the enolate yields acetyl CoA.

43.

$$ACP-S \xrightarrow{(0)}_{H_2} \xrightarrow{(0)}_{C \to 0} \xrightarrow{(0)}_{C \to 0} \xrightarrow{(0)}_{C \to 0}$$

Malonyl-ACP



Acetoacetyl-ACP

44. (a) Fats burn in the flame of carbohydrates. Without carbohydrates, there would be no anapleurotic reactions to replenish the components of the citric acid cycle. With a diet of fats only, the acetyl CoA from fatty acid degradation would build up. (b) Acetone from ketone bodies

(c) Yes. Odd-chain fatty acids would lead to the production of propionyl CoA, which can be converted into succinyl CoA, a citric acid cycle component. It would serve to replenish the citric acid cycle and mitigate the halitosis.

45. A labeled fat can enter the citric acid cycle as acetyl CoA and yield labeled oxaloacetate, but only after two carbon atoms have been lost as CO_2 . Consequently, even though oxaloacetate may be labeled, there can be no net synthesis of oxaloacetate and hence no net synthesis of glucose or glycogen.

46. (a) The V_{max} is decreased and the K_{m} is increased. V_{max} (wild type) = 13 nmol minute⁻¹ mg⁻¹; K_{m} (wild type) = 45 μ M; V_{max} (mutant) = 8.3 nmol minute⁻¹ mg⁻¹; K_{m} (mutant) = 74 μ M. (b) Both the V_{max} and the K_{m} are decreased. V_{max} (wild type) = 41 nmol minute⁻¹ mg⁻¹; K_{m} (wild type) = 104 μ M; V_{max} (mutant) = 23 nmol minute⁻¹ mg⁻¹; K_{m} (mutant) = 69 μ M. (c) The wild type is significantly more sensitive to malonyl CoA. (d) With respect to carnitine, the mutant displays approximately 65% of the activity of the wild type; with respect to palmitoyl CoA, approximately 50% activity. On the other hand, 10 μ M of malonyl CoA inhibits approximately 80% of the wild type but has essentially no effect on the mutant enzyme. (e) The glutamate appears to play a more prominent role in regulation by malonyl CoA than in catalysis.

Chapter 23

1. When the proteins are denatured, all of the peptide bonds are accessible to proteolytic enzymes. If the three-dimensional structure of a protein is maintained, access to many peptide bonds is denied to the proteolytic enzymes.

2. First, the ubiquitin-activating enzyme (E1) links ubiquitin to a sulfhydryl group on E1 itself. Next, the ubiquitin is transferred to a cysteine residue on the ubiquitin-conjugating enzyme (E2) by E2. The ubiquitin-protein ligase (E3), using the ubiquitinated E2 as a substrate, transfers the ubiquitin to the target protein.

3. (a) 7; (b) 4; (c) 2; (d) 10; (e) 5; (f) 3; (g) 9; (h) 1; (i) 6; (j) 8.
4. (a) The ATPase activity of the 26S proteasome resides in the 19S subunit. The energy of ATP hydrolysis could be used to unfold the substrate, which is too large to enter the catalytic barrel. ATP may also be required for translocation of the substrate into the barrel.

(b) Substantiates the answer in part *a*. Because they are small, the peptides do not need to be unfolded. Moreover, small peptides could probably enter all at once and not require translocation.
5. (a) Pyruvate; (b) oxaloacetate; (c) α-ketoglutarate; (d) α-ketoi-

socaproate; (e) phenylpyruvate; (f) hydroxyphenylpyruvate. 6. (a) Aspartate + α -ketoglutarate + GTP + ATP + 2 H₂O + NADH + H⁺ $\rightarrow \frac{1}{2}$ glucose + glutamate + CO₂ + ADP + GDP + NAD⁺ + 2 P_i.

The required coenzymes are pyridoxal phosphate in the transamination reaction and NAD⁺/NADH in the redox reactions. (b) Aspartate + CO_2 + NH_4^+ + 3 ATP + NAD^+ + 4 $H_2O \rightarrow$ oxaloacetate + urea + 2 ADP + 4 P_i + AMP + NADH + H^+ . 7. In the eukaryotic proteasome, the distinct β subunits have different substrate specificities, allowing proteins to be more thoroughly degraded.

8. The six subunits probably exist as a heterohexamer. Crosslinking experiments could test the model and help determine which subunits are adjacent to one another.

9. Thiamine pyrophosphate

10. Aminotransferases transfer the α -amino group to α -ketoglutarate to form glutamate. Glutamate is oxidatively deaminated to form an ammonium ion.

11. Aspartate (oxaloacetate), glutamate (α -ketoglutarate), alanine (pyruvate).

12. Serine and threonine

13. They are either fuels for the citric acid cycle, components of the citric acid cycle, or molecules that can be converted into a fuel for the citric acid cycle in one step.

14. It acts as an electron sink.

15. Carbamoyl phosphate and aspartate

16. (a) 4; (b) 5; (c) 1; (d) 6; (e) 7; (f) 3; (g) 2.

17. A, arginine; B, citrulline; C, ornithine; D, arginosuccinate. The order of appearance: C, B, D, E.

18. $CO_2 + NH_4^+ + 3 ATP + NAD^+ + aspartate + 3 H_2O \rightarrow$ urea + 2 ADP + 2 P_i + AMP + PP_i + NADH + H⁺ + oxaloacetate.

Four high-transfer-potential phosphoryl groups are spent. Note, however, that an NADH is generated if fumarate is converted into oxaloacetate. NADH can generate 2.5 ATP in the electron-transport chain. Taking these ATP into account, only 1.5 high-transferpotential phosphoryl groups are spent.

19. The synthesis of fumarate by the urea cycle is important because it links the urea cycle and the citric acid cycle. Fumarate is hydrated

to malate, which, in turn, is oxidized to oxaloacetate. Oxaloacetate has several possible fates: (1) transamination to aspartate, (2) conversion into glucose by the gluconeogenic pathway, (3) condensation with acetyl CoA to form citrate, or (4) conversion into pyruvate. You can collect.

20. Ornithine transcarbamoylase (analogous to PALA; see Chapter 10).

21. Ammonia could lead to the amination of α -ketoglutarate, producing a high concentration of glutamate in an unregulated fashion. α -Ketoglutarate for glutamate synthesis could be removed from the citric acid cycle, thereby diminishing the cell's respiration capacity.

22. The mass spectrometric analysis strongly suggests that three enzymes—pyruvate dehydrogenase, α -ketoglutarate dehydrogenase, and the branched-chain α -ketoacid dehydrogenase are deficient. Most likely, the common E₃ component of these enzymes is missing or defective. This proposal could be tested by purifying these three enzymes and assaying their ability to catalyze the regeneration of lipoamide.

23. Benzoate, phenylacetate, and arginine would be given to supply a protein-restricted diet. Nitrogen would emerge in hippurate, phenylacetylglutamine, and citrulline.

24. The liver is the primary tissue for capturing nitrogen as urea. If the liver is damaged (for instance, by hepatitis or the excessive consumption of alcohol), free ammonia is released into the blood.

25. This defect can be partly bypassed by providing a surplus of arginine in the diet and restricting the total protein intake. In the liver, arginine is split into urea and ornithine, which then reacts with carbamoyl phosphate to form citrulline. This urea-cycle intermediate condenses with aspartate to yield argininosuccinate, which is then excreted. Note that two nitrogen atoms—one from carbamoyl phosphate and the other from aspartate—are eliminated from the body per molecule of arginine provided in the diet. In essence, argininosuccinate substitutes for urea in carrying nitrogen out of the body. The formation of argininosuccinate removes the nitrogen, and the restriction on protein intake relieves the aciduria.

26. Aspartame, a dipeptide ester (L-aspartyl-L-phenylalanine methyl ester), is hydrolyzed to L-aspartate and L-phenylalanine. High levels of phenylalanine are harmful in phenylketonurics. 27. *N*-Acetylglutamate is synthesized from acetyl CoA and glutamate. Once again, acetyl CoA serves as an activated acetyl donor. This reaction is catalyzed by *N*-acetylglutamate synthase.

28. Not all proteins are created equal: some are more important than others. Some proteins would be degraded to provide the missing amino acid. The nitrogen from the other amino acids would be excreted as urea. Consequently, more nitrogen would be excreted than ingested.

29. The carbon skeletons of ketogenic amino acids can be converted into ketone bodies or fatty acids. Only leucine and lysine are purely ketogenic. Glucogenic amino acids are those whose carbon skeletons can be converted into glucose.

30. The branched-chain amino acids leucine, isoleucine, and valine. The required enzyme is the branched-chain α -ketoacid dehydrogenase complex.

31. Pyruvate (glycolysis and gluconeogenesis), acetyl CoA (citric acid cycle and fatty acid synthesis), acetoacetyl CoA (ketone-body formation), α -ketoglutarate (citric acid cycle), succinyl CoA (citric acid cycle), fumarate (citric acid cycle), and oxaloacetate (citric acid cycle and gluconeogenesis).

A 3 0

ANSWERS TO PROBLEMS





The equilibrium constant for the interconversion of L-serine and D-serine is exactly 1.

34. Exposure of such a domain suggests that a component of a multiprotein complex has failed to form properly or that one component has been synthesized in excess. This exposure leads to rapid degradation and the restoration of appropriate stoichiometries.

35. (a) Depletion of glycogen stores. When they are gone, proteins must be degraded to meet the glucose needs of the brain. The resulting amino acids are deaminated, and the nitrogen atoms are excreted as urea.

(b) The brain has adapted to the use of ketone bodies, which are derived from fatty acid catabolism. In other words, the brain is being powered by fatty acid breakdown.(c) When the glycogen and lipid stores are gone, the only available energy source is protein.

36. Deamination to α -keto- β -methylvalerate; oxidative decarboxylation to α -methylbutyryl CoA; oxidation to tiglyl CoA; hydration, oxidation, and thiolysis yield acetyl CoA and propionyl CoA; propionyl CoA to succinyl CoA. 37. Glycogen phosphorylase. The coenzyme serves as an acid–base catalyst.

38. In the Cori cycle, the carbon atoms are transferred from muscle to liver as lactate. For lactate to be of any use, it must be reduced to pyruvate. This reduction requires high-energy electrons in the form of NADH. When the carbon atoms are transferred as alanine, transamination yields pyruvate directly.

39. (a) Virtually no digestion in the absence of nucleotides. (b) Protein digestion is greatly stimulated by the presence of ATP. (c) AMP-PNP, a nonhydrolyzable analog of ATP, is no more effective than ADP. (d) The proteasome requires neither ATP nor PAN to digest small substrates. (e) PAN and ATP hydrolysis may be required to unfold the peptide and translocate it into the proteasome. (f) Although *Thermoplasma* PAN is not as effective with the other proteasomes, it nonetheless results in threefold to fourfold stimulation of digestion. (g) In light of the fact that the archaea and eukarya diverged several billion years ago, the fact that

Thermoplasma PAN can stimulate rabbit muscle suggests homology not only between the proteasomes, but also between PAN and the 19S subunit (most likely the ATPases) of the mammalian 26S proteasome.

Chapter 24

1. Nitrogen fixation is the conversion of atmospheric N2 into NH₃⁺. Diazotrophic (nitrogen-fixing) microorganisms are able to fix nitrogen.

2. Oxaloacetate, pyruvate, ribose-5-phosphate, phosphoenolpyruvate, erythrose-4-phosphate, α -ketoglutarate, and 3-phosphoglycerate.

3. Human beings do not have the biochemical pathways to synthesize certain amino acids from simpler precursors. Consequently, these amino acids are "essential" and must be obtained from the diet.

4. Glucose + 2 ADP + 2 P_i + 2 NAD⁺ + 2 glutamate \rightarrow 2 alanine + 2 α -ketoglutrate + 2 ATP + 2 NADH +

 $2 H_2O + 2 H^+.$ 5. N₂ \rightarrow NH₄⁺ \rightarrow glutamate \rightarrow serine \rightarrow glycine \rightarrow δ -aminolevulinate \rightarrow porphobilinogen \rightarrow heme.

6. False. Nitrogen fixation is thermodynamically favorable. Nitrogenase is required because the process is kinetically disfavored

7. Pyridoxal phosphate (PLP)

8. S-Adenosylmethionine, tetrahydrofolate, and methylcobalamin.

9. (a) N^5 , N^{10} -Methylenetetrahydrofolate;

(b) N^5 -methyltetrahydrofolate.

10. γ -Glutamyl phosphate is a likely reaction intermediate.

11. The synthesis of asparagine from aspartate passes through an acyl-adenylate intermediate. One of the products of the reaction will be ¹⁸O-labeled AMP.

12. The administration of glycine leads to the formation of isovalerylglycine. This water-soluble conjugate, in contrast with isovaleric acid, is excreted very rapidly by the kidneys.

13. The nitrogen atom shaded red is derived from glutamine. The carbon atom shaded blue is derived from serine.

HN

14. They carry out nitrogen fixation. The absence of photosystem II provides an environment in which O₂ is not produced. Recall that the nitrogenase is very rapidly inactivated by O₂.

15. The cytoplasm is a reducing environment, whereas the extracellular milieu is an oxidizing environment.

16. (a) None; (b) D-glutamate and oxaloacetate. 17. Succinvl CoA is formed in the mitochondrial matrix.

18. Alanine from pyruvate; aspartate from oxaloacetate; glutamate from α -ketoglutarate.

19. Lysine cyclodeaminase converts L-lysine into the six-membered ring analog of proline, also referred to as L-homoproline or L-pipecolate:



20. Y could inhibit the C \rightarrow D step, Z could inhibit the C \rightarrow F step, and C could inhibit $A \rightarrow B$. This scheme is an example of sequential feedback inhibition. Alternatively, Y could inhibit the $C \rightarrow D$ step, Z could inhibit the $C \rightarrow F$ step, and the $A \rightarrow B$

step would be inhibited only in the presence of both Y and Z. This scheme is called concerted feedback inhibition.

21. The rate of the A \rightarrow B step in the presence of high levels of Y and Z would be 24 s⁻¹ (0.6 × 0.4 × 100 s⁻¹).

22. Lysine 258 is absolutely essential for the activity of aspartate aminotransferase, as it is responsible both for the formation of the internal aldimine with the pyridoxal phosphate cofactor and for transferring the proton between the ketimine and quinonoid intermediates. Mutation of this residue to cysteine would be expected to dramatically impair catalysis, as cysteine cannot occupy the same space as lysine and also exhibits differing pK_a properties. Upon treatment with 2-bromoethylamine, however, the resulting thioether now has a shape and pK_a similar to the original lysine side chain. Hence, some catalytic activity is restored.

23. An external aldimine forms with SAM, which is deprotonated to form the quinonoid intermediate. The deprotonated carbon atom attacks the carbon atom adjacent to the sulfur atom to form the cyclopropane ring and release methylthioadenosine, the other product. 24. An external aldimine forms with L-serine, which is deprotonated to form the quinonoid intermediate. This intermediate is reprotonated on its opposite face to form an aldimine with D-serine. This compound is cleaved to release D-serine. The equilibrium constant

for a racemization reaction is 1 because the reactant and product are exact mirror images of each other.

25. (a) In the first step, histidine attacks the methylene group from the methionine subgroup of SAM (rather than the usual methyl substituent), resulting in the transfer of an aminocarboxypropyl group. Three subsequent conventional SAM-mediated methylations of the primary amine yield diphthine.



(b) In this chapter, we have observed two examples of an ATPdependent conversion of a carboxylate into an amide: glutamine synthetase, which uses an acyl-phosphate intermediate, and asparagine synthetase, which uses an acyl-adenylate intermediate. Either mechanism is possible in formation of diphthamide from diphthine. 26. Synthesis from oxaloacetate and α -ketoglutarate would deplete the citric acid cycle, which would decrease ATP production. Anapleurotic reactions would be required to replenish the citric acid cycle.

27. SAM is the donor for DNA methylation reactions that protect a host from digestion by its own restriction enzymes. A lack of SAM would render the bacterial DNA susceptible to digestion by the cell's own restriction enzymes.

28. Acetate \rightarrow acetyl-CoA \rightarrow citrate \rightarrow isocitrate \rightarrow α -ketoglutarate \rightarrow succinyl-CoA.

29. (a) Asparagine is much more abundant in the dark. More glutamine is present in the light. These amino acids show the most dramatic effects. Glycine also is more abundant in the light.

(b) Glutamine is a more metabolically reactive amino acid, used in the synthesis of many other compounds. Consequently, when energy is available as light, glutamine will be preferentially synthesized. Asparagine, which carries more nitrogen per carbon atom and is thus a more-efficient means of storing nitrogen when energy is short, is synthesized in the dark. Glycine is more prevalent in the light because of photorespiration.

(c) White asparagus has an especially high concentration of asparagine, which accounts for its intense taste. All asparagus has a large amount of asparagine. In fact, as suggested by its name, asparagine was first isolated from asparagus.

Chapter 25

1. In de novo synthesis, the nucleotides are synthesized from simpler precursor compounds, in essence from scratch. In salvage pathways, preformed bases are recovered and attached to riboses.

 Carbon 2 and nitrogen 3 come from carbamoyl phosphate. Nitrogen 1 and carbons 4, 5, and 6 are derived from aspartate.
 Nitrogen 1: aspartate; carbon 2: N¹⁰-formyltetrahydrofolate; nitrogen 3: glutamine; carbons 4 and 5 and nitrogen 7: glycine; carbon 6: CO₂; carbon 8: N¹⁰-formyltetrahydrofolate; nitrogen 9: glutamine.

4. Energy currency: ATP; signal transduction: ATP and GTP; RNA synthesis: ATP, GTP, CTP, and UTP; DNA synthesis: dATP, dCTP, dGTP, and TTP; components of coenzymes: ATP in CoA, FAD, and NAD(P)⁺; carbohydrate synthesis: UDP-glucose. They are just some of the uses.

5. A nucleoside is a base attached to ribose. A nucleotide is a nucleoside with the ribose bearing one or more phosphates.
6. (a) 9; (b) 7; (c) 6; (d) 10; (e) 2; (f) 4; (g) 1; (h) 11; (i) 8; (j) 3; (k) 5.
7. Substrate channeling is the process whereby the product of one active site moves to become a substrate at another active site without ever leaving the enzyme. A channel connects the active

sites. Substrate channeling greatly enhances enzyme efficiency and minimizes the diffusion of a substrate to an active site. 8. Glucose + 2 ATP + 2 NADP⁺ + H₂O \rightarrow PRPP + CO₂ +

8. Glucose + 2 ATP + 2 NADP + $H_2O \rightarrow PRPP + CO$ ADP + AMP + 2 NADPH + 3 H⁺.

9. Glutamine + aspartate + CO_2 + 2 ATP + $NAD^+ \rightarrow orotate$ + 2 ADP + 2 P_i + glutamate + NADH + H^+ .

10. (a, c, and d) PRPP; (b) carbamoyl phosphate.

11. PRPP and formylglycinamide ribonucleotide

12. dUMP + serine + NADPH + $H^+ \rightarrow dTMP$ + NADP⁺ + glycine.

13. There is a deficiency of N^{10} -formyltetrahydrofolate. Sulfanilamide inhibits the synthesis of folate by acting as an analog of *p*-aminobenzoate, one of the precursors of folate.

14. (a) Cell A cannot grow in a HAT medium, because it cannot synthesize TMP either from thymidine or from dUMP. Cell B cannot grow in this medium, because it cannot synthesize purines by either the de novo pathway or the salvage pathway. Cell C can grow in a HAT medium because it contains active thymidine kinase from cell B (enabling it to phosphorylate thymidine to TMP) and hypoxanthine guanine phosphoribosyltransferase from cell A (enabling it to synthesize purines from hypoxanthine by the salvage pathway).

(b) Transform cell A with a plasmid containing foreign genes of interest and a functional thymidine kinase gene. The only cells that will grow in a HAT medium are those that have acquired a thymidylate kinase gene; nearly all of these transformed cells will also contain the other genes on the plasmid.

15. The reciprocal substrate relation refers to the fact that AMP synthesis requires GTP, whereas GMP synthesis requires ATP. These requirements tend to balance the synthesis of ATP and GTP.

16. Ring carbon 6 in cytosine will be labeled. In guanine, only carbon 5 will be labeled with 13 C.

17. The enzyme that uses ammonia synthesizes carbamoyl phosphate for a reaction with ornithine, the first step of the urea cycle. The enzyme that uses glutamine synthesizes carbamoyl phosphate for use in the first step of pyrimidine biosynthesis.

18. These patients have a high level of urate because of the breakdown of nucleic acids. Allopurinol prevents the formation of kidney stones and blocks other deleterious consequences of hyperuricemia by preventing the formation of urate.

19. The free energies of binding are -57.7 (wild type),

-49.8 (Asn 27), and -38.1 (Ser 27) kJ mol⁻¹ (-13.8, -11.9, and -9.1 kcal mol⁻¹, respectively). The loss in binding energy is 7.9 kJ mol⁻¹ (1.9 kcal mol⁻¹) and 19.7 kJ mol⁻¹ (4.7 kcal mol⁻¹). 20. Inosine or hypoxanthine could be administered.

21. N-1 in both cases, and the amine group linked to C-6 in ATP.

22. Nitrogen atoms 3 and 9 in the purine ring

23. Allopurinol, an analog of hypoxanthine, is a suicide inhibitor of xanthine oxidase.

24. An oxygen atom is added to allopurinol to form alloxanthine. 25.

The synthesis of carbamoyl phosphate requires 2 ATP	2 ATP
The formation of PRPP from ribose 5-phosphate	
yields an AMP*	2 ATP
The conversion of UMP to UTP requires 2 ATP	2 ATP
The conversion of UTP to CTP requires 1 ATP	1 ATP
Total	7 ATP

*Remember that AMP is the equivalent of 2 ATP because an ATP must be expended to generate ADP, the substrate for ATP synthesis.

26. (a) Carboxyaminoimidazole ribonucleotide; (b) glycinamide ribonucleotide; (c) phosphoribosyl amine; (d) formylglycinamide ribonucleotide.

27. The first reaction proceeds by phosphorylation of glycine to form an acyl phosphate followed by nucleophilic attack by the amine of phosphoribosylamine to displace orthophosphate. The second reaction consists of adenylation of the carbonyl group of xanthylate followed by nucleophilic attack by ammonia to displace AMP. 28. The $-NH_2$ group attacks the carbonyl carbon atom to form a tetrahedral intermediate. Removal of a proton leads to the elimination of water to form inosinate.

29. PRPP is the activated intermediate in the synthesis of phosphoribosylamine in the de novo pathway of purine formation; of purine nucleotides from free bases by the salvage pathway; of orotidylate in the formation of pyrimidines; of nicotinate ribonucleotide; of phosphoribosyl ATP in the pathway leading to histidine; and of phosphoribosylanthranilate in the pathway leading to tryptophan.

30. (a) cAMP; (b) ATP; (c) UDP-glucose; (d) acetyl CoA;
(e) NAD⁺, FAD; (f) dideoxynucleotides; (g) fluorouracil; (h) CTP inhibits ATCase.

31. In vitamin B_{12} deficiency, methyltetrahydrofolate cannot donate its methyl group to homocysteine to regenerate methionine. Because the synthesis of methyltetrahydrofolate is irreversible, the cell's tetrahydrofolate will ultimately be converted into this form. No formyl or methylene tetrahydrofolate will be left for nucleotide synthesis. Vitamin B_{12} is also required to metabolize propionyl CoA generated in the oxidation of odd-chain fatty acids and in the degradation of methionine.

32. Because folate is required for nucleotide synthesis, cells that are dividing rapidly would be most readily affected. They would include cells of the intestine, which are constantly replaced, and precursors to blood cells. A lack of intestinal cells and blood cells would account for the symptoms often observed.

33. The cytoplasmic level of ATP in the liver falls and that of AMP rises above normal in all three conditions. The excess AMP is degraded to urate.

34. Succinate \rightarrow malate \rightarrow oxaloacetate by the citric acid cycle. Oxaloacetate \rightarrow aspartate by transamination, followed by pyrimidine synthesis. Carbons 4, 5, and 6 are labeled.

35. Glucose will most likely be converted into two molecules of pyruvate, one of which will be labeled in the 2 position:



Now consider two common fates of pyruvate—conversion into acetyl CoA and subsequent processing by the citric acid cycle or carboxylation by pyruvate carboxylase to form oxaloacetate. Formation of citrate by condensing the labeled pyruvate with oxaloacetate will yield labeled citrate:



The labeled carbon will be retained through one round of the citric acid cycle but, on the formation of the symmetric succinate, the label will appear in two different positions. Thus, when succinate

is metabolized to oxaloacetate, which may be aminated to form aspartate, two carbons will be labeled:



When this aspartate is used to form uracil, the labeled COO^- attached to the α -carbon is lost and the other COO^- becomes incorporated into uracil as carbon 4.

Suppose, instead, that labeled $2 \cdot [{}^{14}C]$ pyruvate is carboxylated to form oxaloacetate and processed to form aspartate. In this case, the α -carbon of aspartate bears the label.



When this aspartate is used to synthesize uracil, carbon 6 bears the label.

36. (a) Some ATP can be salvaged from the ADP that is being generated. (b) There are equal numbers of high-phosphoryl-transferpotential groups on each side of the equation. (c) Because the adenylate kinase reaction is at equilibrium, the removal of AMP would lead to the formation of more ATP. (d) Essentially, the cycle serves as an anaplerotic reaction for the generation of the citric acid cycle intermediate fumarate.

37. (i) The formation of 5-aminoididazole-4-carboxamide ribonucleotide from 5-aminoimidazole-4-(*N*-succinylcarboxamide) ribonucleotide in the synthesis of IMP. (ii) The formation of AMP from adenylosuccinate. (iii) The formation of arginine from argininosuccinate in the urea cycle.

38. Allopurinol is an inhibitor of xanthine oxidase, which is on the pathway for urate synthesis. In your pet duck, this pathway is the means by which excess nitrogen is excreted. If xanthine oxidase were inhibited in your duck, nitrogen could not be excreted, with severe consequences such as the formation of a dead duck.

Chapter 26

1. Glycerol 3-phosphate is the foundation for both triacylglycerol and phospholipid synthesis. Glycerol 3-phosphate is acylated twice to form phosphatidate. In triacylglycerol synthesis, the phosphoryl group is removed from glycerol 3-phosphate to form diacylglycerol, which is then acylated to form triacylglycerol. In phospholipid synthesis, phosphatidate commonly reacts with CTP to form CDP-diacylglycerol, which then reacts with an alcohol to form a phospholipid. Alternatively, diacylglycerol may react with a CDP-alcohol to form a phospholipid.

2. Glycerol 3-phosphate is formed primarily by the reduction of dihydroxyacetone phosphate, a gluconeogenic intermediate, and to a lesser extent by the phosphorylation of glycerol.

3. Glycerol + 4 ATP + 3 fatty acids + 4 $H_2O \rightarrow$

triacylglycerol + ADP + 3 AMP + 7 P_i + 4 H^+ .

4. Glycerol + 3 ATP + 2 fatty acids + 2 H_2O + CTP +

ethanolamine \rightarrow phosphatidylethanolamine + CMP + ADP + 2 AMP + 6 P_i + 3 H⁺.

 Three. One molecule of ATP to form phosphorylethanolamine and two molecules of ATP to regenerate CTP from CMP.
 All are synthesized from ceramide. In sphingomyelin, the terminal hydroxyl group of ceramide is modified with phosphorylcholine. In a cerebroside, the hydroxyl group has a glucose or galactose attached. In a ganglioside, oligosaccharide chains are attached to the hydroxyl group.

7. (i) Activate the diacylglycerol as CDP-DAG. (ii) Activate the alcohol as CDP-alcohol. (iii) Use the base-exchange reaction.
8. (a) CDP-diacylglycerol; (b) CDP-ethanolamine; (c) acyl CoA; (d) phosphatidylcholine; (e) UDP-glucose or UDP-galactose; (f) UDP-galactose; (g) geranyl pyrophosphate.

9. Such mutations are seen in mice. The amount of adipose tissue would decrease severely because diacylglycerol could not be formed. Normally, diacylglycerol is acylated to form triacylglycerols. If there were deficient phosphatidic acid phosphatase activity, no triacylglycerols would form.

10. (i) The synthesis of activated isoprene units (isopentyl pyrophosphate), (ii) the condensation of six of the activated isoprene units to form squalene, and (iii) cyclization of the squalene to form cholesterol. 11. The amount of reductase and its activity control the regulation of cholesterol biosynthesis. Transcriptional control is mediated by SREBP. Translation of the reductase mRNA also is controlled. The reductase itself may undergo regulated proteolysis. Finally, the activity of the reductase is inhibited by phosphorylation by AMP kinase when ATP levels are low.

12. (a and b) None, because the label is lost as CO_2 .

13. The hallmark of this genetic disease is elevated cholesterol levels in the blood of even young children. The excess cholesterol is taken up by marcrophages, which eventually results in the formation of plaques and heart disease. There are many mutations that cause the disease, but all result in malfunctioning of the LDL receptor.

14. The categories of mutations are: (i) no receptor is synthesized; (ii) receptors are synthesized but do not reach the plasma membrane, because they lack signals for intracellular transport or do not fold properly; (iii) receptors reach the cell surface, but they fail to bind LDL normally because of a defect in the LDLbinding domain; (iv) receptors reach the cell surface and bind LDL, but they fail to cluster in coated pits because of a defect in their carboxyl-terminal regions.

15. "None of your business" and "I don't talk biochemistry until after breakfast" are appropriate but rude and uninformative answers. A better answer might be: "Although it is true that cholesterol is a precursor to steroid hormones, the rest of the statement is oversimplified. Cholesterol is a component of membranes, and membranes literally define cells, and cells make up tissues. But to say that cholesterol 'makes' cells and tissues is wrong."

16. Statins are competitive inhibitors of HMG-CoA reductase. They are used as drugs to inhibit cholesterol synthesis in patients with high levels of cholesterol.

17. No. Cholesterol is essential for membrane function and as a precursor for bile salts and steroid hormones. The complete lack of cholesterol would be lethal.

18. Deamination of cytidine to uridine changes CAA (Gln) into UAA (stop).

19. The LDL contains apolipoprotein B-100, which binds to an LDL receptor on the cell surface in a region known as a coated pit. On binding, the complex is internalized by endocytosis to

form an internal vesicle. The vesicle is separated into two components. One, with the receptor, is transported back to the cell surface and fuses with the membrane, allowing continued use of the receptor. The other vesicle fuses with lysosomes inside the cell. The cholesteryl esters are hydrolyzed, and free cholesterol is made available for cellular use. The LDL protein is hydrolyzed to free amino acids.

20. Benign prostatic hypertrophy can be treated by inhibiting 5α -reductase. Finasteride, the 4-azasteroid analog of dihydrotes-tosterone, competitively inhibits the reductase but does not act on androgen receptors. Patients taking finasteride have a markedly lower plasma level of dihydrotestosterone and a nearly normal level of testosterone. The prostate gland becomes smaller, whereas testosterone-dependent processes such as fertility, libido, and muscle strength appear to be unaffected.



21. Patients who are most sensitive to debrisoquine have a deficiency of a liver P450 enzyme encoded by a member of the CYP2 subfamily. This characteristic is inherited as an autosomal recessive trait. The capacity to degrade other drugs may be impaired in people who hydroxylate debrisoquine at a slow rate, because a single P450 enzyme usually handles a broad range of substrates.

22. Many hydrophobic odorants are deactivated by hydroxylation. Molecular oxygen is activated by a cytochrome P450 monooxygenase. NADPH serves as the reductant. One oxygen atom of O_2 goes into the odorant substrate, whereas the other is reduced to water.

23. Recall that dihydrotestosterone is crucial for the development of male characteristics in the embryo. If a pregnant woman were to be exposed to Propecia, the 5α -reductase of the male embryo would be inhibited, which could result in severe developmental abnormalities.

24. The oxygenation reactions catalyzed by the cytochrome P450 family permit greater flexibility in biosynthesis. Because plants are not mobile, they must rely on physical defenses, such as thorns, and chemical defenses, such as toxic alkaloids. The larger P450 array might permit greater biosynthetic versatility.

25. This knowledge would enable clinicians to characterize the likelihood of a patient's having an adverse drug reaction or being susceptible to chemical-induced illnesses. It would also permit a personalized and especially effective drug-treatment regime for diseases such as cancer.

26. The honey bees may be especially sensitive to environmental toxins, including pesticides, because these chemicals are not readily detoxified, owing to the minimal P450 system.

27. The core structure of a steroid is four fused rings: three cyclohexane rings and one cyclopentane ring. In vitamin D, the B ring is split by ultraviolet light.

28. The negatively charged phosphoserine residue interacts with the positively charged protonated histidine residue and decreases its ability to transfer a proton to the thiolate.



29. The methyl group is first hydroxylated. The hydroxymethylamine eliminated formaldehyde to form methylamine. 30. Note that a cytidine nucleotide plays the same role in the synthesis of these phosphoglycerides as a uridine nucleotide does in the formation of glycogen (Section 21.4). In all of these biosyntheses, an activated intermediate (UDP-glucose, CDP-diacylglycerol, or CDP-alcohol) is formed from a phosphorylated substrate (glucose 1-phosphate, phosphatidate, or a phosphorylalcohol) and a nucleoside triphosphate (UTP or CTP). The activated intermediate then reacts with a hydroxyl group (the terminus of glycogen, the side chain of serine, or a diacylglycerol).

31. The attachment of isoprenoid side chains confers hydrophobic character. Proteins having such a modification are targeted to membranes.

32. 3-Hydroxy-3-methylglutaryl CoA is also a precursor for ketone-body synthesis. If fuel is needed elsewhere in the body, as might be the case during a fast, 3-hydroxy-3-methylglutaryl CoA is converted into the ketone acetoacetate. If energy needs are met, the liver will synthesize cholesterol.

33. One way in which phosphatidylcholine can be synthesized is by the addition of three methyl groups to phosphatidylethanolamine. The methyl donor is a modified form of methionine, S-adenosylmethionine or SAM (Section 24.2).

34. Citrate is transported out of the mitochondria in times of plenty. ATP-citrate lyase yields acetyl CoA and oxaloacetate. The acetyl CoA can then be used to synthesize cholesterol.
35. (a) There is no effect. (b) Because actin is not controlled by cholesterol, the amount isolated should be the same in both experimental groups; a difference would suggest a problem in the RNA isolation. (c) The presence of cholesterol in the diet dramatically reduces the amount of HMG-CoA reductase protein. (d) A common means of regulating the amount of a protein present is to regulate transcription, which is clearly not the case here. (e) The translation of mRNA could be inhibited, and the protein could be rapidly degraded.

Chapter 27

1. Adipose tissue is now known to be an active endocrine organ, secreting signal molecules called adipokines.

2. Caloric homeostasis is the condition in which the energy expenditure of an organism is equal to the energy intake.

3. Leptin and insulin

4. CCK produces a feeling of satiety and stimulates the secretion of digestive enzymes by the pancreas and the secretion of bile salts by the gall bladder. GLP-1 also produces a feeling of satiety; in addition, it potentiates the glucose-induced secretion of insulin by the β cells of the pancreas.

5. Obviously, something is amiss. Although the answer is not known, the leptin-signaling pathway appears to be inhibited by suppressors of cytokine signaling, the regulatory proteins.

6. 1: a, b; 2: f; 3: c, d, f; 4: c, d; 5: c; 6: f; 7: e; 8: e; 9: e.

7. Phosphorylation of dietary glucose after it enters the liver; gluconeogenesis; glycogen breakdown.

8. Type 1 diabetes is due to autoimmune destruction of the insulin-producing cells of the pancreas. Type 1 diabetes is also called insulin-dependent diabetes because affected people require insulin to survive. Type 2 diabetes is characterized by insulin resistance. Insulin is produced, but the tissues that should respond to insulin, such as muscle, do not.

9. Leptin stimulates processes impaired in diabetes. For instance, leptin stimulates fatty acid oxidation, inhibits triacylglycerol synthesis, and increases the sensitivity of muscle and the liver to insulin.

10. (a) A watt is equal to 1 joule (J) per second (0.239 calorie per second). Hence, 70 W is equivalent to 0.07 kJ s⁻¹ (0.017 kcal s⁻¹). (b) A watt is a current of 1 ampere (A) across a potential of 1 volt (V). For simplicity, let us assume that all the electron flow is from NADH to O₂ (a potential drop of 1.14 V). Hence, the current is 61.4 A, which corresponds to 3.86×10^{20} electrons per second (1 A = 1 coulomb s⁻¹ = 6.28×10^{18} charge s⁻¹).

(c) About 2.5 molecules of ATP are formed per molecule of NADH oxidized (two electrons). Hence, 1 molecule of ATP is formed per 0.8 electron transferred. A flow of 3.86×10^{20} electrons per second therefore leads to the generation of 4.83×10^{20} molecules of ATP per second, or 0.80 mmol s⁻¹.

(d) The molecular weight of ATP is 507. The total body content of ATP of 50 g is equal to 0.099 mol. Hence, ATP turns over about once in 125 seconds when the body is at rest.

11. (a) The stoichiometry of the complete oxidation of glucose is

$$C_6H_{12}O_6 + 6 O_2 \longrightarrow 6 CO_2 + 6 H_2O$$

and that of tripalmitoylglycerol is

 $C_{51}H_{98}O_2 + 72.5 O_2 \longrightarrow 51 CO_2 + 49 H_2O$

Hence, the RQ values are 1.0 and 0.703, respectively. (b) An RQ value reveals the relative use of carbohydrates and fats as fuels. The RQ of a marathon runner typically decreases from 0.97 to 0.77 in the course of a race. The lowering of the RQ indicates the shift in fuel from carbohydrates to fat.

12. One gram of glucose (molecular weight 180.2) is equal to 5.55 mmol, and one gram of tripalmitoylglycerol (molecular weight 807.3) is equal to 1.24 mmol. The reaction stoichiometries (see Problem 11) indicate that 6 mol of H_2O is produced per mole of glucose oxidized, and 49 mol of H_2O is produced per mole of tripalmitoylglycerol oxidized. Hence, the H_2O yields per gram of fuel are 33.3 mmol (0.6 g) for glucose and 60.8 mmol (1.09 g) for tripalmitoylglycerol. Thus, complete oxidation of this fat gives 1.82 times as much water as does glucose. Another advantage of triacylglycerols is that they can be stored in essentially anhydrous form, whereas glucose is stored as glycogen, a highly hydrated polymer. A hump consisting mainly of glycogen would be an intolerable burden—far more than the straw that broke the camel's back.

13. The starved-fed cycle is the nightly hormonal cycle that humans experience during sleep and on eating. The cycle maintains adequate amounts of blood glucose. The starved part sleep—is characterized by increased glucagon secretion and decreased insulin secretion. After a meal, glucagon concentration falls and insulin concentration rises.

14. Ethanol is oxidized to yield acetaldehyde by alcohol dehydrogenase, which is subsequently oxidized to acetate acetaldehyde. Ethanol is also metabolized to acetaldehyde by the MEOS, with the subsequent depletion of NADPH.

15. First, fatty liver develops owing to the increased amounts of NADH that inhibit fatty acid oxidation and stimulate fatty acid

synthesis. Second, alcoholic hepatitis begins owing to oxidative damage and damage due to excess acetaldehyde that results in cell death. Finally, fibrous tissues form, creating scars that impair blood flow and biochemical function. Ammonia cannot be converted into urea, and its toxicity leads to coma and death.

16. A typical macadamia nut has a mass of about 2 g. Because it consists mainly of fats (~37 kJ g⁻¹, ~9 kcal g⁻¹), a nut has a value of about 75 kJ (18 kcal). The ingestion of 10 nuts results in an intake of about 753 kJ (180 kcal). As stated in the answer to Problem 10, a power consumption of 1 W corresponds to 1 J s⁻¹ (0.239 cal s⁻¹), and so 400-W running requires 0.4 kJ s⁻¹ (0.0956 kcal s⁻¹). Hence, a person would have to run 1882 s, or about 31 minutes, to spend the calories provided by 10 nuts.

17. A high blood-glucose level triggers the secretion of insulin, which stimulates the synthesis of glycogen and triacylglycerols. A high insulin level would impede the mobilization of fuel reserves during the marathon.

18. A lack of adipose tissue leads to an accumulation of fats in the muscle, with the generation of insulin resistance. The experiment shows that adipokines secreted by the adipose tissue, here leptin, facilitate in some fashion the action of insulin in muscle.

19. Such a mutation would increase the phosphorylation of the insulin receptor and IRS in muscle and would improve insulin sensitivity. Indeed, PTP1B is an attractive therapeutic target for type 2 diabetes. 20. Lipid mobilization can be so rapid that it exceeds the ability of the liver to oxidize the lipids or convert them into ketone bodies. The excess is reesterified and released into the blood as VLDLs. 21. A role of the liver is to provide glucose for other tissues. In the liver, glycolysis is used not for energy production but for biosynthetic purposes. Consequently, in the presence of glucagon, liver glycolysis stops so that the glucose can be released into the blood. 22. The urea cycle and gluconeogenesis

23. (a) Insulin inhibits lipid utilization.

(b) Insulin stimulates protein synthesis, but there are no amino acids in the children's diet. Moreover, insulin inhibits protein breakdown. Consequently, muscle proteins cannot be broken down and used for the synthesis of essential proteins.

(c) Because proteins cannot be synthesized, blood osmolarity is too low. Consequently, fluid leaves the blood. An especially important protein for maintaining blood osmolarity is albumin.

24. During strenuous exercise, muscle converts glucose into pyruvate through glycolysis. Some of the pyruvate is processed by cellular respiration. However, some of it is converted into lactate and released into the blood. The liver takes up the lactate and converts it into glucose through gluconeogenesis. Muscle may process the carbon skeletons of branched-chain amino acids aerobically. The nitrogens of these amino acids are transferred to pyruvate to form alanine, which is released into the blood and taken up by the liver. After the transamination of the amino group to α -ketoglutarate, the resulting pyruvate is converted into glucose. Finally, muscle glycogen may be mobilized, and the released glucose can be used by muscle.

25. This conversion allows muscle to function anaerobically. NAD⁺ is regenerated when pyruvate is reduced to lactate, and so energy can continue to be extracted from glucose during strenuous exercise. The liver converts the lactate into glucose.

26. Fatty acids and glucose, respectively.

27. This practice is called carbo-loading. Depleting the glycogen stores will initially cause the muscles to synthesis a large amount of glycogen when dietary carbohydrates are provided and will lead to the supercompensation of glycogen stores.

28. The oxygen consumption at the end of exercise is used to replenish ATP and creatine phosphate and to oxidize any lactate produced. 29. Oxygen is used in oxidative phosphorylation to resynthesize ATP and creatine phosphate. The liver converts lactate released by the muscle into glucose. Blood must be circulated to return the body temperature to normal, and so the heart cannot return to its resting rate immediately. Hemoglobin must be reoxygenated to replace the oxygen used in exercise. The muscles that power breathing must continue working at the same time as the exercised muscles are returning to resting states. In essence, all the biochemical systems activated in intense exercise need increased oxygen to return to the resting state.

30. Ethanol may replace water that is hydrogen bonded to proteins and membrane surfaces. This alteration of the hydration state of the protein would alter its conformation and hence function. Ethanol may also alter phospholipid packing in membranes. The two effects suggest that integral membrane proteins would be most sensitive to ethanol, as indeed seems to be the case.

31. Cells from the type I fiber would be rich in mitochondria, whereas those of the type II fiber would have few mitochondria.32. (a) The ATP expended during this race amounts to about 8380 kg, or 18,400 pounds. (b) The cyclist would need about \$1,260,000,000 to complete the race.

33. 55 pounds = 25 kg = 25,000 g = total weight gain

40 years \times 365 days year⁻¹ = 14,600 days

 $25,000 \text{ g}/14,600 \text{ days} = 1.7 \text{ g day}^{-1}$

which is equivalent to an extra pat of butter per day. Her BMI is 26.5, and she would be considered overweight but not obese.
34. Exercise greatly enhances the ATP needs of muscle cells. To more efficiently meet these needs, more mitochondria are synthesized.
35. The inability of muscle mitochondria to process all of the fatty acids produced by overnutrition leads to excessive levels of diacylglycerol and ceramide in the muscle cytoplasm. These second-messenger molecules activate enzymes that impair insulin signaling.
36. Both are due to a lack of thiamine (vitamin B₁). Thiamine, which is sometimes called aneurin, is required most notably for the proper functioning of pyruvate dehydrogenase.

37. (a) Red blood cells always produce lactate, and fast-twitch muscle fibers (see Problem 31) also produce a large amount of lactate. (b) At that point, the athlete is beginning to move into anaerobic exercise, in which most energy is produced by anaerobic glycolysis. (c) The lactate threshold is essentially the point at which the athlete switches from aerobic exercise, which can be done for extended periods, to anaerobic exercise, essentially sprinting, which can be done for only short periods. The idea is to race at the extreme of his or her aerobic capacity until the finish line is in sight and then to switch to anaerobic.

(d) Training increases the amount of blood vessels and the number of muscle mitochondria. Together, they increase the ability to process glucose aerobically. Consequently, a greater effort can be expended before the switch to anaerobic energy production.

Chapter 28

1. DNA polymerase I uses deoxyribonucleoside triphosphates; pyrophosphate is the leaving group. DNA ligase uses DNA– adenylate (AMP joined to the 5'-phosphoryl group) as a reaction partner; AMP is the leaving group. Topoisomerase I uses a DNA–tyrosyl intermediate (5'-phosphoryl group linked to the phenolic OH group); the tyrosine residue of the enzyme is the leaving group.

2. Positive supercoiling resists the unwinding of DNA. The melting temperature of DNA increases in proceeding from negatively supercoiled to relaxed to positively supercoiled DNA. Positive supercoiling is probably an adaptation to high temperature.
The nucleotides used for DNA synthesis have the triphosphate attached to the 5'-hydroxyl group with free 3'-hydroxyl groups.
 Such nucleotides can be utilized only for 5'-to-3' DNA synthesis.
 DNA replication requires RNA primers. Without appropriate

ribonucleotides, such primers cannot be synthesized.

5. This close contact prevents the incorporation of ribonucleotides rather than 2'-deoxyribonucleotides.

6. (a) 96.2 revolutions per second (1000 nucleotides per second divided by 10.4 nucleotides per turn for B-DNA gives 96.2 rps). (b) 0.34 μ m s⁻¹ (1000 nucleotides per second corresponds to 3400 Å s⁻¹ because the axial distance between nucleotides in B-DNA is 3.4 Å).

7. Eventually, the DNA would become so tightly wound that movement of the replication complex would be energetically impossible. 8. Linking number Lk = Tw + Wr = 48 + 3 = 51. If Tw = 50, the Wr = 1.

9. A hallmark of most cancer cells is prolific cell division, which requires DNA replication. If the telomerase were not activated, the chromosomes would shorten until they became nonfunctional, leading to cell death.

10. No.

11. Treat the DNA briefly with endonuclease to occasionally nick each strand. Add the polymerase with the radioactive dNTPs. At the broken bond, or nick, the polymerase will degrade the existing strand with its $5' \rightarrow 3'$ exonuclease activity and replace it with a radioactive complementary copy by using its polymerase activity. This reaction scheme is referred to as nick translation because the nick is moved, or translated, along the DNA molecule without ever becoming sealed.

12. If replication were unidirectional, tracks with a low grain density at one end and a high grain density at the other end would be seen. On the other hand, if replication were bidirectional, the middle of a track would have a low density, as shown in the diagram below. For *E. coli*, the grain tracks are denser on both ends than in the middle, indicating that replication is bidirectional.



13. (a) Pro (CCC), Ser (UCC), Leu (CUC), and Phe (UUC). Alternatively, the last base of each of these codons could be U.

(b) Nitrous acid.

14. Potentially deleterious side reactions are prevented. The enzyme itself might be damaged by light if it could be activated by light in the absence of bound DNA harboring a pyrimidine dimer.

15. The free DNA ends that appear in the absence of telomeres are repaired by DNA fusion.

16. The free energy of ATP hydrolysis under standard conditions is $-30.5 \text{ kJ mol}^{-1}$ ($-7.3 \text{ kcal mol}^{-1}$). In principle, it could be used to break three base pairs.

17. The oxidation of guanine could lead to DNA repair: DNA strand cleavage could allow looping out of the triplet repeat regions and triplet expansion.

18. The release of DNA topoisomerase II after the enzyme has acted on its DNA substrate requires ATP hydrolysis. Negative supercoiling requires only the binding of ATP, not its hydrolysis.
19. (a) Size; the top is relaxed and the bottom is supercoiled DNA. (b) Topoisomers. (c) The DNA is becoming progressively more unwound, or relaxed, and thus slower moving.
20. (a) It was used to determine the number of spontaneous revertants—that is, the background mutation rate.
(b) To firmly establish that the system was working. A known mutagen's failure to produce revertants would indicate that something was wrong with the experimental system.
(c) The chemical itself has little mutagenic ability but is apparently activated into a mutagen by the liver homogenate.
(d) Cytochrome P450 system.

Chapter 29

1. The sequence of the coding (+, sense) strand is

5'-ATGGGGAACAGCAAGAGTGGGGCCCTGTCCAAGGAG-3'

and the sequence of template (-, antisense) strand is

3'-TACCCCTTGTCGTTCTCACCCCGGGACAGGTTCCTC-5'

2. An error will affect only one molecule of mRNA of many synthesized from a gene. In addition, the errors do not become a permanent part of the genomic information.

3. At any given instant, only a fraction of the genome (total DNA) is being transcribed. Consequently, speed is not essential.

4. The active sites are related by convergent evolution.

5. Heparin, a glycosaminoglycan, is highly anionic. Its negative charges, like the phosphodiester bridges of DNA templates, allow it to bind to lysine and arginine residues of RNA polymerase.

6. This mutant σ will competitively inhibit the binding of holoenzyme and prevent the specific initiation of RNA chains at promoter sites.

7. The core enzyme without σ binds more tightly to the DNA template than does the holoenzyme. The retention of σ after chain initiation would make the mutant RNA polymerase less processive. Hence, RNA synthesis would be much slower than normal.

8. A 100-kd protein contains about 910 residues, which are encoded by 2730 nucleotides. At a maximal transcription rate of 50 nucleotides per second, the mRNA would be synthesized in 54.6 s.

 The RNA polymerase slides along the DNA rapidly rather than simply diffusing through three-dimensional space.
 The start site is in red time:

10. The start site is in red type:

5'-GCCGTTGACACCGTTCGGCGATCGATCCGCTATAATGTGTGGATCCGCTT-3'

11. Initiation at strong promoters takes place every 2 s. In this interval, 100 nucleotides are transcribed. Hence, centers of transcription bubbles are 34 nm (340 Å) apart.

12. (a) The lowest band on the gel will be that of strand 3 alone (i), whereas the highest will be that of stands 1, 2, and 3 and core polymerase (v). Band ii will be at the same position as band i because the RNA is not complementary to the nontemplate strand, whereas band iii will be higher because a complex is formed between RNA and the template strand. Band iv will be higher than the others because strand 1 is complexed to 2, and strand 2 is complexed to 3. Band v is the highest because core polymerase associates with the three strands.

(b) None, because rifampicin acts before the formation of the open complex.

ANSWERS TO PROBLEMS

(c) RNA polymerase is processive. When the template is bound, heparin cannot enter the DNA-binding site.

(d) When GTP is absent, synthesis stops when the first cytosine residue downstream of the bubble is encountered in the template strand. In contrast, with all four nucleoside triphosphates present, synthesis will continue to the end of the template.

13. RNA polymerase must backtrack before cleavage, leading to dinucleotide products.

14. The base-pairing energy of the di- and trinucleotide DNA-RNA hybrids formed at the very beginning of transcription is not sufficient to prevent strand separation and loss of product. 15. (a) Because cordycepin lacks a 3'-OH group, it cannot participate in $3' \rightarrow 5'$ bond formation. (b) Because the poly(A) tail is a long stretch of adenosine nucleotides, the likelihood that a molecule of cordycepin would become incorporated is higher than with most RNA. (c) Yes, it must be converted into cordycepin 5'-triphosphate.

16. There are $2^8 = 256$ possible products.

17. The relation between the -10 and -35 sequences could be affected by torsional strain. The fact that topoisomerase II introduces negative supercoils in DNA prevents this enzyme from overstimulating the expression of its own gene.

18. Ser-Ile-Phe-His-Pro-Stop

19. A mutation that disrupted the normal AAUAAA recognition sequence for the endonuclease could account for this finding. In fact, a change from U to C in this sequence caused this defect in a thalassemic patient. Cleavage was at the AAUAAA 900 nucleotides downstream of this mutant AACAAA site.

20. One possibility is that the 3' end of the poly(U) donor strand cleaves the phosphodiester bond on the 5' side of the insertion site. The newly formed 3' terminus of the acceptor strand then cleaves the poly(U) strand on the 5' side of the nucleotide that initiated the attack. In other words, a uridine residue could be added by two transesterification reactions. This postulated mechanism is similar to the one in RNA splicing.

21. Alternative splicing and RNA editing. Covalent modification of the proteins subsequent to synthesis further enhances the complexity. 22. Attach an oligo(dT) or oligo(U) sequence to an inert support to create an affinity column. When RNA is passed through the column, only poly(A)-containing RNA will be retained.

23. (a) Different amounts of RNA are present for the various genes.(b) Although all of the tissues have the same genes, the genes are expressed to different extents in different tissues.

(c) These genes are called housekeeping genes—genes that most tissues express. They might include genes for glycolysis or citric acid cycle enzymes.

(d) The point of the experiment is to determine which genes are initiated in vivo. The initiation inhibitor is added to prevent initiation at start sites that may have been activated during the isolation of the nuclei.

24. DNA is the single strand that forms the trunk of the tree. Strands of increasing length are RNA molecules; the beginning of transcription is where growing chains are the smallest; the end of transcription is where chain growth stops. Direction is left to right. Many enzymes are actively transcribing each gene.

Chapter 30

1. The Oxford English Dictionary defines translation as the action or process of turning from one language into another. Protein synthesis converts nucleic acid sequence information into amino acid sequence information.

2. An error frequency of 1 incorrect amino acid every 10⁴ incorporations allows for the rapid and accurate synthesis of proteins as

large as 1000 amino acids. Higher error rates would result in too many defective proteins. Lower error rates would likely slow the rate of protein synthesis without a significant gain in accuracy. 3. (i) Each is a single chain. (ii) They contain unusual bases. (iii) Approximately half of the bases are base-paired to form double helices. (iv) The 5' end is phosphorylated and is usually pG. (v) The amino acid is attached to the hydroxyl group of the A residue of the CCA sequence at the 3' end of the tRNA. (vi) The anticodon is located in a loop near the center of the tRNA sequence. (vii) The molecules are L-shaped.

4. First is the formation of the aminoacyl adenylate, which then reacts with the tRNA to form the aminoacyl-tRNA. Both steps are catalyzed by aminoacyl-tRNA synthetase.

5. Unique features are required so that the aminoacyl-tRNA synthetases can distinguish among the tRNAs and attach the correct amino acid to the proper tRNA. Common features are required because all tRNAs must interact with the same protein-synthesizing machinery.

6. An activated amino acid is one linked to the appropriate tRNA. 7. (a) No; (b) no; (c) yes.

8. The ATP is cleaved to AMP and PP_i. Consequently, a second ATP is required to convert AMP into ADP, the substrate for oxidative phosphorylation.

9. Amino acids larger than the correct amino acid cannot fit into the active site of the tRNA. Smaller but incorrect amino acids that become attached to the tRNA fit into the editing site and are cleaved from the tRNA.

10. Recognition sites on both faces of the tRNAs may be required to uniquely identify the 20 different tRNAs.

11. The first two bases in a codon form Watson–Crick base pairs that are checked for fidelity by bases of the 16S rRNA. The third base is not inspected for accuracy, and so some variation is tolerated.

12. Four bands: light, heavy, a hybrid of light 30S and heavy 50S, and a hybrid of heavy 30S and light 50S

13. Two hundred molecules of ATP are converted into 200 AMP + 400 P_i to activate the 200 amino acids, which is equivalent to 400 molecules of ATP. One molecule of GTP is required for initiation, and 398 molecules of GTP are needed to form 199 peptide bonds.

14. (a, d, and e) Type 2; (b, c, and f) type 1.

15. The reading frame is a set of contiguous, nonoverlapping three-nucleotide codons that begins with a start codon and ends with a stop codon.

16. A mutation caused by the insertion of an extra base can be suppressed by a tRNA that contains a fourth base in its anticodon. For example, UUUC rather than UUU is read as the codon for phenylalanine by a tRNA that contains 3'-AAAG-5' as its anticodon.

17. One approach is to synthesize a tRNA that is acylated with a reactive amino acid analog. For example, bromoacetyl-phenyla-lanyl-tRNA is an affinity-labeling reagent for the P site of *E. coli* ribosomes.

18. The sequence GAGGU is complementary to a sequence of five bases at the 3' end of 16S rRNA and is located several bases upstream of an AUG start codon. Hence, this region is a start signal for protein synthesis. The replacement of G by A would be expected to weaken the interaction of this mRNA with the 16S rRNA and thereby diminish its effectiveness as an initiation signal. In fact, this mutation results in a 10-fold decrease in the rate of synthesis of the protein specified by this mRNA.

19. The peptide would be Phe-Cys-His-Val-Ala-Ala. The codons UGC and UGU encode cysteine but, because the

cysteine was modified to alanine, alanine is incorporated in place of cysteine.

20. Proteins are synthesized from the amino to the carboxyl end on ribosomes, whereas they are synthesized in the reverse direction in the solid-phase method. The activated intermediate in ribosomal synthesis is an aminoacyl-tRNA; in the solid-phase method, it is the adduct of the amino acid and dicyclohexylcarbodiimide. 21. The error rates of DNA, RNA, and protein synthesis are of the order of 10^{-10} , 10^{-5} , and 10^{-4} , respectively, per nucleotide (or amino acid) incorporated. The fidelity of all three processes depends on the precision of base-pairing to the DNA or mRNA template. Few errors are corrected in RNA synthesis. In contrast, the fidelity of DNA synthesis is markedly increased by the $3' \rightarrow 5'$ proofreading nuclease activity and by postreplicative repair. In protein synthesis. the mischarging of some tRNAs is corrected by the hydrolytic action of aminoacyl-tRNA synthetase. Proofreading also takes place when aminoacyl-tRNA occupies the A site on the ribosome; the GTPase activity of EF-Tu sets the pace of this final stage of editing. 22. GTP is not hydrolyzed until aminoacyl-tRNA is delivered to the A site of the ribosome. An earlier hydrolysis of GTP would be wasteful because EF-Tu-GDP has little affinity for aminoacyl-tRNA. 23. The translation of an mRNA molecule can be blocked by antisense RNA, an RNA molecule with the complementary sequence. The antisense-sense RNA duplex cannot serve as a template for translation; single-stranded mRNA is required. Furthermore, the antisense-sense duplex is degraded by nucleases. Antisense RNA added to the external medium is spontaneously taken up by many cells. A precise quantity can be delivered by microinjection. Alternatively, a plasmid encoding the antisense RNA can be introduced into target cells.

24. (a) A_5 . (b) $A_5 > A_4 > A_3 > A_2$. (c) Synthesis is from the amino terminus to the carboxyl terminus.

25. These enzymes convert nucleic acid information into protein information by interpreting the tRNA and linking it to the proper amino acid.

26. The rate would fall because the elongation step requires that the GTP be hydrolyzed before any further elongation can take place.
27. Protein factors modulate the initiation of protein synthesis. The role of IF1 and IF3 is to prevent premature binding of the 30S and 50S ribosomal subunits, whereas IF2 delivers Met-tRNAf to the ribosome. Protein factors are also required for elongation (EF-G and EF-Tu), for termination (release factors, RFs), and for ribosome dissociation (ribosome release factors, RRFs).
28. The signal sequence, signal-recognition particle (SRP), the

28. The signal sequence, signal-recognition particle (SRP), the SRP receptor, and the translocon.

29. The formation of peptide bonds, which in turn are powered by the hydrolysis of the aminoacyl-tRNAs.

30. The Shine–Dalgarno sequence of the mRNA base-pairs with a part of the 16S rRNA of the 30S subunit, which positions the subunit so that the initiator AUG is recognized.

	Prokaryote	Eukaryote
Ribosome size	60S	80S
mRNA	polycistronic	Not polycistronic
Initiation	Shine–Dalgarno is required	First AUG is used
Protein factors	Required	Many more required
Relation to transcription	Translation can start before transcription is completed	Transcription and translation are spatially separated
First amino acid	fMet	Met

32. The SRP binds to the signal sequence and inhibits further translation. The SRP ushers the inhibited ribosome to the ER, where it interacts with the SRP receptor (SR). The SRP–SR complex binds the translocon and simultaneously hydrolyzes GTP. On GTP hydrolysis, SRP and SR dissociate from each other and from the ribosome. Protein synthesis resumes and the nascent protein is channeled through the translocon.

33. The alternative would be to have a single ribosome translating a single mRNA molecule. The use of polysomes allows more protein synthesis per mRNA molecule in a given period of time and thus the production of more protein.

34. (a) 1, 2, 3, 5, 6, 10; (b) 1, 2, 7, 8,; (c) 1, 4, 8, 9.

35. Transfer RNAs have roles in several recognition processes. A tRNA must be recognized by the appropriate aminoacyl-tRNA synthetase, and the tRNA must interact with the ribosome and, in particular, with the peptidyl transferase.

36. The nucleophile is the amino group of the aminoacyl-tRNA. This amino group attacks the carbonyl group of the ester of peptidyl-tRNA to form a tetrahedral intermediate, which eliminates the tRNA alcohol to form a new peptide bond.

37. The aminoacyl-tRNA can be initially synthesized. However, the side-chain amino group attacks the ester linkage to form a six-membered amide, releasing the tRNA.

38. EF-Ts catalyzes the exchange of GTP for GDP bound to EF-Tu. In G-protein cascades, an activated 7TM receptor catalyzes GTP–GDP exchange in a G protein.

39. The α subunits of G proteins are inhibited by a similar mechanism in cholera and whooping cough (Section 14.5).

40. Glu-tRNA^{Gln} is formed by misacylation. The activated glutamate is subsequently amidated to form Gln-tRNA^{Gln}. Ways in which glutamine is formed from glutamate were discussed in Section 24.2. In regard to *H. pylori*, a specific enzyme, Glu-tRNA^{Gln} amidotransferase, catalyzes the following reaction:

$$Gln + Glu - tRNA^{Gln} + ATP \longrightarrow$$

 $Gln - tRNA^{Gln} + glu + ADP + P_i$

Glu-tRNA^{Glu} is not a substrate for the enzyme; so the transferase must also recognize aspects of the structure of tRNA^{Gln}.
41. The primary structure determines the three-dimensional structure of the protein. Thus, the final phase of information trans-

fer from DNA to RNA to protein synthesis is the folding of the protein into its functional state. 42. (a) eIF-4H has two effects: (1) the extent of unwinding is

42. (a) eIF-4H has two effects: (1) the extent of unwinding is increased and (2) the rate of unwinding is increased, as indicated by the increased rise in activity at early reaction times.
(b) To firmly establish that the effect of eIF-H4 was not due to any inherent helicase activity.

(c) Half-maximal activity was achieved at 0.11 μ M of eIF-4H. Therefore, maximal stimulation would be achieved at a ratio of 1:1. (d) eIF-4H enhances the rate of unwinding of all helices, but the effect is greater as the helices increase in stability.

(e) The results in graph C suggest that eIF-4H increases the processivity.

43. (a) The three peaks represent, from left to right, the 40S ribosomal subunit, the 60S ribosomal subunit, and the 80S ribosome. (b) Not only are ribosomal subunits and the 80S ribosome present, but polysomes of various lengths also are apparent. The individual peaks in the polysome region represent polysomes of discrete length.

(c) The treatment significantly inhibited the number of polysomes while increasing the number of free ribosomal subunits. This outcome could be due to inhibited protein-synthesis initiation or inhibited transcription.

A40

ANSWERS TO PROBLEMS

Chapter 31

1. (a) Cells will express β -galactosidase, *lac* permease, and thiogalactoside transacetylase even in the absence of lactose. (b) Cells will express β -galactosidase, *lac* permease, and thiogalactoside transacetylase even in the absence of lactose. (c) The levels of catabolic enzymes such as β -galactosidase and arabinose isomerase will remain low even at low levels of glucose.

2. The concentration is $1/(6 \times 10^{23})$ moles per 10^{-15} liter = 1.7×10^{-9} M. Because $K_{\rm d} = 10^{-13}$ M, the single molecule should be bound to its specific binding site.

3. The number of possible 8-bp sites is $4^8 = 65,536$. In a genome of 4.6×10^6 base pairs, the average site should appear $(4.6 \times 10^6)/65,536 = 70$ times. Each 10-bp site should appear 4 times. Each 12-bp site should appear 0.27 times (many 12-bp sites will not appear at all).

4. The *lac* repressor does not bind DNA when the repressor is bound to a small molecule (the inducer), whereas the *pur* repressor binds DNA only when the repressor is bound to a small molecule (the corepressor). The *E. coli* genome contains only a single *lac* repressor-binding region, whereas it has many sites for the *pur* repressor 5. Anti-inducers bind to the conformation of repressors, such as the *lac* repressor, that are capable of binding DNA. They occupy a site that overlaps that for the inducer and, therefore, compete for binding to the repressor.

6. The inverted repeat may be a binding site for a dimeric DNAbinding protein or it may correspond to a stem-loop structure in the encoded RNA.

7. Bacteriophage λ would be more likely to enter the lytic phase because the cooperative binding of the λ repressor to $O_R 2$ and $O_R 1$, which supports the lysogenic pathway, would be disrupted.

 λ repressor gene -10 region GATTTA -35 region TAGATA Cro gene -10 region TAATGG -35 region TTGACT There are four differences in the -10 region and three differences in the -35 region.

9. Increased Cro concentration reduces the expression of the λ repressor gene. Increased λ repressor concentration reduces the expression of the Cro gene. At low λ repressor concentration, increased λ repressor concentration increases the expression of the λ repressor gene. At higher λ repressor concentrations, increased λ repressor concentration decreases the expression of the λ repressor concentration decreases the expression of the λ repressor gene.

10. Normally, bacterial mRNAs have a leader sequence in which a Shine–Delgarno sequence precedes the AUG start codon. The absence of a leader would be expected to lead to inefficient translation.

11. Add each compound to a culture of *V. fischeri* at low density and look for the development of luminescence.

12. ACC, 7; ACA, 1, ACU, 0; ACG, 0.

13. The reaction takes place with overall retention of configuration. Each step likely takes place with inversion of configuration, which suggests that the reaction consists of two (or some other even number of) steps. A possible mechanism is nucleophilic attack by the carboxylate group of Glu 537 on the C-1 carbon atom of the galactose moiety within glucose, releasing glucose and forming an intermediate with the galactose linked to the enzyme through an ester linkage. Water then attacks this carbon atom, displacing the glutamate carboxylate and releasing galactose. 14. The binding appears to be half complete at a concentration of λ repressor near 3.7 nM. Thus, K_d is approximately 3.7 nM and $\Delta G^{\circ} = -48$ kJ/mol (-11 kcal/mol) at 298 K.

Chapter 32

1. The distribution of charged amino acids is H2A (13 K, 13 R, 2 D, 7 E, charge = +17), H2B (20 K, 8 R, 3 D, 7 E, charge = +18), H3 (13 K, 18 R, 4 D, 7 E, charge = +20), H4 (11 K, 14 R,

3 D, 4 E, charge = +18). The total charge of the histone octamer is estimated to be $2 \times (17 + 18 + 20 + 18) = +146$. The total charge on 150 base pairs of DNA is -300. Thus, the histone octamer neutralizes approximately one-half of the charge. 2. The presence of a particular DNA fragment could be detected by hybridization, by PCR, or by direct sequencing.

3. The total length of the DNA is estimated to be 145 bp × 3.4 Å/bp = 493 Å, which represents 1.75 turns or $1.75 \times 2\pi r = 11.0r$. Thus, the radius is estimated to be r = 493 Å/11.0 = 44.8 Å.

4. 5-Azacytidine cannot be methylated. Some genes, normally repressed by methylation, will be active.

5. Proteins containing these domains will be targeted to methylated DNA in repressed promoter regions. They would likely bind in the major groove because that is where the methyl group is located.

6. Gene expression is not expected to respond to the presence of estrogen. However, genes for which expression normally responds to estrogen will respond to the presence of progesterone.

7. The acetylation of lysine will reduce the charge from +1 to 0. The methylation of lysine will not reduce the charge.

8. On the basis of the pattern of cysteine and histidine residues, this region appears to contain three zinc-finger domains.

9. 10/4000 = 0.25% of 12 Mb = 30 kilobase pairs. 10. The addition of an IRE to the 5' end of the mRNA is expected to block translation in the absence of iron. The addition of an IRE to the 3' end of the mRNA is not expected to block translation, but it might affect mRNA stability.

11. The sequences of all of the mRNAs would be searched for sequences that are fully or nearly complementary to the sequence of the miRNA. These sequences would be candidates for regulation by this mRNA.

12. The amino group of the lysine residue, formed from the protonated form by a base, attacks the carbonyl group of acetyl CoA to generate a tetrahedral intermediate. This intermediate collapses to form the amide bond and release CoA.

13. In mouse DNA, most of the HpaII sites are methylated and therefore not cut by the enzyme, resulting in large fragments. Some small fragments are produced from CpG islands that are unmethylated. For *Drosophila* and *E. coli* DNA, there is no methylation and all sites are cut.

Chapter 33

1. The transgenic nematode would avoid the compound. The identity of the ligand is determined by the receptor, whereas the behavioral response is dictated by the neuron in which the receptor is expressed.

2. Only a mixture of compounds C_5 -COOH and HOOC- C_7 -COOH is predicted to yield this pattern.

3. Bitter and sweet sensations are mediated by G proteins coupled to 7TM receptors, leading to millisecond time resolution. Salty and sour sensations are mediated directly by ion channels, which may lead to faster time resolution.

4. Sound travels 0.15 m in 428 μ s. The human hearing system is capable of sensing time differences of close to a microsecond, and so the difference in arrival times at the two ears is substantial. A system based on G proteins is unlikely to be able to reliably distinguish between signals arriving at the two ears, because G proteins typically respond in milliseconds.

5. If a plant tastes bitter, animals will avoid eating it even if it is nontoxic, which may provide a selective advantage to the plant.
6. Using mice in which either the gene for T1R1 or the gene for T1R3 has been disrupted, test the taste responses of these mice to glutamate, aspartate, and a wide variety of other amino acids.

7. These women have four functional color receptors: blue, red, green, and a red–green hybrid. The additional color receptor allows some colors that appear identical to most people to be distinguished. 8. 380 (one for each receptor); there are $(380 \times 379)/2! = 72,010$ combinations of two receptors; $(380 \times 379 \times 378)/3! = 9,073,260$ combinations of three receptors.

9. The absorption of light converts 11-*cis*-retinal into all-*trans*-retinal. 10. These compounds are enantiomers and must bind to protein receptors to elicit a smell. Even these subtle structural differences can affect relative receptor binding affinities and, hence, the elicited odor.

11. Vision: cGMP-gated channel; taste: amiloride-sensitive sodium channel; hearing; tip-link channel.

12. For all senses, ATP hydrolysis is required to generate and maintain ion gradients and membrane potential. Olfaction: ATP is required for the synthesis of cAMP. Gustation: ATP is required for the synthesis of cyclic nucleotides, and GTP is required for the action of gustducin in the detection of bitter and sweet tastes. Vision: GTP is required for the synthesis of cGMP and for the action of transducin. Hearing and touch: ATP hydrolysis is required to generate and maintain ion gradients and membrane potential and may be required for other roles as well. 13.



Chapter 34

1. The innate immune system responds rapidly to common features present in many pathogens. The genes for the innate immune system's key molecules are expressed without substantial modification. In contrast, the adaptive immune system responds to specific features present only in a given pathogen. Its genes undergo significant rearrangement and mutation to enable specific recogition of a vast number of potential binding surfaces.

VJ and V(D)J recombination; variability in segment joining by the action of terminal deoxyribonucleotidyl transferase; somatic mutation.
 Affinity refers to the strength of a single interaction; avidity refers to the cumulative strength of multiple independent binding interactions. Avidity may play a significant role in the interaction between IgM and antigen because this immunoglobulin class features 10 binding sites.

4. The intracellular signaling domain common to each of the TLRs is responsible for docking other proteins and reporting that a targeted pathogen-associated molecular pattern (PAMP), such as LPS, has been detected. If a mutation within this domain interfered with the intracellular docking and signal transduction, then TLR-4 would not respond to LPS.

5. Viruses that contain dsRNA genomes would be expected to stimulate a TLR-3-mediated immune response.

6. (a) $\Delta G^{\circ\prime} = -37 \text{ kJ mol}^{-1} (-8.9 \text{ kcal mol}^{-1})$

(b) $K_a = 3.3 \times 10^6 \,\mathrm{M}^{-1}$

(c) $k_{on} = 4 \times 10^8 \text{ M}^{-1} \text{ s}^{-1}$. This value is close to the diffusion-controlled limit for the combination of a small molecule with a

protein (see p. 245). Hence, the extent of structural change is likely to be small; extensive conformational transitions take time. 7. The fluorescence enhancement and the shift to blue indicate that water is largely excluded from the combining site when the hapten is bound. Hydrophobic interactions contribute significantly to the formation of most antigen–antibody complexes. 8. (a) An antibody combining site is formed by CDRs from both the H and the L chains. The V_H and V_L domains are essential. A small proportion of F_{ab} fragments can be further digested to produce F_{v} , a fragment that contains just these two domains. C_H1 and C_L contribute to the stability of F_{ab} but not to antigen binding. (b) A synthetic F_v analog 248 residues long was prepared by expressing a synthetic gene consisting of a V_H gene joined to a V_L gene through a linker. See J. S. Huston et al., *Proc. Natl. Acad. Sci.* U. S. A. 85:5879–5883, 1988.

9. (a) Multivalent antigens lead to the dimerization or oligomerization of transmembrane immunoglobulins, an essential step in their activation. This mode of activation is reminiscent of that of receptor tyrosine kinases (Section 14.2).

(b) An antibody specific for a transmembrane immunoglobulin will activate a B cell by cross-linking these receptors. This experiment can be carried out by using, for example, a goat antibody to cross-link receptors on a mouse B cell.

10. B cells do not express T-cell receptors. The hybridization of T-cell cDNAs with B-cell mRNAs removes cDNAs that are expressed in both cells. Hence, the mixture of cDNAs subsequent to this hybridization are enriched in those encoding T-cell receptors. This procedure, called subtractive hybridization, is generally useful in isolating low-abundance cDNAs. Hybridization should be carried out by using mRNAs from a closely related cell that does not express the gene of interest. See S. M. Hedrick, M. M. Davis, D. I. Cohen, E. A. Nielsen, and M. M. Davis, *Nature* 308:149–153, 1984, for an interesting account of how this method was used to obtain genes for T-cell receptors.

11. TLR-4 is the receptor for LPS, a toxin found specifically in the walls of Gram-negative bacteria. Mutations that inhibit the function of TLR4 impair an affected person's defenses against this class of bacteria.

12. If the HLA alleles are not matched, then the recipient's T cell receptors will identify the MHC proteins of the transplanted tissue as nonself and transplant rejection is likely.

13. Purify an antibody with a specificity to one antigen. Unfold the antibody and allow it to re-fold either in the presence of the antigen or in the absence of the antigen. Test the re-folded antibodies for antigen-binding ability.

14. In some cases, V–D–J rearrangement will result in combining V, D, and J segments out of frame. mRNA molecules produced from such rearranged genes will produce truncated molecules if translated. This possibility is excluded by degrading the mRNA. 15. The mutant bacteria may still stimulate an immune response without causing disease. Hence, they may be valuable starting points for the design of a live attenuated vaccine for the original pathogenic strain.

16. The peptide is LLQATYSAV (L in second position, V in last).17. Catalysis is likely to require a base for removing a proton from a water molecule. A histidine, glutamate, or aspartate residue is most likely. In addition, a potential hydrogen-bond donor may be present and will interact with the negatively charged oxygen atom that forms in the transition state.

18. A phosphotyrosine residue in the carboxyl terminus of Src and related protein tyrosine kinases binds to its own SH2 domain to generate the inhibited from of Src (Section 14.5). Removal of the phosphoryl group from this residue will activate the kinase.

A42

ANSWERS TO PROBLEMS

19. (a) $K_{\rm d} = 10^{-7}$ M; (b) $K_{\rm d} = 10^{-9}$ M. The gene was probably generated by a point mutation in the gene for antibody A rather than by de novo rearrangement.

Chapter 35

1. (a) Skeletal muscle and eukaryotic cilia derive their free energy from ATP hydrolysis; the bacterial flagellar motor uses a protonmotive force.

(b) Skeletal muscle requires myosin and actin. Eukaryotic cilia require microtubules and dynein. The bacterial flagellar motor requires MotA, MotB, and FliG, as well as many ancillary components. 2. 6400 Å/80 Å = 80 body lengths per second. For a 10-foot automobile, this body-length speed corresponds to a speed of 80 \times 10 feet = 800 feet per second, or 545 miles per hour.

3. 4 pN = 8.8×10^{-13} pounds. The weight of a single motor domain is 100,000 g mol⁻¹/(6.023 × 10²³ molecules mol⁻¹) = 1.7×10^{-19} g = 3.7×10^{-22} pounds. Thus, a motor domain can lift ($8.8 \times 10^{-13}/3.7 \times 10^{-22}$) = 2.4×10^{9} times its weight.

4. Both actin filaments and microtubles are built from subunits and these subunits bind and hydrolyze nucleoside triphosphates. Actin filaments are built of a single type of subunit and these subunits bind ATP. Microtubules are built of two different types of subunits and these subunits bind GTP.

5. The light chains in myosin stiffen the lever arm. The light chains in kinesin bind cargo to be transported.

6. After death, the ratio of ADP to ATP increases rapidly. In the ADP form, myosin motor domains bind tightly to actin. Myosin-actin interactions are possible because the drop in ATP concentration also allows the calcium concentration to rise. clearing the blockage of actin by tropomyosin through the action of the troponin complex.

7. Above its critical concentration, ATP-actin will polymerize. The ATP will hydrolyze through time to form ADP-actin, which has a higher critical concentration. Thus, if the initial subunit concentration is between the critical concentrations of ATP-actin and ADP-actin, filaments will form initially and then disappear on ATP hydrolysis.

8. A one-base step is approximately 3.4 Å = 3.4×10^{-4} µm. If a stoichiometry of one molecule of ATP per step is assumed, this distance corresponds to a velocity of 0.017 μ m s⁻¹. Kinesin moves at a velocity of 6400 Å per second, or 0.64 $\mu m~s^{-1}$

9. A proton-motive force across the plasma membrane is necessary to drive the flagellar motor. Under conditions of starvation, this proton-motive force is depleted. In acidic solution, the pH difference across the membrane is sufficient to power the motor.

10. The mean distance between tumbles would be longer when the bacterium is moving up a gradient of a chemoattractant.

11. (a) 1.13×10^{-9} dyne

(b) 6.8×10^{14} erg (c) 6.6×10^{-11} erg per 80 molecules of ATP. A single kinesin motor provides more than enough free energy to power the transport of micrometer-size cargoes at micrometer-per-second velocities. 12. The spacing between identical subunits on microtubules is 8 nm. Thus, a kinesin molecule with a step size that is not a multiple of 8 nm would have to be able to bind at more than one type of site on the microtubule surface.

13. KIF1A must be tethered to an additional microtubule-binding element that retains an attachment to the microtubule when the motor domain releases.

14. Filaments built from subunits can be arbitrarily long, can be dynamically assembled and disassembled, and require only a small amount of genetic information to encode.

15. Protons still flow from outside to inside the cell. Each proton might pass into the outer half-channel of one MotA-MotB complex, bind to the MS ring, rotate clockwise, and pass into the inner half-channel of the neighboring MotA-MotB complex. 16. At a high concentration of calcium ion, Ca^{2+} binds to cal-

modulin. In turn, calmodulin binds to a protein kinase that phosphorylates myosin light chains and activates it. At low calcium ion concentration, the light chains are dephosphorylated by a Ca^{2+} independent phosphatase.

17. (a) The value of k_{cat} is approximately 13 molecules per second, whereas the $K_{\rm M}$ value for ATP is approximately 12 μ M. (b) The step size is approximately (380 - 120)/7 = 37 nm. (c) The step size is very large, which is consistent with the presence of six light-chain-binding sites and, hence, very long lever arms. The rate of ADP release is essentially identical with the overall k_{cat} ; so ADP release is rate limiting, which suggests that both motor domains can bind to sites 37 nm apart simultaneously. ADP release from the hindmost domain allows ATP to bind, leading to actin release and lever-arm motion.

Chapter 36

1. (a) Before; (b) after; (c) after; (d) after; (e) before; (f) after.

2. (a) Yes; (b) yes; (c) no (MW > 600). 3. If computer programs could estimate log(P) values on the basis of chemical structure, then the required laboratory time for drug development could be shortened. The determination of the relative solubilities of pharmaceutical candidates by allowing each compound to equilibrate between water and an organic phase would no longer be necessary.

4. Perhaps N-acetylcysteine would conjugate to some of the *N*-acetyl-*p*-benzoquinone imine that is produced by the metabolism of acetaminophen, thereby preventing the depletion of the liver's supply of glutathione.

5. In phase 1 clinical trials, approximately 10 to 100 healthy volunteers are typically enrolled in a study designed to assess safety. In contrast, a larger number of subjects are enrolled in a typical phase 2 trial. Moreover, these persons may benefit from the drug administered. In a phase 2 trial, efficacy, dosage, and safety can be assessed. 6. The binding of other drugs to albumin could cause extra coumadin to be released. (Albumin is a general carrier for hydrophobic molecules.)

7. A drug that inhibits a P450 enzyme may dramatically affect the disposition of another drug that is metabolized by that same enzyme. If this inhibited metabolism is not accounted for when dosing, the second drug may reach very high, and sometimes toxic, levels in the blood.

8. Unlike competitive inhibition, noncompetitive inhibition cannot be overcome with additional substrate. Hence, a drug that acts by a noncompetitive mechanism will be unaffected by changing levels of the physiological substrate.

9. An inhibitor of MDR could prevent the efflux of a chemotherapeutic drug from tumor cells. Hence, this type of an inhibitor could be useful in averting resistance to cancer chemotherapy. 10. Agents that inhibit one or more enzymes of the glycolytic pathway could act to deprive trypanosomes of energy and thus be useful for treating sleeping sickness. A difficulty is that glycolysis in the host cells also would be inhibited.

11. Imatinib is an inhibitor of the Bcr-Abl kinase, a mutant kinase present only in tumor cells that have undergone a translocation between chromosomes 9 and 22 (see Figure 14.33). Before initiating treatment with imatinib, we could sequence the DNA of the tumor cells and determine (a) whether this translocation has taken place and (b) whether the sequence of bcr-abl carries any mutations that would render the kinase resistant to imatinib. If the translocation has not taken place or if the gene carries resistance mutations, then imatinib would likely not be an effective treatment for the patient carrying this particular tumor.

12. Sildenafil increases cGMP levels by inhibiting the phosphodiesterase-mediated breakdown of cGMP to GMP. Intracellular cGMP levels can also be increased by activating its synthesis. This activation can be achieved with the use of NO donors (such as sodium nitroprusside and nitroglycerin) or compounds that activate guanylate cyclase activity. Drugs that act by the latter mechanism are currently in clinical trials.

13. A reasonable mechanism would be an oxidative deamination following an overall mechanism similar to that in Figure 36.9, with release of ammonia.



14. $K_{\rm I} \approx 0.3$ nM. IC₅₀ ≈ 2.0 nM. Yes, compound A should be effective when taken orally because 400 nM is much greater than the estimated values of $K_{\rm I}$ and IC₅₀.

This page intentionally left blank

Chapter 2

Where to Start

- Service, R. F. 2008. Problem solved*(*sort of) (a brief review of protein folding). *Science* 321:784–786.
- Doolittle, R. F. 1985. Proteins. Sci. Am. 253(4):88-99.
- Richards, F. M. 1991. The protein folding problem. *Sci. Am.* 264(1): 54–57.
- Weber, A. L., and Miller, S. L. 1981. Reasons for the occurrence of the twenty coded protein amino acids. J. Mol. Evol. 17:273–284.

Books

- Petsko, G. A., and Ringe, D. 2004. Protein Structure and Function. New Science Press.
- Tanford, C., and Reynolds, J. 2004. Nature's Robots: A History of Proteins. Oxford.
- Branden, C., and Tooze, J. 1999. Introduction to Protein Structure (2d ed.). Garland.
- Creighton, T. E. 1992. Proteins: Structures and Molecular Principles (2d ed.). W. H. Freeman and Company.

Conformation of Proteins

- Smock, R. G., and Gierasch, L. M. 2009. Sending signals dynamically. Science 324:198–203.
- Tokuriki, N., and Tawfik, D. S. 2009. Protein dynamism and evolvability. Science 324:203–207.
- Pace, C. N., Grimsley, G. R., and Scholtz, J. M. 2009. Protein ionizable groups: pK values and their contribution to protein stability and solubility. J. Biol. Chem. 284:13285–13289.
- Ronald Breslow, R., and Cheng, Z.-L. 2009. On the origin of terrestrial homochirality for nucleosides and amino acids. *Proc. Natl. Acad. Sci. U.S.A.* 106:9144–9146.

Secondary Structure

- Shoulders, M. D., and Raines, R. T. 2009. Collagen structure and stability. Annu. Rev. Biochem. 78:929–58.
- O'Neil, K. T., and DeGrado, W. F. 1990. A thermodynamic scale for the helix-forming tendencies of the commonly occurring amino acids. *Science* 250:646–651.
- Zhang, C., and Kim, S. H. 2000. The anatomy of protein beta-sheet topology. J. Mol. Biol. 299:1075–1089.
- Regan, L. 1994. Protein structure: Born to be beta. Curr. Biol. 4:656-658.
- Srinivasan, R., and Rose, G. D. 1999. A physical basis for protein secondary structure. Proc. Natl. Acad. Sci. U.S.A. 96:14258–14263.

Intrinsically Unstructured Proteins

- Galea, C. A., Wang, Y., Sivakolundu, S. G., and Kriwacki, R. W. 2008. Regulation of cell division by intrinsically unstructured proteins: Intrinsic flexibility, modularity, and signaling conduits. *Biochemistry* 47:7598–7609.
- Raychaudhuri, S., Dey, S., Bhattacharyya, N. P., and Mukhopadhyay, D. 2009. The role of intrinsically unstructured proteins in neurodegenerative diseases. *PLoS One* 4:e5566.
- Tompa, P., and Fuxreiter, M. 2008. Fuzzy complexes: Polymorphism and structural disorder in protein-protein interactions. *Trends Biochem. Sci.* 33:2–8.
- Tuinstra, R. L., Peterson, F. C., Kutlesa, E. S., Elgin, S., Kron, M. A., and Volkman, B. F. 2008. Interconversion between two unrelated protein folds in the lymphotactin native state. *Proc. Natl. Acad. Sci.* U.S.A. 105:5057–5062.

Domains

Jin, J., Xie, X., Chen, C., Park, J. G., Stark, C., James, D. A., Olhovsky, M., Linding, R., Mao, Y., and Pawson, T. 2009. Eukaryotic protein domains as functional units of cellular evolution. *Sci. Signal.* 2:ra76.

- Bennett, M. J., Choe, S., and Eisenberg, D. 1994. Domain swapping: Entangling alliances between proteins. *Proc. Natl. Acad. Sci. U.S.A.* 91:3127–3131.
- Bergdoll, M., Eltis, L. D., Cameron, A. D., Dumas, P., and Bolin, J. T. 1998. All in the family: Structural and evolutionary relationships among three modular proteins with diverse functions and variable assembly. *Protein Sci.* 7:1661–1670.
- Hopfner, K. P., Kopetzki, E., Kresse, G. B., Bode, W., Huber, R., and Engh, R. A. 1998. New enzyme lineages by subdomain shuffling. *Proc. Natl. Acad. Sci. U.S.A.* 95:9813–9818.
- Ponting, C. P., Schultz, J., Copley, R. R., Andrade, M. A., and Bork, P. 2000. Evolution of domain families. Adv. Protein Chem. 54:185–244.

Protein Folding

- Caughey, B., Baron, G. S., Chesebro, B., and Jeffrey, M. 2009. Getting a grip on prions: Oligomers, amyloids, and pathological membrane interactions. *Annu. Rev. Biochem.* 78:177–204.
- Cobb, N. J., and Surewicz, W. K. 2009. Prion diseases and their biochemical mechanisms. *Biochemistry* 48:2574–2585.
- Daggett, V., and Fersht, A. R. 2003. Is there a unifying mechanism for protein folding? *Trends Biochem. Sci.* 28:18–25.
- Selkoe, D. J. 2003. Folding proteins in fatal ways. Nature 426:900-904.
- Anfinsen, C. B. 1973. Principles that govern the folding of protein chains. Science 181:223–230.
- Baldwin, R. L., and Rose, G. D. 1999. Is protein folding hierarchic? I. Local structure and peptide folding. *Trends Biochem. Sci.* 24:26–33.
- Baldwin, R. L., and Rose, G. D. 1999. Is protein folding hierarchic? II. Folding intermediates and transition states. *Trends Biochem.* Sci. 24:77–83.
- Kuhlman, B., Dantas, G., Ireton, G. C., Varani, G., Stoddard, B. L., and Baker, D. 2003. Design of a novel globular protein with atomiclevel accuracy. *Science* 302:1364–1368.
- Staley, J. P., and Kim, P. S. 1990. Role of a subdomain in the folding of bovine pancreatic trypsin inhibitor. *Nature* 344:685–688.

Covalent Modification of Proteins

- Tarrant, M. K., and Cole, P. A. 2009. The chemical biology of protein phosphorylation. Annu. Rev. Biochem. 78:797–825.
- Krishna, R. G., and Wold, F. 1993. Post-translational modification of proteins. Adv. Enzymol. Relat. Areas. Mol. Biol. 67:265–298.
- Aletta, J. M., Cimato, T. R., and Ettinger, M. J. 1998. Protein methylation: A signal event in post-translational modification. *Trends Biochem. Sci.* 23:89–91.
- Tsien, R. Y. 1998. The green fluorescent protein. Annu. Rev. Biochem. 67:509–544.

Chapter 3

Where to Start

- Sanger, F. 1988. Sequences, sequences, sequences. Annu. Rev. Biochem. 57:1–28.
- Merrifield, B. 1986. Solid phase synthesis. Science 232:341-347.
- Hunkapiller, M. W., and Hood, L. E. 1983. Protein sequence analysis: Automated microsequencing. *Science* 219:650–659.
- Milstein, C. 1980. Monoclonal antibodies. Sci. Am. 243(4):66-74.
- Moore, S., and Stein, W. H. 1973. Chemical structures of pancreatic ribonuclease and deoxyribonuclease. *Science* 180:458–464.

Books

- Methods in Enzymology. Academic Press.
- Wilson, K., and Walker, J. (Eds.). 2000. Principles and Techniques of Practical Biochemistry (5th ed.). Cambridge University Press.
- Van Holde, K. E., Johnson, W. C., and Ho, P.-S. 1998. Principles of Physical Biochemistry. Prentice Hall.

- Wilkins, M. R., Williams, K. L., Appel, R. D., and Hochstrasser, D. F. 1997. Proteome Research: New Frontiers in Functional Genomics (Principles and Practice). Springer Verlag.
- Johnstone, R. A. W. 1996. Mass Spectroscopy for Chemists and Biochemists (2d ed.). Cambridge University Press.
- Kyte, J. 1994. Structure in Protein Chemistry. Garland.
- Creighton, T. E. 1993. Proteins: Structure and Molecular Properties (2d ed.). W. H. Freeman and Company.
- Cantor, C. R., and Schimmel, P. R. 1980. *Biophysical Chemistry*. W. H. Freeman and Company.

Protein Purification and Analysis

- Blackstock, W. P., and Weir, M. P. 1999. Proteomics: Quantitative and physical mapping of cellular proteins. *Trends Biotechnol.* 17:121–127.
- Deutscher, M. (Ed.). 1997. Guide to Protein Purification. Academic Press. Dunn, M. J. 1997. Quantitative two-dimensional gel electrophoresis: From proteins to proteomes. Biochem. Soc. Trans. 25:248–254.
- Scopes, R. K., and Cantor, C. 1994. Protein Purification: Principles and Practice (3d ed.). Springer Verlag.
- Aebersold, R., Pipes, G. D., Wettenhall, R. E., Nika, H., and Hood, L. E. 1990. Covalent attachment of peptides for high sensitivity solid-phase sequence analysis. *Anal. Biochem.* 187:56–65.

Ultracentrifugation and Mass Spectrometry

- Steen, H., and Mann, M. 2004. The ABC's (and XYZ's) of peptide sequencing. Nat. Rev. Mol. Cell Biol. 5:699–711.
- Glish, G. L., and Vachet, R. W. 2003. The basics of mass spectrometry in the twenty-first century. Nat. Rev. Drug Discovery 2:140–150.
- Li, L., Garden, R. W., and Sweedler, J. V. 2000. Single-cell MALDI: A new tool for direct peptide profiling. *Trends Biotechnol.* 18: 151–160.
- Yates, J. R., 3d. 1998. Mass spectrometry and the age of the proteome. J. Mass Spectrom. 33:1–19.
- Pappin, D. J. 1997. Peptide mass fingerprinting using MALDI-TOF mass spectrometry. *Methods Mol. Biol.* 64:165–173.
- Schuster, T. M., and Laue, T. M. 1994. Modern Analytical Ultracentrifugation. Springer Verlag.
- Arnott, D., Shabanowitz, J., and Hunt, D. F. 1993. Mass spectrometry of proteins and peptides: Sensitive and accurate mass measurement and sequence analysis. *Clin. Chem.* 39:2005–2010.
- Chait, B. T., and Kent, S. B. H. 1992. Weighing naked proteins: Practical, high-accuracy mass measurement of peptides and proteins. *Science* 257:1885–1894.
- Edmonds, C. G., Loo, J. A., Loo, R. R., Udseth, H. R., Barinaga, C. J., and Smith, R. D. 1991. Application of electrospray ionization mass spectrometry and tandem mass spectrometry in combination with capillary electrophoresis for biochemical investigations. *Biochem. Soc. Trans.* 19:943–947.
- Jardine, I. 1990. Molecular weight analysis of proteins. *Methods* Enzymol. 193:441–455.

Proteomics

- Yates, J. R., 3d. 2004. Mass spectral analysis in proteomics. Annu. Rev. Biophys. Biomol. Struct. 33:297–316.
- Weston, A. D., and Hood, L. 2004. Systems biology, proteomics, and the future of health care: Toward predictive, preventative, and personalized medicine. J. Proteome Res. 3:179–196.
- Pandey, A., and Mann, M. 2000. Proteomics to study genes and ge-nomes. *Nature* 405:837–846.
- Dutt, M. J., and Lee, K. H. 2000. Proteomic analysis. Curr. Opin. Biotechnol. 11:176–179.
- Rout, M. P., Aitchison, J. D., Suprapto, A., Hjertaas, K., Zhao, Y., and Chait, B. T. 2000. The yeast nuclear pore complex: Composition, architecture, and transport mechanism. J. Cell Biol. 148:635–651.

X-ray Crystallography and NMR Spectroscopy

Rhodes, G. 2006. Crystallography Made Crystal Clear. Elsevier/ Academic Press.

- Moffat, K. 2003. The frontiers of time-resolved macromolecular crystallography: Movies and chirped X-ray pulses. *Faraday Discuss.* 122:65–88.
- Bax, A. 2003. Weak alignment offers new NMR opportunities to study protein structure and dynamics. *Protein Sci.* 12:1–16.
- Wery, J. P., and Schevitz, R. W. 1997. New trends in macromolecular x-ray crystallography. Curr. Opin. Chem. Biol. 1:365–369.
- Glusker, J. P. 1994. X-ray crystallography of proteins. Methods Biochem. Anal. 37:1–72.
- Clore, G. M., and Gronenborn, A. M. 1991. Structures of larger proteins in solution: Three- and four-dimensional heteronuclear NMR spectroscopy. *Science* 252:1390–1399.
- Wüthrich, K. 1989. Protein structure determination in solution by nuclear magnetic resonance spectroscopy. *Science* 243:45–50.
- Wüthrich, K. 1986. NMR of Proteins and Nucleic Acids. Wiley-Interscience.

Monoclonal Antibodies and Fluorescent Molecules

Immunology Today. 2000. Volume 21, issue 8.

- Tsien, R. Y. 1998. The green fluorescent protein. Annu. Rev. Biochem. 67:509–544.
- Kendall, J. M., and Badminton, M. N. 1998. Aequorea victoria bioluminescence moves into an exciting era. Trends Biotechnol. 16:216–234.
- Goding, J. W. 1996. Monoclonal Antibodies: Principles and Practice. Academic Press.
- Köhler, G., and Milstein, C. 1975. Continuous cultures of fused cells secreting antibody of predefined specificity. *Nature* 256:495–497.

Chemical Synthesis of Proteins

- Bang, D., Chopra, N., and Kent, S. B. 2004. Total chemical synthesis of crambin. J. Am. Chem. Soc. 126:1377–1383.
- Dawson, P. E., and Kent, S. B. 2000. Synthesis of native proteins by chemical ligation. Annu. Rev. Biochem. 69:923–960.
- Mayo, K. H. 2000. Recent advances in the design and construction of synthetic peptides: For the love of basics or just for the technology of it. *Trends Biotechnol.* 18:212–217.

Chapter 4

Where to Start

- Felsenfeld, G. 1985. DNA. Sci. Am. 253(4):58-67.
- Darnell, J. E., Jr. 1985. RNA. Sci. Am. 253(4):68-78.
- Dickerson, R. E. 1983. The DNA helix and how it is read. Sci. Am. 249(6):94–111.
- Crick, F. H. C. 1954. The structure of the hereditary material. Sci. Am. 191(4): 54–61.
- Chambon, P. 1981. Split genes. Sci. Am. 244(5):60-71.
- Watson, J. D., and Crick, F. H. C. 1953. Molecular structure of nucleic acids: A structure for deoxyribose nucleic acid. *Nature* 171:737–738.
- Watson, J. D., and Crick, F. H. C. 1953. Genetic implications of the structure of deoxyribonucleic acid. *Nature* 171:964–967.
- Meselson, M., and Stahl, F. W. 1958. The replication of DNA in *Escherichia coli. Proc. Natl. Acad. Sci. U.S.A.* 44:671–682.

Books

- Bloomfield, V. A., Crothers, D. M., Tinoco, I., and Hearst, J. 2000. Nucleic Acids: Structures, Properties, and Functions. University Science Books.
- Singer, M., and Berg, P. 1991. Genes and Genomes: A Changing Perspective. University Science Books.
- Lodish, H., Berk, A., Kaiser, C. A., Krieger, M., Scott, M. P., Bretscher, A., Ploegh, H., and Matsudaira, P. 2007. *Molecular Cell Biology* (6th ed.). W. H. Freeman and Company.

Selected Readings

Lewin, B. 2007. Genes IX. Jones and Bartlett.

Watson, J. D., Baker, T. A., Bell, S. P., Gann, A., Levine, M., and Losick, R. 2007. *Molecular Biology of the Gene* (6th ed.). Benjamin Cummings.

DNA Structure

Neidle, S. 2007. Principles of Nucleic Acid Structure. Academic Press.

Dickerson, R. E., Drew, H. R., Conner, B. N., Wing, R. M., Fratini, A. V., and Kopka, M. L. 1982. The anatomy of A-, B-, and Z-DNA. Science 216:475–485.

Sinden, R. R. 1994. DNA Structure and Function. Academic Press.

DNA Replication

- Lehman, I. R. 2003. Discovery of DNA polymerase. J. Biol. Chem. 278:34733–34738.
- Hübscher, U., Maga, G., and Spardari, S. 2002. Eukaryotic DNA polymerases. Annu. Rev. Biochem. 71:133–163.
- Hübscher, U., Nasheuer, H.-P., and Syväoja, J. E. 2000. Eukaryotic DNA polymerases: A growing family. *Trends Biochem. Sci.* 25:143–147.
- Brautigam, C. A., and Steitz, T. A. 1998. Structural and functional insights provided by crystal structures of DNA polymerases and their substrate complexes. *Curr. Opin. Struct. Biol.* 8:54–63.
- Kornberg, A., and Baker, T. A. 1992. DNA Replication (2d ed.). W. H. Freeman and Company.

Discovery of Messenger RNA

- Jacob, F., and Monod, J. 1961. Genetic regulatory mechanisms in the synthesis of proteins. J. Mol. Biol. 3:318–356.
- Brenner, S., Jacob, F., and Meselson, M. 1961. An unstable intermediate carrying information from genes to ribosomes for protein synthesis. *Nature* 190:576–581.
- Hall, B. D., and Spiegelman, S. 1961. Sequence complementarity of T2-DNA and T2-specific RNA. Proc. Natl. Acad. Sci. U.S.A. 47:137–146.

Genetic Code

- Koonin, E. V., and Novozhilov, A. S. 2009. Origin and evolution of the genetic code: The universal enigma. *IUBMB Life* 61:99–111.
- Yarus, M., Caporaso, J. G., and Knight, R. 2005. Origins of the genetic code: The escaped triplet theory. Annu. Rev. Biochem. 74:179–198.
- Freeland, S. J., and Hurst, L. D. 2004. Evolution encoded. Sci. Am. 290(4):84-91.
- Crick, F. H. C., Barnett, L., Brenner, S., and Watts-Tobin, R. J. 1961. General nature of the genetic code for proteins. *Nature* 192:1227–1232.

Woese, C. R. 1967. The Genetic Code. Harper & Row.

Knight, R. D., Freeland, S. J., and Landweber L. F. 1999. Selection, history and chemistry: The three faces of the genetic code. *Trends Biochem. Sci.* 24(6):241–247.

Introns, Exons, and Split Genes

- Liu, M., and Grigoriev, A. 2004. Protein domains correlate strongly with exons in multiple eukaryotic genomes—evidence of exon shuffling? *Trends Genet.* 20:399–403.
- Dorit, R. L., Schoenbach, L., and Gilbert, W. 1990. How big is the universe of exons? *Science* 250:1377–1382.
- Cochet, M., Gannon, F., Hen, R., Maroteaux, L., Perrin, F., and Chambon, P. 1979. Organization and sequence studies of the 17-piece chicken conalbumin gene. *Nature* 282:567–574.
- Tilghman, S. M., Tiemeier, D. C., Seidman, J. G., Peterlin, B. M., Sullivan, M., Maizel, J. V., and Leder, P. 1978. Intervening sequence of DNA identified in the structural portion of a mouse β-globin gene. *Proc. Natl. Acad. Sci. U.S.A.* 75:725–729.

Reminiscences and Historical Accounts

Nirenberg, M. 2004. Deciphering the genetic code—a personal account. Trends Biochem. Sci. 29:46–54.

- Clayton, J., and Dennis, C. (Eds.). 2003. 50 Years of DNA. Palgrave Macmillan.
- Watson, J. D. 1968. The Double Helix. Atheneum.
- McCarty, M. 1985. The Transforming Principle: Discovering That Genes Are Made of DNA. Norton.
- Cairns, J., Stent, G. S., and Watson, J. D. 2000. Phage and the Origins of Molecular Biology. Cold Spring Harbor Laboratory.
- Olby, R. 1974. The Path to the Double Helix. University of Washington Press.
- Portugal, F. H., and Cohen, J. S. 1977. A Century of DNA: A History of the Discovery of the Structure and Function of the Genetic Substance. MIT Press.
- Judson, H. F. 1996. The Eighth Day of Creation. Cold Spring Harbor Laboratory.
- Sayre, A. 2000. Rosalind Franklin and DNA. Norton.

Chapter 5

Where to Start

- Berg, P. 1981. Dissections and reconstructions of genes and chromosomes. Science 213:296–303.
- Gilbert, W. 1981. DNA sequencing and gene structure. Science 214:1305–1312.
- Sanger, F. 1981. Determination of nucleotide sequences in DNA. Science 214:1205–1210.
- Mullis, K. B. 1990. The unusual origin of the polymerase chain reaction. *Sci. Am.* 262(4):56–65.

Books on Recombinant DNA Technology

- Watson, J. D., Myers, R. M., Caudy, A. A., and Witkowski, J. 2007. *Recombinant DNA: Genes and Genomes* (3d ed.). W. H. Freeman and Company.
- Grierson, D. (Ed.). 1991. Plant Genetic Engineering. Chapman and Hall.
- Mullis, K. B., Ferré, F., and Gibbs, R. A. (Eds.). 1994. The Polymerase Chain Reaction. Birkhaüser.
- Russel, D., Sambrook, J., and Russel, D. 2000. *Molecular Cloning: A Laboratory Manual* (3d ed.). Cold Spring Harbor Laboratory Press.
- Ausubel, F. M., Brent, R., Kingston, R. E., and Moore, D. D. (Eds.). 1999. Short Protocols in Molecular Biology: A Compendium of Methods from Current Protocols in Molecular Biology. Wiley.
- Birren, B., Green, E. D., Klapholz, S., Myers, R. M., Roskams, J., Riethamn, H., and Hieter, P. (Eds.). 1999. *Genome Analysis* (vols. 1–4). Cold Spring Harbor Laboratory Press.
- Methods in Enzymology. Academic Press. [Many volumes in this series deal with recombinant DNA technology.]

DNA Sequencing and Synthesis

- Hunkapiller, T., Kaiser, R. J., Koop, B. F., and Hood, L. 1991. Large-scale and automated DNA sequence determination. *Science* 254:59–67.
- Sanger, F., Nicklen, S., and Coulson, A. R. 1977. DNA sequencing with chain-terminating inhibitors. *Proc. Natl. Acad. Sci. U.S.A.* 74:5463–5467.
- Maxam, A. M., and Gilbert, W. 1977. A new method for sequencing DNA. Proc. Natl. Acad. Sci. U.S.A. 74:560–564.
- Smith, L. M., Sanders, J. Z., Kaiser, R. J., Hughes, P., Dodd, C., Connell, C. R., Heiner, C., Kent, S. B. H., and Hood, L. E. 1986. Fluorescence detection in automated DNA sequence analysis. *Nature* 321:674–679.
- Pease, A. C., Solas, D., Sullivan, E. J., Cronin, M. T., Holmes, C. P., and Fodor, S. P. A. 1994. Light-generated oligonucleotide arrays for rapid DNA sequence analysis. *Proc. Natl. Acad. Sci. U.S.A.* 91:5022–5026.
- Venter, J. C., Adams, M. D., Sutton, G. G., Kerlavage, A. R., Smith, H. O., and Hunkapiller, M. 1998. Shotgun sequencing of the human genome. *Science* 280:1540–1542.

Mardis, E. R. 2008. Next-generation DNA sequencing methods. Annu. Rev. Genomics Hum. Genet. 9:387–402.

Polymerase Chain Reaction

- Arnheim, N., and Erlich, H. 1992. Polymerase chain reaction strategy. Annu. Rev. Biochem. 61:131–156.
- Kirby, L. T. (Ed.). 1997. DNA Fingerprinting: An Introduction. Stockton Press.
- Eisenstein, B. I. 1990. The polymerase chain reaction: A new method for using molecular genetics for medical diagnosis. N. Engl. J. Med. 322:178–183.
- Foley, K. P., Leonard, M. W., and Engel, J. D. 1993. Quantitation of RNA using the polymerase chain reaction. *Trends Genet*. 9:380–386. Pääbo, S. 1993. Ancient DNA. *Sci. Am.* 269(5):86–92.
- Paabo, S. 1993. Ancient DNA. Sci. Am. 209(5):80-92.
- Hagelberg, E., Gray, I. C., and Jeffreys, A. J. 1991. Identification of the skeletal remains of a murder victim by DNA analysis. *Nature* 352:427–429.
- Lawlor, D. A., Dickel, C. D., Hauswirth, W. W., and Parham, P. 1991. Ancient HLA genes from 7500-year-old archaeological remains. *Nature* 349:785–788.
- Krings, M., Geisert, H., Schmitz, R. W., Krainitzki, H., and Pääbo, S. 1999. DNA sequence of the mitochondrial hypervariable region II for the Neanderthal type specimen. *Proc. Natl. Acad. Sci. U.S.A.* 96:5581–5585.
- Ovchinnikov, I. V., Götherström, A., Romanova, G. P., Kharitonov, V. M., Lidén, K., and Goodwin, W. 2000. Molecular analysis of Neanderthal DNA from the northern Caucasus. *Nature* 404:490–493.

Genome Sequencing

- International Human Genome Sequencing Consortium. 2004. Finishing the euchromatic sequence of the human genome. *Nature* 431:931–945.
- Lander, E. S., Linton, L. M., Birren, B., Nusbaum, C., Zody, M. C., Baldwin, J., Devon, K., Dewar, K., Doyle, M., FitzHugh, W., et al. 2001. Initial sequencing and analysis of the human genome. *Nature* 409:860–921.
- Venter, J. C., Adams, M. D., Myers, E. W., Li, P. W., Mural, R. J., Sutton, G. G., Smith, H. O., Yandell, M., Evans, C. A., Holt, R. A., et al. 2001. The sequence of the human genome. *Science* 291:1304–1351.
- Waterston, R. H., Lindblad-Toh, K., Birney, E., Rogers, J., Abril, J. F., Agarwal, P., Agarwala, R., Ainscough, R., Alexandersson, M., An, P., et al. 2002. Initial sequencing and comparative analysis of the mouse genome. *Nature* 420:520–562.
- Koonin, E. V. 2003. Comparative genomics, minimal gene-sets and the last universal common ancestor. Nat. Rev. Microbiol. 1:127–236.
- Gilligan, P., Brenner, S., and Venkatesh, B. 2002. Fugu and human sequence comparison identifies novel human genes and conserved non-coding sequences. *Gene* 294:35–44.
- Enard, W., and Pääbo, S. 2004. Comparative primate genomics. Annu. Rev. Genomics Hum. Genet. 5:351–378.

Quantitative PCR and DNA Arrays

- Duggan, D. J., Bittner, J. M., Chen, Y., Meltzer, P., and Trent, J. M. 1999. Expression profiling using cDNA microarrays. *Nat. Genet.* 21:10–14.
- Golub, T. R., Slonim, D. K., Tamayo, P., Huard, C., Gaasenbeek, M., Mesirov, J. P., Coller, H., Loh, M. L., Downing, J. R., Caligiuri, M. A., Bloomfield, C. D., and Lander, E. S. 1999. Molecular classification of cancer: Class discovery and class prediction by gene expression monitoring. *Science* 286:531–537.
- Perou, C. M., Sørlie, T., Eisen, M. B., van de Rijn, M., Jeffery, S. S., Rees, C. A., Pollack, J. R., Ross, D. T., Johnsen, H., Akslen, L. A., Fluge, Ø., Pergamenschikov, A., Williams, C., Zhu, S. X., Lønning, P. E., Børresen-Dale, A.-L., Brown, P. O., and Botstein, D. 2000. Molecular portraits of human breast tumours. *Nature* 406:747–752.
- Walker, N. J. 2002. A technique whose time has come. Science 296:557–559.

Introduction of Genes into Animal Cells

Anderson, W. F. 1992. Human gene therapy. Science 256:808-813.

- Friedmann, T. 1997. Overcoming the obstacles to gene therapy. Sci. Am. 277(6):96–101.
- Blaese, R. M. 1997. Gene therapy for cancer. Sci. Am. 277 (6): 111-115.
- Brinster, R. L., and Palmiter, R. D. 1986. Introduction of genes into the germ lines of animals. *Harvey Lect.* 80:1–38.
- Capecchi, M. R. 1989. Altering the genome by homologous recombination. Science 244:1288–1292.
- Hasty, P., Bradley, A., Morris, J. H., Edmondson, D. G., Venuti, J. M., Olson, E. N., and Klein, W. H. 1993. Muscle deficiency and neonatal death in mice with a targeted mutation in the myogenin gene. *Nature* 364:501–506.
- Parkmann, R., Weinberg, K., Crooks, G., Nolta, J., Kapoor, N., and Kohn, D. 2000. Gene therapy for adenosine deaminase deficiency. *Annu. Rev. Med.* 51:33–47.

RNA Interference

- Rana, T. M. 2007. Illuminating the silence: Understanding the structure and function of small RNAs. Nat. Rev. Mol. Cell Biol. 8:23–36.
- Novina, C. D., and Sharp, P. A. 2004. The RNAi revolution. *Nature* 430:161–164.
- Hannon, G. J., and Rossi, J. J. 2004. Unlocking the potential of the human genome with RNA interference. *Nature* 431:371–378.
- Meister, G., and Tuschl, T. 2004. Mechanisms of gene silencing by double-stranded RNA. *Nature* 431:343–349.
- Elbashir, S. M., Harborth, J., Lendeckel, W., Yalcin, A., Weber, K., and Tuschl, T. 2001. Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells. *Nature* 411:494–498.
- Fire, A., Xu, S., Montgomery, M. K., Kostas, S. A., Driver, S. E., and Mello, C. C. 1998. Potent and specific genetic interference by doublestranded RNA in *Caenorhabditis elegans*. *Nature* 391:806–811.

Genetic Engineering of Plants

- Gasser, C. S., and Fraley, R. T. 1992. Transgenic crops. Sci. Am. 266(6):62-69.
- Gasser, C. S., and Fraley, R. T. 1989. Genetically engineering plants for crop improvement. Science 244:1293–1299.
- Shimamoto, K., Terada, R., Izawa, T., and Fujimoto, H. 1989. Fertile transgenic rice plants regenerated from transformed protoplasts. *Nature* 338:274–276.
- Chilton, M.-D. 1983. A vector for introducing new genes into plants. Sci. Am. 248(6):50–59.
- Hansen, G., and Wright, M. S. 1999. Recent advances in the transformation of plants. *Trends Plant Sci.* 4:226–231.
- Hammond, J. 1999. Overview: The many uses of transgenic plants. Curr. Top. Microbiol. Immunol. 240:1–20.
- Finer, J. J., Finer, K. R., and Ponappa, T. 1999. Particle bombardment mediated transformation. *Curr. Top. Microbiol. Immunol.* 240:60–80.

Amyotrophic Lateral Sclerosis

- Siddique, T., Figlewicz, D. A., Pericak-Vance, M. A., Haines, J. L., Rouleau, G., Jeffers, A. J., Sapp, P., Hung, W.-Y., Bebout, J., McKenna-Yasek, D., et al. 1991. Linkage of a gene causing familial amyotrophic lateral sclerosis to chromosome 21 and evidence of genetic-locus heterogeneity. *New Engl. J. Med.* 324:1381–1384.
- Rosen, D. R., Siddique, T., Patterson, D., Figlewicz, D. A., Sapp, P., Hentati, A., Donaldson, D., Goto, J., O'Regan, J. P., Deng, H.-X., et al. 1993. Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis. *Nature* 362:59–62.
- Gurney, M. E., Pu, H., Chiu, A. Y., Dal Canto, M. C., Polchow, C. Y., Alexander, D. D., Caliendo, J., Hentati, A., Kwon, Y. W., Deng, H.-X., Chen, W., Zhai, P., Sufit, R. L., and Siddique, T. 1994. Motor neuron degeneration in mice that express a human Cu,Zn superoxide dismutase mutation. *Science* 264:1772–1774.

Borchelt, D. R., Lee, M. K., Slunt, H. S., Guarnieri, M., Xu, Z.-S., Wong, P. C. Brown, R. H., Jr., Price, D. L., Sisodia, S. S., and Cleveland, D. W. 1994. Superoxide dismutase 1 with mutations linked to familial amyotrophic lateral sclerosis possesses significant activity. *Proc. Natl. Acad. Sci. U.S.A.* 91:8292–8296.

Chapter 6

Books

Claverie, J.-M., and Notredame, C. 2003. *Bioinformatics for Dummies*. Wiley.

Pevsner, J. 2003. Bioinformatics and Functional Genomics. Wiley-Liss. Doolittle, R. F. 1987. Of URFS and ORFS. University Science Books.

Sequence Alignment

- Schaffer, A. A., Aravind, L., Madden, T. L., Shavirin, S., Spouge, J. L., Wolf, Y. I., Koonin, E. V., and Altschul, S. F. 2001. Improving the accuracy of PSI-BLAST protein database searches with composition-based statistics and other refinements. *Nucleic Acids Res.* 29:2994–3005.
- Henikoff, S., and Henikoff, J. G. 1992. Amino acid substitution matrices from protein blocks. Proc. Natl. Acad. Sci. U.S.A. 89:10915–10919.
- Johnson, M. S., and Overington, J. P. 1993. A structural basis for sequence comparisons: An evaluation of scoring methodologies. J. Mol. Biol. 233:716–738.
- Eddy, S. R. 2004. Where did the BLOSUM62 alignment score matrix come from? Nat. Biotechnol. 22:1035–1036.
- Aravind, L., and Koonin, E. V. 1999. Gleaning non-trivial structural, functional and evolutionary information about proteins by iterative database searches. J. Mol. Biol. 287:1023–1040.
- Altschul, S. F., Madden, T. L., Schaffer, A. A., Zhang, J., Zhang, Z., Miller, W., and Lipman, D. J. 1997. Gapped BLAST and PSI-BLAST: A new generation of protein database search programs. *Nucleic Acids Res.* 25:3389–3402.

Structure Comparison

- Orengo, C. A., Bray, J. E., Buchan, D. W., Harrison, A., Lee, D., Pearl, F. M., Sillitoe, I., Todd, A. E., and Thornton, J. M. 2002. The CATH protein family database: A resource for structural and functional annotation of genomes. *Proteomics* 2:11–21.
- Bashford, D., Chothia, C., and Lesk, A. M. 1987. Determinants of a protein fold: Unique features of the globin amino acid sequences. *J. Mol. Biol.* 196:199–216.
- Harutyunyan, E. H., Safonova, T. N., Kuranova, I. P., Popov, A. N., Teplyakov, A. V., Obmolova, G. V., Rusakov, A. A., Vainshtein, B. K., Dodson, G. G., Wilson, J. C., et al. 1995. The structure of deoxy- and oxy-leghaemoglobin from lupin. J. Mol. Biol. 251:104–115.
- Flaherty, K. M., McKay, D. B., Kabsch, W., and Holmes, K. C. 1991. Similarity of the three-dimensional structures of actin and the ATPase fragment of a 70-kDa heat shock cognate protein. *Proc. Natl. Acad. Sci. U.S.A.* 88:5041–5045.
- Murzin, A. G., Brenner, S. E., Hubbard, T., and Chothia, C. 1995. SCOP: A structural classification of proteins database for the investigation of sequences and structures. J. Mol. Biol. 247: 536–540.
- Hadley, C., and Jones, D. T. 1999. A systematic comparison of protein structure classification: SCOP, CATH and FSSP. Struct. Fold. Des. 7:1099–1112.

Domain Detection

Marchler-Bauer, A., Anderson, J. B., DeWeese-Scott, C., Fedorova, N. D., Geer, L. Y., He, S., Hurwitz, D. I., Jackson, J. D., Jacobs, A. R., Lanczycki, C. J., Liebert, C. A., Liu, C., Madej, T., Marchler, G. H., Mazumder, R., Nikolskaya, A. N., Panchenko, A. R., Rao, B. S., Shoemaker, B. A., Simonyan, V., Song, J. S., Thiessen, P. A., Vasudevan, S., Wang, Y., Yamashita, R. A., Yin, J. J., and Bryant, S. H. 2003. CDD: A curated Entrez database of conserved domain alignments. *Nucleic Acids Res.* 31:383–387.

- Ploegman, J. H., Drent, G., Kalk, K. H., and Hol, W. G. 1978. Structure of bovine liver rhodanese I: Structure determination at 2.5 Å resolution and a comparison of the conformation and sequence of its two domains. J. Mol. Biol. 123:557–594.
- Nikolov, D. B., Hu, S. H., Lin, J., Gasch, A., Hoffmann, A., Horikoshi, M., Chua, N. H., Roeder, R. G., and Burley, S. K. 1992. Crystal structure of TFIID TATA-box binding protein. *Nature* 360: 40–46.
- Doolittle, R. F. 1995. The multiplicity of domains in proteins. Annu. Rev. Biochem. 64:287–314.
- Heger, A., and Holm, L. 2000. Rapid automatic detection and alignment of repeats in protein sequences. *Proteins* 41:224–237.

Evolutionary Trees

- Wolf, Y. I., Rogozin, I. B., Grishin, N. V., and Koonin, E. V. 2002. Genome trees and the tree of life. *Trends Genet*. 18:472–479.
- Doolittle, R. F. 1992. Stein and Moore Award address. Reconstructing history with amino acid sequences. *Protein Sci.* 1:191–200.
- Zuckerkandl, E., and Pauling, L. 1965. Molecules as documents of evolutionary history. J. Theor. Biol. 8:357-366.

Ancient DNA

- Green, R. E., Malaspinas, A.-S., Krause, J., Briggs, A. W., Johnson, P. L. F., Uhler, C., Meyer, M., Good, J. M., Maricic, T., Stenzel, U., Prüfer, K., Siebauer, M., Burbano, H. A., Ronan, M., Rothberg, J. M., Egholm, M., Rudan, P., Brajković, D., Kućan, Ž., Gušić, I., Wikström, M., Laakkonen, L., Kelso, J., Slatkin, M., and Pääbo, S. 2008. A complete Neandertal mitochondrial genome sequence determined by high-throughput sequencing. *Cell* 134:416–426.
- Pääbo, S., Poinar, H., Serre, D., Jaenicke-Despres, V., Hebler, J., Rohland, N., Kuch, M., Krause, J., Vigilant, L., and Hofreiter, M. 2004. Genetic analyses from ancient DNA. Annu. Rev. Genet. 38:645–679.
- Krings, M., Stone, A., Schmitz, R. W., Krainitzki, H., Stoneking, M., and Pääbo, S. 1997. Neandertal DNA sequences and the origin of modern humans. *Cell* 90:19–30.
- Krings, M., Geisert, H., Schmitz, R. W., Krainitzki, H., and Pääbo, S. 1999. DNA sequence of the mitochondrial hypervariable region II from the Neanderthal type specimen. *Proc. Natl. Acad. Sci. U.S.A.* 96:5581–5585.

Evolution in the Laboratory

- Sassanfar, M., and Szostak, J. W. 1993. An RNA motif that binds ATP. Nature 364:550–553.
- Gold, L., Polisky, B., Uhlenbeck, O., and Yarus, M. 1995. Diversity of oligonucleotide functions. Annu. Rev. Biochem. 64:763–797.
- Wilson, D. S., and Szostak, J. W. 1999. In vitro selection of functional nucleic acids. Annu. Rev. Biochem. 68:611–647.
- Hermann, T., and Patel, D. J. 2000. Adaptive recognition by nucleic acid aptamers. Science 287:820–825.

Web Sites

- The Protein Data Bank (PDB) site is the repository for three-dimensional macromolecular structures. It currently contains more than 30,000 structures. (http://www.rcsb.org/pdb/).
- National Center for Biotechnology Information (NCBI) contains molecular biological databases and software for analysis. (http:// www.ncbi.nlm.nih.gov/).

Chapter 7

Where to Start

- Perutz, M. F. 1978. Hemoglobin structure and respiratory transport. Sci. Am. 239(6):92–125.
- Perutz, M. F. 1980. Stereochemical mechanism of oxygen transport by haemoglobin. Proc. R. Soc. Lond. Biol. Sci. 208:135–162.

Kilmartin, J. V. 1976. Interaction of haemoglobin with protons, CO₂, and 2,3-diphosphoglycerate. *Brit. Med. Bull.* 32:209–222.

Structure

- Kendrew, J. C., Bodo, G., Dintzis, H. M., Parrish, R. G., Wyckoff, H., and Phillips, D. C. 1958. A three-dimensional model of the myoglobin molecule obtained by x-ray analysis. *Nature* 181:662–666.
- Shaanan, B. 1983. Structure of human oxyhaemoglobin at 2.1 Å resolution. J. Mol. Biol. 171:31–59.
- Frier, J. A., and Perutz, M. F. 1977. Structure of human foetal deoxyhaemoglobin. J. Mol. Biol. 112:97–112.
- Perutz, M. F. 1969. Structure and function of hemoglobin. *Harvey Lect.* 63:213–261.
- Perutz, M. F. 1962. Relation between structure and sequence of haemoglobin. *Nature* 194:914–917.

Interaction of Hemoglobin with Allosteric Effectors

- Benesch, R., and Beesch, R. E. 1969. Intracellular organic phosphates as regulators of oxygen release by haemoglobin. *Nature* 221: 618–622.
- Fang, T. Y., Zou, M., Simplaceanu, V., Ho, N. T., and Ho, C. 1999. Assessment of roles of surface histidyl residues in the molecular basis of the Bohr effect and of β 143 histidine in the binding of 2,3-bisphosphoglycerate in human normal adult hemoglobin. *Biochemistry* 38:13423–13432.
- Arnone, A. 1992. X-ray diffraction study of binding of 2,3-diphosphoglycerate to human deoxyhaemoglobin. *Nature* 237:146–149.

Models for Cooperativity

- Monod, J., Wyman, J., and Changeux, J.-P. 1965. On the nature of allosteric interactions: A plausible model. J. Mol. Biol. 12:88–118.
- Koshland, D. L., Jr., Nemethy, G., and Filmer, D. 1966. Comparison of experimental binding data and theoretical models in proteins containing subunits. *Biochemistry* 5:365–385.
- Ackers, G. K., Doyle, M. L., Myers, D., and Daugherty, M. A. 1992. Molecular code for cooperativity in hemoglobin. *Science* 255: 54–63.

Sickle-Cell Anemia and Thalasssemia

- Herrick, J. B. 1910. Peculiar elongated and sickle-shaped red blood corpuscles in a case of severe anemia. Arch. Intern. Med. 6: 517–521.
- Pauling, L., Itano, H. A., Singer, S. J., and Wells, L. C. 1949. Sickle cell anemia: A molecular disease. Science 110:543–548.
- Ingram, V. M. 1957. Gene mutation in human hemoglobin: The chemical difference between normal and sickle cell haemoglobin. *Nature* 180:326–328.
- Eaton, W. A., and Hofrichter, J. 1990. Sickle cell hemoglobin polymerization. Adv. Prot. Chem. 40:63–279.
- Weatherall, D. J. 2001. Phenotype genotype relationships in monogenic disease: Lessons from the thalassemias. Nat. Rev. Genet. 2:245–255.

Globin-Binding Proteins and Other Globins

- Kihm, A. J., Kong, Y., Hong, W., Russell, J. E., Rouda, S., Adachi, K., Simon, M. C., Blobel, G. A., and Weiss, M. J. 2002. An abundant erythroid protein that stabilizes free α-haemoglobin. *Nature* 417:758–763.
- Feng, L., Zhou, S., Gu, L., Gell, D. A., Mackay, J. P., Weiss, M. J., Gow, A. J., and Shi, Y. 2005. Structure of oxidized α-haemoglobin bound to AHSP reveals a protective mechanism for haem. *Nature* 435:697–701.
- Yu, X., Kong, Y., Dore, L. C., Abdulmalik, O., Katein, A. M., Zhou, S., Choi, J. K., Gell, D., Mackay, J. P., Gow, A. J., and Weiss, M. J. 2007. An erythroid chaperone that facilitates folding of α-globin subunits for hemoglobin synthesis. J. Clin. Invest. 117:1856–1865.
- Burmester, T., Haberkamp, M., Mitz, S., Roesner, A., Schmidt, M., Ebner, B., Gerlach, F., Fuchs, C., and Hankeln, T. 2004. Neuroglobin and cytoglobin: Genes, proteins and evolution. *IUBMB Life* 56:703–707.

- Hankeln, T., Ebner, B., Fuchs, C., Gerlach, F., Haberkamp, M., Laufs, T. L., Roesner, A., Schmidt, M., Weich, B., Wystub, S., Saaler-Reinhardt, S., Reuss, S., Bolognesi, M., De Sanctis, D., Marden, M. C., Kiger, L., Moens, L., Dewilde, S., Nevo, E., Avivi, A., Weber, R. E., Fago, A., and Burmester, T. 2005. Neuroglobin and cytoglobin in search of their role in the vertebrate globin family. J. Inorg. Biochem. 99:110–119.
- Burmester, T., Ebner, B., Weich, B., and Hankeln, T. 2002. Cytoglobin: A novel globin type ubiquitously expressed in vertebrate tissues. *Mol. Biol. Evol.* 19:416–421.
- Zhang, C., Wang, C., Deng, M., Li, L., Wang, H., Fan, M., Xu, W., Meng, F., Qian, L., and He, F. 2002. Full-length cDNA cloning of human neuroglobin and tissue expression of rat neuroglobin. *Biochem. Biophys. Res. Commun.* 290:1411–1419

Chapter 8

Where to Start

- Zalatan, J. G., and Herschlag, D. 2009. The far reaches of enzymology. Nat. Chem. Biol. 5:516–520.
- Hammes, G. G. 2008. How do enzymes really work? J. Biol. Chem. 283:22337-22346.
- Koshland, D. E., Jr. 1987. Evolution of catalytic function. Cold Spring Harbor Symp. Quant. Biol. 52:1–7.
- Jencks, W. P. 1987. Economics of enzyme catalysis. Cold Spring Harbor Symp. Quant. Biol. 52:65–73.
- Lerner, R. A., and Tramontano, A. 1988. Catalytic antibodies. Sci. Am. 258(3):58–70.

Books

- Cook, P. F., and Cleland, W. W. 2007. Enzyme Kinetics and Mechanism. Garland Press.
- Fersht, A. 1999. Structure and Mechanism in Protein Science: A Guide to Enzyme Catalysis and Protein Folding. W. H. Freeman and Company.
- Walsh, C. 1979. Enzymatic Reaction Mechanisms. W. H. Freeman and Company.
- Page, M. I., and Williams, A. (Eds.). 1987. Enzyme Mechanisms. Royal Society of Chemistry.
- Bender, M. L., Bergeron, R. J., and Komiyama, M. 1984. The Bioorganic Chemistry of Enzymatic Catalysis. Wiley-Interscience.
- Abelson, J. N., and Simon, M. I. (Eds.). 1992. Methods in Enzymology. Academic Press.
- Boyer, P. D. (Ed.). 1970. The Enzymes (3d ed.). Academic Press.
- Friedmann, H. C. (Ed.). 1981. Benchmark Papers in Biochemistry. Vol. 1, Enzymes. Hutchinson Ross.

Transition-State Stabilization, Analogs, and Other Enzyme Inhibitors

- Schramm, V. L. 2007. Enzymatic transition state theory and transition state analog design. J. Biol. Chem. 282:28297–28300.
- Pauling, L. 1948. Nature of forces between large molecules of biological interest. Nature 161:707–709.
- Leinhard, G. E. 1973. Enzymatic catalysis and transition-state theory. Science 180:149–154.
- Kraut, J. 1988. How do enzymes work? Science 242:533-540.
- Waxman, D. J., and Strominger, J. L. 1983. Penicillin-binding proteins and the mechanism of action of β-lactam antibiotics. Annu. Rev. Biochem. 52:825–869.
- Abraham, E. P. 1981. The B-lactam antibiotics. Sci. Am. 244:76-86.
- Walsh, C. T. 1984. Suicide substrates, mechanism-based enzyme inactivators: Recent developments. Annu. Rev. Biochem. 53:493–535.

Catalytic Antibodies

Hilvert, D. 2000. Critical analysis of antibody catalysis. Annu. Rev. Biochem. 69:751–794.

Selected Readings

- Wade, H., and Scanlan, T. S. 1997. The structural and functional basis of antibody catalysis. Annu. Rev. Biophys. Biomol. Struct. 26:461–493.
- Lerner, R. A., Benkovic, S. J., and Schultz, P. G. 1991. At the crossroads of chemistry and immunology: Catalytic antibodies. *Science* 252:659–667.
- Cochran, A. G., and Schultz, P. G. 1990. Antibody-catalyzed porphyrin metallation. Science 249:781–783.

Enzyme Kinetics and Mechanisms

- Hammes-Schiller, S. and Benkovic, S. J. 2006. Relating protein motion to catalysis. Annu. Rev. Biochem. 75:519–541.
- Benkovic, S. J., and Hammes-Schiller, S. 2003. A perspective on enzyme catalysis. Science 301:1196–1202.
- Hur, S., and Bruice, T. C. 2003. The near attack conformation approach to the study of the chorismate to prephenate reaction. *Proc. Natl. Acad. Sci. U.S.A.* 100:12015–12020.
- Xie, X. S., and Lu, H. P. 1999. Single-molecule enzymology. J. Biol. Chem. 274:15967–15970.
- Miles, E. W., Rhee, S., and Davies, D. R. 1999. The molecular basis of substrate channeling. J. Biol. Chem. 274:12193–12196.
- Warshel, A. 1998. Electrostatic origin of the catalytic power of enzymes and the role of preorganized active sites. J. Biol. Chem. 273:27035–27038.
- Cannon, W. R., and Benkovic, S. J. 1999. Solvation, reorganization energy, and biological catalysis. J. Biol. Chem. 273:26257–26260.
- Cleland, W. W., Frey, P. A., and Gerlt, J. A. 1998. The low barrier hydrogen bond in enzymatic catalysis. J. Biol. Chem. 273:25529–25532.
- Romesberg, F. E., Santarsiero, B. D., Spiller, B., Yin, J., Barnes, D., Schultz, P. G., and Stevens, R. C. 1998. Structural and kinetic evidence for strain in biological catalysis. *Biochemistry* 37:14404–14409.
- Lu, H. P., Xun, L., and Xie, X. S. 1998. Single-molecule enzymatic dynamics. Science 282:1877–1882.
- Fersht, A. R., Leatherbarrow, R. J., and Wells, T. N. C. 1986. Binding energy and catalysis: A lesson from protein engineering of the tyrosyl-tRNA synthetase. *Trends Biochem. Sci.* 11:321–325.
- Jencks, W. P. 1975. Binding energy, specificity, and enzymic catalysis: The Circe effect. *Adv. Enzymol.* 43:219–410.
- Knowles, J. R., and Albery, W. J. 1976. Evolution of enzyme function and the development of catalytic efficiency. *Biochemistry* 15:5631–5640.

Chapter 9

Where to Start

- Stroud, R. M. 1974. A family of protein-cutting proteins. Sci. Am. 231(1):74-88.
- Kraut, J. 1977. Serine proteases: Structure and mechanism of catalysis. Annu. Rev. Biochem. 46:331–358.
- Lindskog, S. 1997. Structure and mechanism of carbonic anhydrase. *Pharmacol. Ther.* 74:1–20.
- Jeltsch, A., Alves, J., Maass, G., and Pingoud, A. 1992. On the catalytic mechanism of *Eco*RI and *Eco*RV: A detailed proposal based on biochemical results, structural data and molecular modelling. *FEBS Lett.* 304:4–8.
- Bauer, C. B., Holden, H. M., Thoden, J. B., Smith, R., and Rayment, I. 2000. X-ray structures of the apo and MgATP-bound states of *Dictyostelium discoideum* myosin motor domain. J. Biol. Chem. 275:38494–38499.
- Lolis, E., and Petsko, G. A. 1990. Transition-state analogues in protein crystallography: Probes of the structural source of enzyme catalysis. *Annu. Rev. Biochem.* 59:597–630.

Books

- Fersht, A. 1999. Structure and Mechanism in Protein Science: A Guide to Enzyme Catalysis and Protein Folding. W. H. Freeman and Company.
- Silverman, R. B. 2000. The Organic Chemistry of Enzyme-Catalyzed Reactions. Academic Press.

Page, M., and Williams, A. 1997. Organic and Bio-organic Mechanisms. Addison Wesley Longman.

Chymotrypsin and Other Serine Proteases

- Fastrez, J., and Fersht, A. R. 1973. Demonstration of the acyl-enzyme mechanism for the hydrolysis of peptides and anilides by chymotrypsin. *Biochemistry* 12:2025–2034.
- Sigler, P. B., Blow, D. M., Matthews, B. W., and Henderson, R. 1968. Structure of crystalline-chymotrypsin II: A preliminary report including a hypothesis for the activation mechanism. J. Mol. Biol. 35:143–164.
- Kossiakoff, A. A., and Spencer, S. A. 1981. Direct determination of the protonation states of aspartic acid-102 and histidine-57 in the tetrahedral intermediate of the serine proteases: Neutron structure of trypsin. *Biochemistry* 20:6462–6474.
- Carter, P., and Wells, J. A. 1988. Dissecting the catalytic triad of a serine protease. *Nature* 332:564–568.
- Carter, P., and Wells, J. A. 1990. Functional interaction among catalytic residues in subtilisin BPN'. Proteins 7:335–342.
- Koepke, J., Ermler, U., Warkentin, E., Wenzl, G., and Flecker, P. 2000. Crystal structure of cancer chemopreventive Bowman-Birk inhibitor in ternary complex with bovine trypsin at 2.3 Å resolution: Structural basis of Janus-faced serine protease inhibitor specificity. J. Mol. Biol. 298:477–491.
- Gaboriaud, C., Rossi, V., Bally, I., Arlaud, G. J., and Fontecilla-Camps, J. C. 2000. Crystal structure of the catalytic domain of human complement C1s: A serine protease with a handle. *EMBO J.* 19:1755–1765.

Other Proteases

- Vega, S., Kang, L. W., Velazquez-Campoy, A., Kiso, Y., Amzel, L. M., and Freire, E. 2004. A structural and thermodynamic escape mechanism from a drug resistant mutation of the HIV-1 protease. *Proteins* 55:594–602.
- Kamphuis, I. G., Kalk, K. H., Swarte, M. B., and Drenth, J. 1984. Structure of papain refined at 1.65 Å resolution. J. Mol. Biol. 179:233–256.
- Kamphuis, I. G., Drenth, J., and Baker, E. N. 1985. Thiol proteases: Comparative studies based on the high-resolution structures of papain and actinidin, and on amino acid sequence information for cathepsins B and H, and stem bromelain. J. Mol. Biol. 182:317–329.
- Sivaraman, J., Nagler, D. K., Zhang, R., Menard, R., and Cygler, M. 2000. Crystal structure of human procathepsin X: A cysteine protease with the proregion covalently linked to the active site cysteine. J. Mol. Biol. 295:939–951.
- Davies, D. R. 1990. The structure and function of the aspartic proteinases. Annu. Rev. Biophys. Biophys. Chem. 19:189–215.
- Dorsey, B. D., Levin, R. B., McDaniel, S. L., Vacca, J. P., Guare, J. P., Darke, P. L., Zugay, J. A., Emini, E. A., Schleif, W. A., Quintero, J. C., et al. 1994. L-735,524: The design of a potent and orally bioavailable HIV protease inhibitor. J. Med. Chem. 37:3443–3451.
- Chen, Z., Li, Y., Chen, E., Hall, D. L., Darke, P. L., Culberson, C., Shafer, J. A., and Kuo, L. C. 1994. Crystal structure at 1.9-Å resolution of human immunodeficiency virus (HIV) II protease complexed with L-735,524, an orally bioavailable inhibitor of the HIV proteases. J. Biol. Chem. 269:26344–26348.
- Ollis, D. L., Cheah, E., Cygler, M., Dijkstra, B., Frolow, F., Franken, S. M., Harel, M., Remington, S. J., Silman, I., Schrag, J., et al. 1992. The α/β hydrolase fold. *Protein Eng.* 5:197–211.

Carbonic Anhydrase

- Strop, P., Smith, K. S., Iverson, T. M., Ferry, J. G., and Rees, D. C. 2001. Crystal structure of the "cab"-type beta class carbonic anhydrase from the archaeon *Methanobacterium thermoautotrophicum. J. Biol. Chem.* 276:10299–10305.
- Lindskog, S., and Coleman, J. E. 1973. The catalytic mechanism of carbonic anhydrase. Proc. Natl. Acad. Sci. U.S.A. 70:2505–2508.

- Kannan, K. K., Notstrand, B., Fridborg, K., Lovgren, S., Ohlsson, A., and Petef, M. 1975. Crystal structure of human erythrocyte carbonic anhydrase B: Three-dimensional structure at a nominal 2.2-Å resolution. Proc. Natl. Acad. Sci. U.S.A. 72:51–55.
- Boriack-Sjodin, P. A., Zeitlin, S., Chen, H. H., Crenshaw, L., Gross, S., Dantanarayana, A., Delgado, P., May, J. A., Dean, T., and Christianson, D. W. 1998. Structural analysis of inhibitor binding to human carbonic anhydrase II. *Protein Sci.* 7:2483–2489.
- Wooley, P. 1975. Models for metal ion function in carbonic anhydrase. Nature 258:677–682.
- Jonsson, B. H., Steiner, H., and Lindskog, S. 1976. Participation of buffer in the catalytic mechanism of carbonic anhydrase. *FEBS Lett.* 64:310–314.
- Sly, W. S., and Hu, P. Y. 1995. Human carbonic anhydrases and carbonic anhydrase deficiencies. Annu. Rev. Biochem. 64:375–401.
- Maren, T. H. 1988. The kinetics of HCO₃⁻ synthesis related to fluid secretion, pH control, and CO₂ elimination. Annu. Rev. Physiol. 50:695–717.
- Kisker, C., Schindelin, H., Alber, B. E., Ferry, J. G., and Rees, D. C. 1996. A left-hand beta-helix revealed by the crystal structure of a carbonic anhydrase from the archaeon *Methanosarcina thermophila*. *EMBO J.* 15:2323–2330.

Restriction Enzymes

- Selvaraj, S., Kono, H., and Sarai, A. 2002. Specificity of protein-DNA recognition revealed by structure-based potentials: Symmetric/ asymmetric and cognate/non-cognate binding. J. Mol. Biol. 322:907–915.
- Winkler, F. K., Banner, D. W., Oefner, C., Tsernoglou, D., Brown, R. S., Heathman, S. P., Bryan, R. K., Martin, P. D., Petratos, K., and Wilson, K. S. 1993. The crystal structure of *Eco*RV endonuclease and of its complexes with cognate and non-cognate DNA fragments. *EMBO J.* 12:1781–1795.
- Kostrewa, D., and Winkler, F. K. 1995. Mg²⁺ binding to the active site of *Eco*RV endonuclease: A crystallographic study of complexes with substrate and product DNA at 2 Å resolution. *Biochemistry* 34:683–696.
- Athanasiadis, A., Vlassi, M., Kotsifaki, D., Tucker, P. A., Wilson, K. S., and Kokkinidis, M. 1994. Crystal structure of *PvuII* endonuclease reveals extensive structural homologies to *EcoRV*. *Nat. Struct. Biol.* 1:469–475.
- Sam, M. D., and Perona, J. J. 1999. Catalytic roles of divalent metal ions in phosphoryl transfer by *Eco*RV endonuclease. *Biochemistry* 38:6576–6586.
- Jeltsch, A., and Pingoud, A. 1996. Horizontal gene transfer contributes to the wide distribution and evolution of type II restrictionmodification systems. J. Mol. Evol. 42:91–96.

Myosins

- Grigorenko, B. L., Rogov, A. V., Topol, I. A., Burt, S. K., Martinez, H. M., and Nemukhin, A. V. 2007. Mechanism of the myosin catalyzed hydrolysis of ATP as rationalized by molecular modeling. *Proc. Natl. Acad. Sci. U.S.A.* 104:7057–7061.
- Gulick, A. M., Bauer, C. B., Thoden, J. B., and Rayment, I. 1997. X-ray structures of the MgADP, MgATPγS, and MgAMPPNP complexes of the *Dictyostelium discoideum* myosin motor domain. *Biochemistry* 36:11619–11628.
- Kovacs, M., Malnasi-Csizmadia, A., Woolley, R. J., and Bagshaw, C. R. 2002. Analysis of nucleotide binding to *Dictyostelium* myosin II motor domains containing a single tryptophan near the active site. J. Biol. Chem. 277:28459–28467.
- Kuhlman, P. A., and Bagshaw, C. R. 1998. ATPase kinetics of the Dictyostelium discoideum myosin II motor domain. J. Muscle Res. Cell Motil. 19:491–504.
- Smith, C. A., and Rayment, I. 1996. X-ray structure of the magnesium(II) ADP vanadate complex of the *Dictyostelium discoideum* myosin motor domain to 1.9 Å resolution. *Biochemistry* 35: 5404–5417.

Chapter 10

Where to Start

- Kantrowitz, E. R., and Lipscomb, W. N. 1990. Escherichia coli aspartate transcarbamoylase: The molecular basis for a concerted allosteric transition. Trends Biochem. Sci. 15:53–59.
- Schachman, H. K. 1988. Can a simple model account for the allosteric transition of aspartate transcarbamoylase? J. Biol. Chem. 263: 18583–18586.
- Neurath, H. 1989. Proteolytic processing and physiological regulation. Trends Biochem. Sci. 14:268–271.
- Bode, W., and Huber, R. 1992. Natural protein proteinase inhibitors and their interaction with proteinases. *Eur. J. Biochem.* 204:433–451.

Aspartate Transcarbamoylase and Allosteric Interactions

- Rabinowitz, J. D., Hsiao, J. J., Gryncel, K. R., Kantrowitz, E. R., Feng, X.-J., Li, G., and Rabitz H. 2008. Dissecting enzyme regulation by multiple allosteric effectors: Nucleotide regulation of aspartate transcarbamoylase. *Biochemistry* 47:5881–5888.
- West, J. M., Tsuruta, H., and Kantrowitz, E. R. 2004. A fluorescent probelabeled *Escherichia coli* aspartate transcarbamoylase that monitors the allosteric conformation state. *J. Biol. Chem.* 279:945–951.
- Endrizzi, J. A., Beernink, P. T., Alber, T., and Schachman, H. K. 2000. Binding of bisubstrate analog promotes large structural changes in the unregulated catalytic trimer of aspartate transcarbamoylase: Implications for allosteric regulation. *Proc. Natl. Acad. Sci. U. S. A.* 97:5077–5082.
- Beernink, P. T., Endrizzi, J. A., Alber, T., and Schachman, H. K. 1999. Assessment of the allosteric mechanism of aspartate transcarbamoylase based on the crystalline structure of the unregulated catalytic subunit. *Proc. Natl. Acad. Sci. U.S.A.* 96:5388–5393.
- Wales, M. E., Madison, L. L., Glaser, S. S., and Wild, J. R. 1999. Divergent allosteric patterns verify the regulatory paradigm for aspartate transcarbamoylase. J. Mol. Biol. 294:1387–1400.
- Eisenstein, E., Markby, D. W., and Schachman, H. K. 1990. Heterotropic effectors promote a global conformational change in aspartate transcarbamoylase. *Biochemistry* 29:3724–3731.
- Newell, J. O., Markby, D. W., and Schachman, H. K. 1989. Cooperative binding of the bisubstrate analog N-(phosphonacetyl)-L-aspartate to aspartate transcarbamoylase and the heterotropic effects of ATP and CTP. J. Biol. Chem. 264:2476–2481.
- Stevens, R. C., Reinisch, K. M., and Lipscomb, W. N. 1991. Molecular structure of *Bacillus subtilis* aspartate transcarbamoylase at 3.0 Å resolution. *Proc. Natl. Acad. Sci. U.S.A.* 88:6087–6091.
- Stevens, R. C., Gouaux, J. E., and Lipscomb, W. N. 1990. Structural consequences of effector binding to the T state of aspartate carbamoyltransferase: Crystal structures of the unligated and ATP- and CTPcomplexed enzymes at 2.6-Å resolution. *Biochemistry* 29:7691–7701.
- Gouaux, J. E., and Lipscomb, W. N. 1990. Crystal structures of phosphonoacetamide ligated T and phosphonoacetamide and malonate ligated R states of aspartate carbamoyltransferase at 2.8-Å resolution and neutral pH. *Biochemistry* 29:389–402.
- Labedan, B., Boyen, A., Baetens, M., Charlier, D., Chen, P., Cunin, R., Durbeco, V., Glansdorff, N., Herve, G., Legrain, C., Liang, Z., Purcarea, C., Roovers, M., Sanchez, R., Toong, T. L., Van de Casteele, M., van Vliet, F., Xu, Y., and Zhang, Y. F. 1999. The evolutionary history of carbamoyltransferases: A complex set of paralogous genes was already present in the last universal common ancestor. J. Mol. Evol. 49:461–473.

Covalent Modification

- Tarrant, M. K., and Cole, P.A. 2009. The chemical biology of protein phosphorylation Annu. Rev. Biochem. 78:797–825.
- Johnson, L. N., and Barford, D. 1993. The effects of phosphorylation on the structure and function of proteins. Annu. Rev. Biophys. Biomol. Struct. 22:199–232.

- Ziegler, M. 2000. New functions of a long-known molecule: Emerging roles of NAD in cellular signaling. Eur. J. Biochem. 267:1550-1564.
- Ng, H. H., and Bird, A. 2000. Histone deacetylases: Silencers for hire. Trends Biochem. Sci. 25:121–126.
- Jacobson, M. K., and Jacobson, E. L. 1999. Discovering new ADPribose polymer cycles: Protecting the genome and more. *Trends Biochem. Sci.* 24:415–417.
- Barford, D., Das, A. K., and Egloff, M. P. 1998. The structure and mechanism of protein phosphatases: Insights into catalysis and regulation. Annu. Rev. Biophys. Biomol. Struct. 27:133–164.

Protein Kinase A

- Taylor, S. S., Knighton, D. R., Zheng, J., Sowadski, J. M., Gibbs, C. S., and Zoller, M. J. 1993. A template for the protein kinase family. *Trends Biochem. Sci.* 18:84–89.
- Gibbs, C. S., Knighton, D. R., Sowadski, J. M., Taylor, S. S., and Zoller, M. J. 1992. Systematic mutational analysis of cAMP-dependent protein kinase identifies unregulated catalytic subunits and defines regions important for the recognition of the regulatory subunit. J. Biol. Chem. 267:4806–4814.
- Knighton, D. R., Zheng, J. H., TenEyck, L., Ashford, V. A., Xuong, N. H., Taylor, S. S., and Sowadski, J. M. 1991. Crystal structure of the catalytic subunit of cyclic adenosine monophosphate-dependent protein kinase. *Science* 253:407–414.
- Knighton, D. R., Zheng, J. H., TenEyck, L., Xuong, N. H., Taylor, S. S., and Sowadski, J. M. 1991. Structure of a peptide inhibitor bound to the catalytic subunit of cyclic adenosine monophosphatedependent protein kinase. *Science* 253:414–420.
- Adams, S. R., Harootunian, A. T., Buechler, Y. J., Taylor, S. S., and Tsien, R. Y. 1991. Fluorescence ratio imaging of cyclic AMP in single cells. *Nature* 349:694–697.

Zymogen Activation

- Neurath, H. 1986. The versatility of proteolytic enzymes. J. Cell. Biochem. 32:35–49.
- Bode, W., and Huber, R. 1986. Crystal structure of pancreatic serine endopeptidases. In *Molecular and Cellular Basis of Digestion* (pp. 213–234), edited by P. Desnuelle, H. Sjostrom, and O. Noren. Elsevier.
- James, M. N. 1991. Refined structure of porcine pepsinogen at 1.8 Å resolution. J. Mol. Biol. 219:671–692.

Protease Inhibitors

- Carrell, R., and Travis, J. 1985. α₁-Antitrypsin and the serpins: Variation and countervariation. *Trends Biochem. Sci.* 10:20–24.
- Carp, H., Miller, F., Hoidal, J. R., and Janoff, A. 1982. Potential mechanism of emphysema: α₁-Proteinase inhibitor recovered from lungs of cigarette smokers contains oxidized methionine and has decreased elastase inhibitory capacity. *Proc. Natl. Acad. Sci. U.S.A.* 79:2041–2045.
- Owen, M. C., Brennan, S. O., Lewis, J. H., and Carrell, R. W. 1983. Mutation of antitrypsin to antithrombin. New Engl. J. Med. 309:694–698.
- Travis, J., and Salvesen, G. S. 1983. Human plasma proteinase inhibitors. Annu. Rev. Biochem. 52:655–709.

Clotting Cascade

- Furie, B., and Furie, B. C. 2008. Mechanisms of thrombus formation. New Engl. J. Med. 359:938–949.
- Orfeo, T., Brufatto, N., Nesheim, M. E., Xu, H., Butenas, S., and Mann, K. G. 2004. The factor V activation paradox. *J. Biol. Chem.* 279:19580–19591.
- Mann, K. G. 2003. Thrombin formation. Chest 124:4S-10S.
- Rose, T., and Di Cera, E. 2002. Three-dimensional modeling of thrombin–fibrinogen interaction. J. Biol. Chem. 277:18875–18880.
- Krem, M. M., and Di Cera, E. 2002. Evolution of cascades from embryonic development to blood coagulation. *Trends Biochem.* Sci. 27: 67–74.

- Fuentes-Prior, P., Iwanaga, Y., Huber, R., Pagila, R., Rumennik, G., Seto, M., Morser, J., Light, D. R., and Bode, W. 2000. Structural basis for the anticoagulant activity of the thrombin– thrombomodulin complex. *Nature* 404:518–525.
- Lawn, R. M., and Vehar, G. A. 1986. The molecular genetics of hemophilia. Sci. Am. 254(3):48–65.
- Brown, J. H., Volkmann, N., Jun, G., Henschen-Edman, A. H., and Cohen, C. 2000. The crystal structure of modified bovine fibrinogen. Proc. Natl. Acad. Sci. U.S.A. 97:85–90.
- Stubbs, M. T., Oschkinat, H., Mayr, I., Huber, R., Angliker, H., Stone, S. R., and Bode, W. 1992. The interaction of thrombin with fibrinogen: A structural basis for its specificity. *Eur. J. Biochem.* 206:187–195.

Chapter 11

Where to Start

- Glycochemistry and glycobiology. A series of review articles. 2007. Nature 446:999-1051.
- Maeder, T. 2002. Sweet medicines. Sci. Am. 287(1):40-47.
- Sharon, N., and Lis, H. 1993. Carbohydrates in cell recognition. Sci. Am. 268(1):82–89.
- Lasky, L. A. 1992. Selectins: Interpreters of cell-specific carbohydrate information during inflammation. *Science* 258:964–969.
- Woods, R. J. 1995. Three-dimensional structures of oligosaccharides. Curr. Opin. Struct. Biol. 5:591–598.

Books

- Varki, A., Cummings, R., Esko, J., Freeze, H., Stanley, P., Bertozzi, C., Hart, G., and Etzler, M. E. 2009. *Essentials of Glycobiology*, 2d ed. Cold Spring Harbor Laboratory Press.
- Stick, R. V., and Williams, S. 2008. Carbohydrates: The Essential Molecules of Life, 2d ed. Elsevier Science.
- Sansome, C., and Markman, O. 2007. Glycobiology. Scion.
- Lindhorst, T. K. 2007. Essentials of Carbohydrate Chemistry and Biochemistry, 3d ed. Wiley-VCH.
- Taylor, M. E. 2006. Introduction to Glycobiology, 2d ed. Oxford University Press.

Carbohydrate-Binding Proteins and Glycoproteins

- Lairson, L. L., Henrissat, B., Davies, G. J., and Withers, S. G. 2008. Glycosyltransferases: Structures, functions and mechanisms. *Annu. Rev. Biochem.* 77:521–555.
- Foley, R. N. 2008. Erythropoietin: Physiology and molecular mechanisms. *Heart Failure Rev.* 13:404–414.
- Yan, A., and Lennarz, W. J. 2005. Unraveling the mechanism of protein N-glycosylation. J. Biol. Chem. 280:3121–3124.
- Qasba, P. K., Ramakrishnan, B., and Boeggeman, E. 2005. Substrateinduced conformational changes in glycosyltransferases. *Trends Biochem. Sci.* 30:53–62.
- Pratta, M. A., Yao, W., Decicco, C., Tortorella, M., Liu, R.-Q., Copeland, R. A., Magolda, R., Newton, R. C., Trzaskos, J. M., and Arner, E. C. 2003. Aggrecan protects cartilage collagen from proteolytic cleavage. J. Biol. Chem. 278:45539–45545.
- Fisher, J. W. 2003. Erythropoietin: Physiology and pharmacology update. *Exp. Biol. Med.* 228:1–14.
- Cheetham, J. C., Smith, D. M., Aoki, K. H., Stevenson, J. L., Hoeffel, T. J., Syed, R. S., Egrie, J., and Harvey, T. S. 1998. NMR structure of human erythropoietin and a comparison with its receptor bound conformation. *Nat. Struct. Biol.* 5:861–866.
- Bouckaert, J., Hamelryck, T., Wyns, L., and Loris, R. 1999. Novel structures of plant lectins and their complexes with carbohydrates. *Curr. Opin. Struct. Biol.* 9:572–577.
- Weis, W. I., and Drickamer, K. 1996. Structural basis of lectincarbohydrate recognition. Annu. Rev. Biochem. 65:441–473.

- Vyas, N. K. 1991. Atomic features of protein-carbohydrate interactions. Curr. Opin. Struct. Biol. 1:732-740.
- Weis, W. I., Drickamer, K., and Hendrickson, W. A. 1992. Structure of a C-type mannose-binding protein complexed with an oligosaccharide. *Nature* 360:127–134.
- Shaanan, B., Lis, H., and Sharon, N. 1991. Structure of a legume lectin with an ordered N-linked carbohydrate in complex with lactose. *Science* 254:862–866.

Glycoproteins

- Hattrup, C. L., and Gendler, S. J. 2008. Structure and function of the cell surface (tethered) mucins. Annu. Rev. Physiol. 70:431–457.
- Thorton, D. J., Rousseau, K., and McGuckin, M. A. 2008. Structure and function of mucins in airways mucus. Annu. Rev. Physiol. 70:459–486.
- Rose, M. C., and Voynow, J. A. 2007. Respiratory tract mucin genes and mucin glycoproteins in health and disease. *Physiol. Rev.* 86:245–278.
- Lamoureux, F., Baud'huin, M., Duplomb, L., Heymann, D., and Rédini, F. 2007. Proteoglycans: Key partners in bone cell biology. *Bioessays* 29:758–771.
- Carraway, K. L., Funes, M., Workman, H. C., and Sweeney, C. 2007. Contribution of membrane mucins to tumor progression through modulation of cellular growth signaling pathways. *Curr. Top. Dev. Biol.* 78:1–22.
- Bernfield, M., Götte, M., Park, P. W., Reizes, O., Fitzgerald, M. L., Lincecum, J., and Zako, M. 1999. Functions of cell surface heparan sulfate proteoglycans. *Annu. Rev. Biochem.* 68:729–777.
- Iozzo, R. V. 1998. Matrix proteoglycans: From molecular design to cellular function. Annu. Rev. Biochem. 67:609–652.
- Yanagishita, M., and Hascall, V. C. 1992. Cell surface heparan sulfate proteoglycans. J. Biol. Chem. 267:9451–9454.
- Iozzo, R. V. 1999. The biology of small leucine-rich proteoglycans: Functional network of interactive proteins. J. Biol. Chem. 274: 18843–18846.

Carbohydrates in Recognition Processes

- Wasserman, P. M. 2008. Zona pellucida glycoproteins. J. Biol. Chem. 283:24285–24289.
- Sharon, N. 2008. Lectins: Past, present and future. Biochem. Soc. Trans. 36:1457–1460.
- Balzarini, J, 2007. Targeting the glycans of glycoproteins: A novel paradigm for antiviral therapy. Nat. Rev. Microbiol. 5:583–597.
- Sharon, N. 2007. Lectins: Carbohydrate-specific reagents and biological recognition molecules. J. Biol. Chem. 282:2753–2764.
- Stevens, J., Blixt, O., Tumpey, T. M., Taubenberger, J. K., Paulson, J. C., and Wilson, I. A. 2006. Structure and receptor specificity of hemagglutinin from an H5N1 influenza virus. *Science* 312: 404–409.
- Cambi, A., Koopman, M., and Figdor, C. G. 2005. How C-type lectins detect pathogens. *Cell. Microbiol.* 7:481–488.
- Turner, M. L. 1992. Cell adhesion molecules: A unifying approach to topographic biology. Biol. Rev. Camb. Philos. Soc. 67:359–377.
- Feizi, T. 1992. Blood group-related oligosaccharides are ligands in celladhesion events. *Biochem. Soc. Trans.* 20:274–278.
- Jessell, T. M., Hynes, M. A., and Dodd, J. 1990. Carbohydrates and carbohydrate-binding proteins in the nervous system. Annu. Rev. Neurosci. 13:227–255.
- Clothia, C., and Jones, E. V. 1997. The molecular structure of cell adhesion molecules. Annu. Rev. Biochem. 66:823–862.

Carbohydrate Sequencing

- Venkataraman, G., Shriver, Z., Raman, R., and Sasisekharan, R. 1999. Sequencing complex polysaccharides. *Science* 286:537–542.
- Zhao, Y., Kent, S. B. H., and Chait, B. T. 1997. Rapid, sensitive structure analysis of oligosaccharides. Proc. Natl. Acad. Sci. U.S.A. 94:1629–1633.
- Rudd, P. M., Guile, G. R., Küster, B., Harvey, D. J., Opdenakker, G., and Dwek, R. A. 1997. Oligosaccharide sequencing technology. *Nature* 388:205–207.

Chapter 12

Where to Start

- De Weer, P. 2000. A century of thinking about cell membranes. Annu. Rev. Physiol. 62:919–926.
- Bretscher, M. S. 1985. The molecules of the cell membrane. Sci. Am. 253(4):100–108.
- Unwin, N., and Henderson, R. 1984. The structure of proteins in biological membranes. *Sci. Am.* 250(2):78–94.
- Deisenhofer, J., and Michel, H. 1989. The photosynthetic reaction centre from the purple bacterium *Rhodopseudomonas viridis*. *EMBO J.* 8:2149–2170.
- Singer, S. J., and Nicolson, G. L. 1972. The fluid mosaic model of the structure of cell membranes. Science 175:720–731.
- Jacobson, K., Sheets, E. D., and Simson, R., 1995. Revisiting the fluid mosaic model of membranes. Science 268:1441–1442.

Books

- Gennis, R. B. 1989. Biomembranes: Molecular Structure and Function. Springer Verlag.
- Vance, D. E., and Vance, J. E. (Eds.). 1996. Biochemistry of Lipids, Lipoproteins, and Membranes. Elsevier.
- Lipowsky, R., and Sackmann, E. 1995. The Structure and Dynamics of Membranes. Elsevier.
- Racker, E. 1985. Reconstitutions of Transporters, Receptors, and Pathological States. Academic Press.
- Tanford, C. 1980. The Hydrophobic Effect: Formation of Micelles and Biological Membranes (2d ed.). Wiley-Interscience.

Membrane Lipids and Dynamics

- Lingwood, D., and Simons, K. 2010. Lipid rafts as a membraneorganizing principle. Science. 327:46-50.
- Pike, L. J. 2009. The challenge of lipid rafts. J. Lipid Res. 50:S323-S328.
- Simons, K., and Vaz, W. L. 2004. Model systems, lipid rafts, and cell membranes. Annu. Rev. Biophys. Biomol. Struct. 33:269–295.
- Anderson, T. G., and McConnell, H. M. 2002. A thermodynamic model for extended complexes of cholesterol and phospholipid. *Biophys. J.* 83:2039–2052.
- Saxton, M. J., and Jacobson, K. 1997. Single-particle tracking: Applications to membrane dynamics. Annu. Rev. Biophys. Biomol. Struct. 26:373–399.
- Bloom, M., Evans, E., and Mouritsen, O. G. 1991. Physical properties of the fluid lipid-bilayer component of cell membranes: A perspective. Q. Rev. Biophys. 24:293–397.
- Elson, E. L. 1986. Membrane dynamics studied by fluorescence correlation spectroscopy and photobleaching recovery. Soc. Gen. Physiol. Ser. 40:367–383.
- Zachowski, A., and Devaux, P. F. 1990. Transmembrane movements of lipids. *Experientia* 46:644–656.
- Devaux, P. F. 1992. Protein involvement in transmembrane lipid asymmetry. Annu. Rev. Biophys. Biomol. Struct. 21:417–439.
- Silvius, J. R. 1992. Solubilization and functional reconstitution of biomembrane components. Annu. Rev. Biophys. Biomol. Struct. 21:323–348.
- Yeagle, P. L., Albert, A. D., Boesze-Battaglia, K., Young, J., and Frye, J. 1990. Cholesterol dynamics in membranes. *Biophys. J.* 57:413-424.
- Nagle, J. F., and Tristram-Nagle, S. 2000. Lipid bilayer structure. Curr. Opin. Struct. Biol. 10:474–480.
- Dowhan, W. 1997. Molecular basis for membrane phospholipid diversity: Why are there so many lipids? Annu. Rev. Biochem. 66:199–232.
- Huijbregts, R. P. H., de Kroon, A. I. P. M., and de Kruijff, B. 1998. Rapid transmembrane movement of newly synthesized phosphatidylethanolamine across the inner membrane of *Escherichia coli*. J. Biol.Chem. 273:18936–18942.

Structure of Membrane Proteins

- Walian, P., Cross, T. A., and Jap, B. K. 2004. Structural genomics of membrane proteins. *Genome Biol.* 5:215.
- Werten, P. J., Remigy, H. W., de Groot, B. L., Fotiadis, D., Philippsen, A., Stahlberg, H., Grubmuller, H., and Engel, A. 2002. Progress in the analysis of membrane protein structure and function. *FEBS Lett.* 529:65–72.
- Popot, J.-L., and Engleman, D. M. 2000. Helical membrane protein folding, stability and evolution. Annu. Rev. Biochem. 69:881–922.
- White, S. H., and Wimley, W. C. 1999. Membrane protein folding and stability: Physical principles. Annu. Rev. Biophys. Biomol. Struct. 28:319–365.
- Marassi, F. M., and Opella, S. J. 1998. NMR structural studies of membrane proteins. Curr. Opin. Struct. Biol. 8:640–648.
- Lipowsky, R. 1991. The conformation of membranes. *Nature* 349:475–481.
- Altenbach, C., Marti, T., Khorana, H. G., and Hubbell, W. L. 1990. Transmembrane protein structure: Spin labeling of bacteriorhodopsin mutants. *Science* 248:1088–1092.
- Fasman, G. D., and Gilbert, W. A. 1990. The prediction of transmembrane protein sequences and their conformation: An evaluation. *Trends Biochem. Sci.* 15:89–92.
- Jennings, M. L. 1989. Topography of membrane proteins. Annu. Rev. Biochem. 58:999–1027.
- Engelman, D. M., Steitz, T. A., and Goldman, A. 1986. Identifying non-polar transbilayer helices in amino acid sequences of membrane proteins. *Annu. Rev. Biophys. Biophys. Chem.* 15:321–353.
- Udenfriend, S., and Kodukola, K. 1995. How glycosyl-phosphatidylinositol-anchored membrane proteins are made. Annu. Rev. Biochem. 64:563–591.

Intracellular Membranes

- Skehel, J. J., and Wiley, D. C. 2000. Receptor binding and membrane fusion in virus entry: The influenza hemagglutinin. Annu. Rev. Biochem. 69:531–569.
- Roth, M. G. 1999. Lipid regulators of membrane traffic through the Golgi complex. *Trends Cell Biol*. 9:174–179.
- Jahn, R., and Sudhof, T. C. 1999. Membrane fusion and exocytosis. Annu. Rev. Biochem. 68:863-911.
- Stroud, R. M., and Walter, P. 1999. Signal sequence recognition and protein targeting. Curr. Opin. Struct. Biol. 9:754–759.
- Teter, S. A., and Klionsky, D. J. 1999. How to get a folded protein across a membrane. *Trends Cell Biol.* 9:428–431.
- Hettema, E. H., Distel, B., and Tabak, H. F. 1999. Import of proteins into peroxisomes. *Biochim. Biophys. Acta* 1451:17–34.

Membrane Fusion

- Sollner, T. H., and Rothman, J. E. 1996. Molecular machinery mediating vesicle budding, docking and fusion. *Experientia* 52:1021–1025.
- Ungar, D., and Hughson, F. M. 2003. SNARE protein structure and function. Annu. Rev. Cell Dev. Biol. 19:493-517.

Chapter 13

Where to Start

- Lancaster, C. R. 2004. Structural biology: Ion pump in the movies. Nature 432:286–287.
- Unwin, N. 2003. Structure and action of the nicotinic acetylcholine receptor explored by electron microscopy. FEBS Lett. 555:91–95.
- Abramson, J., Smirnova, I., Kasho, V., Verner, G., Iwata, S., and Kaback, H. R. 2003. The lactose permease of *Escherichia coli*: Overall structure, the sugar-binding site and the alternating access model for transport. *FEBS Lett.* 555:96–101.
- Lienhard, G. E., Slot, J. W., James, D. E., and Mueckler, M. M. 1992. How cells absorb glucose. Sci. Am. 266(1):86–91.

- King, L. S., Kozono, D., and Agre, P. 2004. From structure to disease: The evolving tale of aquaporin biology. Nat. Rev. Mol. Cell Biol. 5:687–698.
- Neher, E., and Sakmann, B. 1992. The patch clamp technique. Sci. Am. 266(3):28–35.
- Sakmann, B. 1992. Elementary steps in synaptic transmission revealed by currents through single ion channels. *Science* 256:503–512.

Books

- Ashcroft, F. M. 2000. Ion Channels and Disease. Academic Press.
- Conn, P. M. (Ed.). 1998. Ion Channels, vol. 293, Methods in Enzymology. Academic Press.
- Aidley, D. J., and Stanfield, P. R. 1996. Ion Channels: Molecules in Action. Cambridge University Press.
- Hille, B. 2001. Ionic Channels of Excitable Membranes (3d ed.). Sinauer.
- Läuger, P. 1991. Electrogenic Ion Pumps. Sinauer.
- Stein, W. D. 1990. Channels, Carriers, and Pumps: An Introduction to Membrane Transport. Academic Press.
- Hodgkin, A. 1992. Chance and Design: Reminiscences of Science in Peace and War. Cambridge University Press.

P-Type ATPases

- Sorensen, T. L., Moller, J. V., and Nissen, P. 2004. Phosphoryl transfer and calcium ion occlusion in the calcium pump. *Science* 304:1672–1675.
- Sweadner, K. J., and Donnet, C. 2001. Structural similarities of Na,K-ATPase and SERCA, the Ca²⁺-ATPase of the sarcoplasmic reticulum. *Biochem. J.* 356:685–704.
- Toyoshima, C., and Mizutani, T. 2004. Crystal structure of the calcium pump with a bound ATP analogue. *Nature* 430:529–535.
- Toyoshima, C., Nakasako, M., Nomura, H., and Ogawa, H. 2000. Crystal structure of the calcium pump of sarcoplasmic reticulum at 2.6 Å resolution. *Nature* 405:647–655.
- Auer, M., Scarborough, G. A., and Kuhlbrandt, W. 1998. Threedimensional map of the plasma membrane H⁺-ATPase in the open conformation. Nature 392:840–843.
- Axelsen, K. B., and Palmgren, M. G. 1998. Evolution of substrate specificities in the P-type ATPase superfamily. J. Mol. Evol. 46:84–101.
- Pedersen, P. A., Jorgensen, J. R., and Jorgensen, P. L. 2000. Importance of conserved a-subunit segment ⁷⁰⁹GDGVND for Mg²⁺ binding, phosphorylation, energy transduction in Na,K-ATPase. J. Biol. Chem. 275:37588–37595.
- Blanco, G., and Mercer, R. W. 1998. Isozymes of the Na-K-ATPase: heterogeneity in structure, diversity in function. Am. J. Physiol. 275:F633-F650.
- Estes, J. W., and White, P. D. 1965. William Withering and the purple foxglove. Sci. Am. 212(6):110–117.

ATP-Binding Cassette Proteins

- Ward, A., Reyes, C. L., Yu, J., Roth, C. B., and Chang, G. 2007. Flexibility in the ABC transporter MsbA: Alternating access with a twist. *Proc. Natl. Acad. Sci. U.S.A.* 104:19005–19010.
- Locher, K. P. 2004. Structure and mechanism of ABC transporters. Curr. Opin. Struct. Biol. 14:426–431.
- Locher, K. P., Lee, A. T., and Rees, D. C. 2002. The *E. coli* BtuCD structure: A framework for ABC transporter architecture and mechanism. *Science* 296:1091–1098.
- Borths, E. L., Locher, K. P., Lee, A. T., and Rees, D. C. 2002. The structure of *Escherichia coli* BtuF and binding to its cognate ATP binding cassette transporter. *Proc. Natl. Acad. Sci. U.S.A.* 99:16642–16647.
- Dong, J., Yang, G., and McHaourab, H. S. 2005. Structural basis of energy transduction in the transport cycle of MsbA. *Science* 308:1023–1028.
- Akabas, M. H. 2000. Cystic fibrosis transmembrane conductance regulator: Structure and function of an epithelial chloride channel. J. Biol. Chem. 275:3729–3732.
- Chen, J., Sharma, S., Quiocho, F. A., and Davidson, A. L. 2001. Trapping the transition state of an ATP-binding cassette transporter: Evidence

SELECTED READINGS

for a concerted mechanism of maltose transport. Proc. Natl. Acad. Sci. U.S.A. 98:1525–1530.

- Sheppard, D. N., and Welsh, M. J. 1999. Structure and function of the CFTR chloride channel. *Physiol. Rev.* 79:S23–S45.
- Jones, P. M., and George, A. M. 2000. Symmetry and structure in P-glycoprotein and ABC transporters: What goes around comes around. *Eur. J. Biochem.* 287:5298–5305.
- Chen, Y., and Simon, S. M. 2000. In situ biochemical demonstration that P-glycoprotein is a drug efflux pump with broad specificity. J. Cell Biol. 148:863–870.
- Saier, M. H., Jr., Paulsen, I. T., Sliwinski, M. K., Pao, S. S., Skurray, R. A., and Nikaido, H. 1998. Evolutionary origins of multidrug and drugspecific efflux pumps in bacteria. *FASEB J*. 12:265–274.

Symporters and Antiporters

- Abramson, J., Smirnova, I., Kasho, V., Verner, G., Kaback, H. R., and Iwata, S. 2003. Structure and mechanism of the lactose permease of *Escherichia coli. Science* 301:610–615.
- Philipson, K. D., and Nicoll, D. A. 2000. Sodium-calcium exchange: A molecular perspective. Annu. Rev. Physiol. 62:111–133.
- Pao, S. S., Paulsen, I. T., and Saier, M. H., Jr. 1998. Major facilitator superfamily. *Microbiol. Mol. Biol. Rev.* 62:1–34.
- Wright, E. M., Hirsch, J. R., Loo, D. D., and Zampighi, G. A. 1997. Regulation of Na⁺/glucose cotransporters. J. Exp. Biol. 200:287–293.
- Kaback, H. R., Bibi, E., and Roepe, P. D. 1990. β-Galactoside transport in *E. coli*: A functional dissection of lac permease. *Trends Biochem. Sci.* 8:309–314.
- Hilgemann, D. W., Nicoll, D. A., and Philipson, K. D. 1991. Charge movement during Na⁺ translocation by native and cloned cardiac Na⁺/Ca²⁺ exchanger. *Nature* 352:715–718.
- Hediger, M. A., Turk, E., and Wright, E. M. 1989. Homology of the human intestinal Na⁺/glucose and *Escherichia coli* Na⁺/proline cotransporters. *Proc. Natl. Acad. Sci. U.S.A.* 86:5748–5752.

Ion Channels

- Zhou, Y., and MacKinnon, R. 2003. The occupancy of ions in the K⁺ selectivity filter: Charge balance and coupling of ion binding to a protein conformational change underlie high conduction rates. J. Mol. Biol. 333:965–975.
- Zhou, Y., Morais-Cabral, J. H., Kaufman, A., and MacKinnon, R. 2001. Chemistry of ion coordination and hydration revealed by a K⁺ channel-Fab complex at 2.0 Å resolution. *Nature* 414:43–48.
- Jiang, Y., Lee, A., Chen, J., Cadene, M., Chait, B. T., and MacKinnon, R. 2002. The open pore conformation of potassium channels. *Nature* 417:523–526.
- Jiang, Y., Lee, A., Chen, J., Ruta, V., Cadene, M., Chait, B. T., and MacKinnon, R. 2003. X-ray structure of a voltage-dependent K⁺ channel. *Nature* 423:33–41.
- Jiang, Y., Ruta, V., Chen, J., Lee, A., and MacKinnon, R. 2003. The principle of gating charge movement in a voltage-dependent K⁺ channel. *Nature* 423:42–48.
- Mackinnon, R. 2004. Structural biology: Voltage sensor meets lipid membrane. Science 306:1304–1305.
- Bezanilla, F. 2000. The voltage sensor in voltage-dependent ion channels. *Physiol. Rev.* 80:555–592.
- Shieh, C.-C., Coghlan, M., Sullivan, J. P., and Gopalakrishnan, M. 2000. Potassium channels: Molecular defects, diseases, and therapeutic opportunities. *Pharmacol. Rev.* 52:557–594.
- Horn, R. 2000. Conversation between voltage sensors and gates of ion channels. *Biochemistry* 39:15653–15658.
- Perozo, E., Cortes, D. M., and Cuello, L. G. 1999. Structural rearrangements underlying K⁺-channel activation gating. Science 285:73–78.
- Doyle, D. A., Morais Cabral, J., Pfuetzner, R. A., Kuo, A., Gulbis, J. M., Cohen, S. L., Chait, B. T., and MacKinnon R. 1998. The structure of the potassium channel: Molecular basis of K⁺ conduction and selectivity. *Science* 280:69–77.

- Marban, E., Yamagishi, T., and Tomaselli, G. F. 1998. Structure and function of the voltage-gated Na⁺ channel. J. Physiol. 508:647–657.
- Miller, R. J. 1992. Voltage-sensitive Ca²⁺ channels. J. Biol. Chem. 267:1403–1406.
- Catterall, W. A. 1991. Excitation-contraction coupling in vertebrate skeletal muscle: A tale of two calcium channels. *Cell* 64:871–874.

Ligand-Gated Ion Channels

- Unwin, N. 2005. Refined structure of the nicotinic acetylcholine receptor at 4 Å resolution. J. Mol. Biol. 346:967–989.
- Miyazawa, A., Fujiyoshi, Y., Stowell, M., and Unwin, N. 1999. Nicotinic acetylcholine receptor at 4.6 Å resolution: Transverse tunnels in the channel wall. J. Mol. Biol. 288:765–786.
- Jiang, Y., Lee, A., Chen, J., Cadene, M., Chait, B. T., and MacKinnon, R. 2002. Crystal structure and mechanism of a calcium-gated potassium channel. *Nature* 417:515–522.
- Barrantes, F. J., Antollini, S. S., Blanton, M. P., and Prieto, M. 2000. Topography of the nicotinic acetylcholine receptor membraneembedded domains. J. Biol. Chem. 275:37333–37339.
- Cordero-Erausquin, M., Marubio, L. M., Klink, R., and Changeux, J. P. 2000. Nicotinic receptor function: New perspectives from knockout mice. *Trends Pharmacol. Sci.* 21:211–217.
- Le Novère, N., and Changeux, J. P. 1995. Molecular evolution of the nicotinic acetylcholine receptor: An example of multigene family in excitable cells. J. Mol. Evol. 40:155–172.
- Kunishima, N., Shimada, Y., Tsuji, Y., Sato, T., Yamamoto, M., Kumasaka, T., Nakanishi, S., Jingami, H., and Morikawa, K. 2000. Structural basis of glutamate recognition by dimeric metabotropic glutamate receptor. *Nature* 407:971–978.
- Betz, H., Kuhse, J., Schmieden, V., Laube, B., Kirsch, J., and Harvey, R. J. 1999. Structure and functions of inhibitory and excitatory glycine receptors. Ann. N. Y. Acad. Sci. 868:667–676.
- Unwin, N. 1995. Acetylcholine receptor channel imaged in the open state. *Nature* 373:37–43.
- Colquhoun, D., and Sakmann, B. 1981. Fluctuations in the microsecond time range of the current through single acetylcholine receptor ion channels. *Nature* 294:464–466.

Long QT Syndrome and hERG

- Saenen, J. B., and Vrints, C. J. 2008. Molecular aspects of the congenital and acquired Long QT Syndrome: clinical implications. J. Mol. Cell. Cardiol. 44:633–646.
- Zaręba, W. 2007. Drug induced QT prolongation. Cardiol. J. 14:523-533.
- Fernandez, D., Ghanta, A., Kauffman, G. W., and Sanguinetti, M. C. 2004. Physicochemical features of the hERG channel drug binding site. J. Biol. Chem. 279:10120–10127.
- Mitcheson, J. S., Chen, J., Lin, M., Culberson, C., and Sanguinetti, M. C. 2000. A structural basis for drug-induced long QT syndrome. *Proc. Natl. Acad. Sci. U.S.A.* 97:12329–12333.

Gap Junctions

- Saez, J. C., Berthoud, V. M., Branes, M. C., Martinez, A. D., and Beyer, E. C. 2003. Plasma membrane channels formed by connexins: Their regulation and functions. *Physiol. Rev.* 83:1359–1400.
- Revilla, A., Bennett, M. V. L., and Barrio, L. C. 2000. Molecular determinants of membrane potential dependence in vertebrate gap junction channels. *Proc. Natl. Acad. Sci. U.S.A.* 97:14760–14765.
- Unger, V. M., Kumar, N. M., Gilula, N. B., and Yeager, M. 1999. Threedimensional structure of a recombinant gap junction membrane channel. *Science* 283:1176–1180.
- Simon, A. M. 1999. Gap junctions: More roles and new structural data. Trends Cell Biol. 9:169–170.
- Beltramello, M., Piazza, V., Bukauskas, F. F., Pozzan, T., and Mammano, F. 2005. Impaired permeability to Ins(1,4,5)P₃ in a mutant connexin underlies recessive hereditary deafness. *Nat. Cell Biol.* 7:63–69.
- White, T. W., and Paul, D. L. 1999. Genetic diseases and gene knockouts reveal diverse connexin functions. *Annu. Rev. Physiol.* 61:283–310.

Water Channels

- Agre, P., King, L. S., Yasui, M., Guggino, W. B., Ottersen, O. P., Fujiyoshi, Y., Engel, A., and Nielsen, S. 2002. Aquaporin water channels: From atomic structure to clinical medicine. J. Physiol. 542:3–16.
- Agre, P., and Kozono, D. 2003. Aquaporin water channels: Molecular mechanisms for human diseases. *FEBS Lett.* 555:72–78.
- de Groot, B. L., Engel, A., and Grubmuller, H. 2003. The structure of the aquaporin-1 water channel: A comparison between cryoelectron microscopy and X-ray crystallography. J. Mol. Biol. 325:485–493.

Chapter 14

Where to Start

- Scott, J. D., and Pawson, T. 2000. Cell communication: The inside story. Sci. Am. 282(6):7279.
- Pawson, T. 1995. Protein modules and signalling networks. Nature 373:573-580.
- Okada, T., Ernst, O. P., Palczewski, K., and Hofmann, K. P. 2001. Activation of rhodopsin: New insights from structural and biochemical studies. *Trends Biochem. Sci.* 26:318–324.
- Tsien, R. Y. 1992. Intracellular signal transduction in four dimensions: From molecular design to physiology. Am. J. Physiol. 263:C723-C728.
- Loewenstein, W. R. 1999. Touchstone of Life: Molecular Information, Cell Communication, and the Foundations of Life. Oxford University Press.

G Proteins and 7TM Receptors

- Palczewski, K., Kumasaka, T., Hori, T., Behnke, C. A., Motoshima, H., Fox, B. A., Le Trong, I., Teller, D. C., Okada, T., Stenkamp, R. E., Yamamoto, M., and Miyano, M. 2000. Crystal structure of rhodopsin: A G protein-coupled receptor. *Science* 289:739–745.
- Rasmussen, S. G. F., Choi, H.-J., Rosenbaum, D. M., Kobilka, T. S., Thian, F. S., Edwards, P. C., Burghammer, M., Ratnala, V. R. P., Sanishvili, R., Fischetti, R. F., Schertler, G. F. X., Weis, W. I., and Kobilka, B. K. 2007. Crystal structure of the human β₂ adrenergic G-protein-coupled receptor. *Nature* 450:383–387.
- Rosenbaum, D. M., Cherezov, V., Hanson, M. A., Rasmussen, S. G. F., Thian, F. S., Kobilka, T. S., Choi, H.-J., Yao, X.-J., Weis, W. I., Stevens, R. C., and Kobilka, B. K. 2007. GPCR engineering yields high-resolution structural insights into β₂-adrenergic receptor function. *Science* 318:1266–1273.
- Lefkowitz, R. J. 2000. The superfamily of heptahelical receptors. *Nat. Cell Biol.* 2:E133–E136.
- Bourne, H. R., Sanders, D. A., and McCormick, F. 1991. The GTPase superfamily: Conserved structure and molecular mechanism. *Nature* 349:117–127.
- Lambright, D. G., Noel, J. P., Hamm, H. E., and Sigler, P. B. 1994. Structural determinants for activation of the α -subunit of a heterotrimeric G protein. *Nature* 369:621–628.
- Noel, J. P., Hamm, H. E., and Sigler, P. B. 1993. The 2.2 Å crystal structure of transducin-α complexed with GTPγS. *Nature* 366:654–663.
- Sondek, J., Lambright, D. G., Noel, J. P., Hamm, H. E., and Sigler, P. B. 1994. GTPase mechanism of G proteins from the 1.7-Å crystal structure of transducin α -GDP-AIF⁻⁴. *Nature* 372:276–279.
- Sondek, J., Bohm, A., Lambright, D. G., Hamm, H. E., and Sigler, P. B. 1996. Crystal structure of a G-protein βγ dimer at 2.1 Å resolution. *Nature* 379:369–374.
- Wedegaertner, P. B., Wilson, P. T., and Bourne, H. R. 1995. Lipid modifications of trimeric G proteins. J. Biol. Chem. 270:503–506.
- Farfel, Z., Bourne, H. R., and Iiri, T. 1999. The expanding spectrum of G protein diseases. N. Engl. J. Med. 340:1012–1020.
- Bockaert, J., and Pin, J. P. 1999. Molecular tinkering of G proteincoupled receptors: An evolutionary success. EMBO J. 18:1723–1729.

Cyclic AMP Cascade

- Hurley, J. H. 1999. Structure, mechanism, and regulation of mammalian adenylyl cyclase. J. Biol. Chem. 274:7599–7602.
- Kim, C., Xuong, N. H., and Taylor, S. S. 2005. Crystal structure of a complex between the catalytic and regulatory (RI) subunits of PKA. *Science* 307:690–696.
- Tesmer, J. J., Sunahara, R. K., Gilman, A. G., and Sprang, S. R. 1997. Crystal structure of the catalytic domains of adenylyl cyclase in a complex with G_{sα}-GTPγS. *Science* 278:1907–1916.
- Smith, C. M., Radzio-Andzelm, E., Madhusudan, Akamine, P., and Taylor, S. S. 1999. The catalytic subunit of cAMP-dependent protein kinase: Prototype for an extended network of communication. *Prog. Biophys. Mol. Biol.* 71:313–341.
- Taylor, S. S., Buechler, J. A., and Yonemoto, W. 1990. cAMP-dependent protein kinase: Framework for a diverse family of regulatory enzymes. Annu. Rev. Biochem. 59:971–1005.

Phosphoinositide Cascade

- Berridge, M. J., and Irvine, R. F. 1989. Inositol phosphates and cell signalling. *Nature* 341:197–205.
- Berridge, M. J. 1993. Inositol trisphosphate and calcium signalling. *Nature* 361:315–325.
- Essen, L. O., Perisic, O., Cheung, R., Katan, M., and Williams, R. L. 1996. Crystal structure of a mammalian phosphoinositide-specific phospholipase C δ. *Nature* 380:595–602.
- Ferguson, K. M., Lemmon, M. A., Schlessinger, J., and Sigler, P. B. 1995. Structure of the high affinity complex of inositol trisphosphate with a phospholipase C pleckstrin homology domain. *Cell* 83:1037–1046.
- Baraldi, E., Carugo, K. D., Hyvonen, M., Surdo, P. L., Riley, A. M., Potter, B. V., O'Brien, R., Ladbury, J. E., and Saraste, M. 1999. Structure of the PH domain from Bruton's tyrosine kinase in complex with inositol 1,3,4,5-tetrakisphosphate. *Struct. Fold. Design* 7:449–460.

Calcium

- Ikura, M., Clore, G. M., Gronenborn, A. M., Zhu, G., Klee, C. B., and Bax, A. 1992. Solution structure of a calmodulin-target peptide complex by multidimensional NMR. *Science* 256:632–638.
- Kuboniwa, H., Tjandra, N., Grzesiek, S., Ren, H., Klee, C. B., and Bax, A. 1995. Solution structure of calcium-free calmodulin. Nat. Struct. Biol. 2:768–776.
- Grynkiewicz, G., Poenie, M., and Tsien, R. Y. 1985. A new generation of Ca²⁺ indicators with greatly improved fluorescence properties. *J. Biol. Chem.* 260:3440–3450.
- Kerr, R., Lev-Ram, V., Baird, G., Vincent, P., Tsien, R. Y., and Schafer, W. R. 2000. Optical imaging of calcium transients in neurons and pharyngeal muscle of *C. elegans. Neuron* 26:583–594.
- Chin, D., and Means, A. R. 2000. Calmodulin: A prototypical calcium sensor. Trends Cell Biol. 10:322–328.
- Dawson, A. P. 1997. Calcium signalling: How do IP₃ receptors work? *Curr. Biol.* 7:R544–R547.

Protein Kinases, Including Receptor Tyrosine Kinases

- Riedel, H., Dull, T. J., Honegger, A. M., Schlessinger, J., and Ullrich, A. 1989. Cytoplasmic domains determine signal specificity, cellular routing characteristics and influence ligand binding of epidermal growth factor and insulin receptors. *EMBO J.* 8:2943–2954.
- Taylor, S. S., Knighton, D. R., Zheng, J., Sowadski, J. M., Gibbs, C. S., and Zoller, M. J. 1993. A template for the protein kinase family. *Trends Biochem. Sci.* 18:84–89.
- Sicheri, F., Moarefi, I., and Kuriyan, J. 1997. Crystal structure of the Src family tyrosine kinase Hck. Nature 385:602–609.
- Waksman, G., Shoelson, S. E., Pant, N., Cowburn, D., and Kuriyan, J. 1993. Binding of a high affinity phosphotyrosyl peptide to the Src SH2 domain: Crystal structures of the complexed and peptide-free forms. *Cell* 72:779–790.

- Schlessinger, J. 2000. Cell signaling by receptor tyrosine kinases. Cell 103:211–225.
- Simon, M. A. 2000. Receptor tyrosine kinases: Specific outcomes from general signals. Cell 103:13–15.
- Robinson, D. R., Wu, Y. M., and Lin, S. F. 2000. The protein tyrosine kinase family of the human genome. *Oncogene* 19:5548–5557.
- Hubbard, S. R. 1999. Structural analysis of receptor tyrosine kinases. Prog. Biophys. Mol. Biol. 71:343–358.
- Carter-Su, C., and Smit, L. S. 1998. Signaling via JAK tyrosine kinases: Growth hormone receptor as a model system. *Recent Prog. Horm. Res.* 53:61–82.

Insulin Signaling Pathway

- Khan, A. H., and Pessin, J. E. 2002. Insulin regulation of glucose uptake: A complex interplay of intracellular signalling pathways. *Diabetologia* 45:1475–1483.
- Bevan, P. 2001. Insulin signalling. J. Cell Sci. 114:1429-1430.
- De Meyts, P., and Whittaker, J. 2002. Structural biology of insulin and IGF1 receptors: Implications for drug design. *Nat. Rev. Drug Discov*. 1:769–783.
- Dhe-Paganon, S., Ottinger, E. A., Nolte, R. T., Eck, M. J., and Shoelson, S. E. 1999. Crystal structure of the pleckstrin homology-phosphotyrosine binding (PH-PTB) targeting region of insulin receptor substrate 1. Proc. Natl. Acad. Sci. U.S.A. 96:8378-8383.
- Domin, J., and Waterfield, M. D. 1997. Using structure to define the function of phosphoinositide 3-kinase family members. FEBS Lett. 410:91–95.
- Hubbard, S. R. 1997. Crystal structure of the activated insulin receptor tyrosine kinase in complex with peptide substrate and ATP analog. *EMBO J.* 16:5572–5581.
- Hubbard, S. R., Wei, L., Ellis, L., and Hendrickson, W. A. 1994. Crystal structure of the tyrosine kinase domain of the human insulin receptor. *Nature* 372:746–754.

EGF Signaling Pathway

- Burgess, A. W., Cho, H. S., Eigenbrot, C., Ferguson, K. M., Garrett, T. P., Leahy, D. J., Lemmon, M. A., Sliwkowski, M. X., Ward, C. W., and Yokoyama, S. 2003. An open-and-shut case? Recent insights into the activation of EGF/ErbB receptors. *Mol. Cell* 12:541–552.
- Cho, H. S., Mason, K., Ramyar, K. X., Stanley, A. M., Gabelli, S. B., Denney, D. W., Jr., and Leahy, D. J. 2003. Structure of the extracellular region of HER2 alone and in complex with the Herceptin Fab. *Nature* 421:756–760.
- Chong, H., Vikis, H. G., and Guan, K. L. 2003. Mechanisms of regulating the Raf kinase family. *Cell. Signal.* 15:463–469.
- Stamos, J., Sliwkowski, M. X., and Eigenbrot, C. 2002. Structure of the epidermal growth factor receptor kinase domain alone and in complex with a 4-anilinoquinazoline inhibitor. J. Biol. Chem. 277:46265–46272.

Ras

- Milburn, M. V., Tong, L., deVos, A. M., Brunger, A., Yamaizumi, Z., Nishimura, S., and Kim, S. H. 1990. Molecular switch for signal transduction: Structural differences between active and inactive forms of protooncogenic Ras proteins. *Science* 247:939–945.
- Boriack-Sjodin, P. A., Margarit, S. M., Bar-Sagi, D., and Kuriyan, J. 1998. The structural basis of the activation of Ras by Sos. *Nature* 394:337–343.
- Maignan, S., Guilloteau, J. P., Fromage, N., Arnoux, B., Becquart, J., and Ducruix, A. 1995. Crystal structure of the mammalian Grb2 adaptor. Science 268:291–293.
- Takai, Y., Sasaki, T., and Matozaki, T. 2001. Small GTP-binding proteins. *Physiol. Rev.* 81:153–208.

Cancer

- Druker, B. J., Sawyers, C. L., Kantarjian, H., Resta, D. J., Reese, S. F., Ford, J. M., Capdeville, R., and Talpaz, M. 2001. Activity of a specific inhibitor of the BCR-ABL tyrosine kinase in the blast crisis of chronic myeloid leukemia and acute lymphoblastic leukemia with the Philadelphia chromosome. N. Engl. J. Med. 344:1038–1042.
- Vogelstein, B., and Kinzler, K. W. 1993. The multistep nature of cancer. Trends Genet. 9:138–141.
- Ellis, C. A., and Clark, G. 2000. The importance of being K-Ras. Cell. Signal. 12:425–434.
- Hanahan, D., and Weinberg, R. A. 2000. The hallmarks of cancer. Cell 100:57–70.
- McCormick, F. 1999. Signalling networks that cause cancer. Trends Cell Biol. 9:M53–M56.

Chapter 15

Where to Start

- Stipanuk, M. H. (Ed.). 2006. Biochemical, Physiological, Molecular Aspects of Human Nutrition. Saunders-Elsevier.
- McGrane, M. M., Yun, J. S., Patel, Y. M., and Hanson, R. W. 1992. Metabolic control of gene expression: In vivo studies with transgenic mice. *Trends Biochem. Sci.* 17:40–44.
- Westheimer, F. H. 1987. Why nature chose phosphates. Science 235:1173–1178.

Books

- Atkins, P., and de Paula, J. 2005. *Physical Chemistry for the Life Sciences*. W. H. Freeman and Company.
- Harold, F. M. 1986. The Vital Force: A Study of Bioenergetics. W. H. Freeman and Company.
- Krebs, H. A., and Kornberg, H. L. 1957. Energy Transformations in Living Matter. Springer Verlag.
- Nicholls, D. G., and Ferguson, S. J. 2002. *Bioenergetics 3* (3d ed.). Academic Press.
- Frayn, K. N. 2010. Metabolic Regulation: A Human Perspective (3d ed.). Wiley-Blackwell.
- Fell, D. 1997. Understanding the Control of Metabolism. Portland Press.
- Harris, D. A. 1995. Bioenergetics at a Glance. Blackwell Scientific.

Von Baeyer, H. C. 1999. Warmth Disperses and Time Passes: A History of Heat. Modern Library.

Thermodynamics

- Alberty, R. A. 1993. Levels of thermodynamic treatment of biochemical reaction systems. *Biophys. J.* 65:1243–1254.
- Alberty, R. A., and Goldberg, R. N. 1992. Standard thermodynamic formation properties for the adenosine 5'-triphosphate series. *Biochemistry* 31:10610–10615.
- Alberty, R. A. 1968. Effect of pH and metal ion concentration on the equilibrium hydrolysis of adenosine triphosphate to adenosine diphosphate. J. Biol. Chem. 243:1337–1343.
- Goldberg, R. N. 1984. Compiled Thermodynamic Data Sources for Aqueous and Biochemical Systems: An Annotated Bibliography (1930–1983). National Bureau of Standards Special Publication 685, U.S. Government Printing Office.
- Frey, P. A., and Arabshahi, A. 1995. Standard free energy change for the hydrolysis of the α,β-phosphoanhydride bridge in ATP. *Biochemistry* 34:11307–11310.

Bioenergetics and Metabolism

- Schilling, C. H., Letscher, D., and Palsson, B. O. 2000. Theory for the systemic definition of metabolic pathways and their use in interpreting metabolic function from a pathway-oriented perspective. *J. Theor. Biol.* 203:229–248.
- DeCoursey, T. E., and Cherny, V. V. 2000. Common themes and problems of bioenergetics and voltage-gated proton channels. *Biochim. Biophys. Acta* 1458:104–119.

- Giersch, C. 2000. Mathematical modelling of metabolism. Curr. Opin. Plant Biol. 3:249–253.
- Rees, D. C., and Howard, J. B. 1999. Structural bioenergetics and energy transduction mechanisms. J. Mol. Biol. 293:343-350.

Regulation of Metabolism

- Kemp, G. J. 2000. Studying metabolic regulation in human muscle. Biochem. Soc. Trans. 28:100–103.
- Towle, H. C., Kaytor, E. N., and Shih, H. M. 1996. Metabolic regulation of hepatic gene expression. *Biochem. Soc. Trans.* 24:364–368.
- Hofmeyr, J. H. 1995. Metabolic regulation: A control analytic perspective. J. Bioenerg. Biomembr. 27:479–490.

Historical Aspects

Kalckar, H. M. 1991. 50 years of biological research: From oxidative phosphorylation to energy requiring transport regulation. Annu. Rev. Biochem. 60:1–37.

Kalckar, H. M. (Ed.). 1969. Biological Phosphorylations. Prentice Hall. Fruton, J. S. 1972. Molecules and Life. Wiley-Interscience.

Lipmann, F. 1971. Wanderings of a Biochemist. Wiley-Interscience.

Chapter 16

Where to Start

- Knowles, J. R. 1991. Enzyme catalysis: Not different, just better. Nature 350:121–124.
- Granner, D., and Pilkis, S. 1990. The genes of hepatic glucose metabolism. J. Biol. Chem. 265:10173–10176.
- McGrane, M. M., Yun, J. S., Patel, Y. M., and Hanson, R. W. 1992. Metabolic control of gene expression: In vivo studies with transgenic mice. *Trends Biochem. Sci.* 17:40–44.
- Pilkis, S. J., and Granner, D. K. 1992. Molecular physiology of the regulation of hepatic gluconeogenesis and glycolysis. Annu. Rev. Physiol. 54:885–909.

Books

- Frayn, K. N. 2010. Metabolic Regulation: A Human Perspective (3d ed.). Wiley-Blackwell.
- Fell, D. 1997. Understanding the Control of Metabolism. Portland.
- Fersht, A. 1999. Structure and Mechanism in Protein Science: A Guide to Enzyme Catalysis and Protein Folding. W. H. Freeman and Company.

Poortmans, J. R. (Ed.). 2004. Principles of Exercise Biochemistry. Krager.

Structure of Glycolytic and Gluconeogenic Enzymes

- Ferreras, C., Hernández, E. D., Martínez-Costa, O. H., and Aragón, J. J. 2009. Subunit interactions and composition of the fructose 6-phosphate catalytic site and the fructose 2,6-bisphosphate allosteric site of mammalian phosphofructokinase. J. Biol. Chem. 284:9124–9131.
- Hines, J. K., Chen, X., Nix, J. C., Fromm, H. J., and Honzatko. R. B. 2007. Structures of mammalian and bacterial fructose-1, 6-bisphosphatase reveal the basis for synergism in AMP/fructose-2, 6-bisphosphate inhibition. J. Biol. Chem. 282:36121–36131.
- Ferreira-da-Silva, F., Pereira, P. J., Gales, L., Roessle, M., Svergun, D. I., Moradas-Ferreira, P., and Damas, A. M. 2006. The crystal and solution structures of glyceraldehyde-3-phosphate dehydrogenase reveal different quaternary structures. J. Biol. Chem. 281:33433–33440.
- Kim, S.-G., Manes, N. P., El-Maghrabi, M. R., and Lee, Y.-H. 2006. Crystal structure of the hypoxia-inducible form of 6-phosphofructo-2-kinase/fructose-2,6-phosphatase (PFKFB3): A possible target for cancer therapy. J. Biol. Chem. 281:2939–2944.
- Aleshin, A. E., Kirby, C., Liu, X., Bourenkov, G. P., Bartunik, H. D., Fromm, H. J., and Honzatko, R. B. 2000. Crystal structures of mutant monomeric hexokinase I reveal multiple ADP binding

sites and conformational changes relevant to allosteric regulation. *J. Mol. Biol.* 296:1001–1015.

- Jeffery, C. J., Bahnson, B. J., Chien, W., Ringe, D., and Petsko, G. A. 2000. Crystal structure of rabbit phosphoglucose isomerase, a glycolytic enzyme that moonlights as neuroleukin, autocrine motility factor, and differentiation mediator. *Biochemistry* 39:955–964.
- Schirmer, T., and Evans, P. R. 1990. Structural basis of the allosteric behaviour of phosphofructokinase. *Nature* 343:140–145.
- Cooper, S. J., Leonard, G. A., McSweeney, S. M., Thompson, A. W., Naismith, J. H., Qamar, S., Plater, A., Berry, A., and Hunter, W. N. 1996. The crystal structure of a class II fructose-l,6-bisphosphate aldolase shows a novel binuclear metal-binding active site embedded in a familiar fold. *Structure* 4:1303–1315.
- Davenport, R. C., Bash, P. A., Seaton, B. A., Karplus, M., Petsko, G. A., and Ringe, D. 1991. Structure of the triosephosphate isomerase– phosphoglycolohydroxamate complex: An analogue of the intermediate on the reaction pathway. *Biochemistry* 30:5821–5826.
- Bernstein, B. E., and Hol, W. G. 1998. Crystal structures of substrates and products bound to the phosphoglycerate kinase active site reveal the catalytic mechanism. *Biochemistry* 37:4429–4436.
- Rigden, D. J., Alexeev, D., Phillips, S. E. V., and Fothergill-Gilmore, L. A. 1998. The 2.3 Å X-ray crystal structure of S. cerevisiae phosphoglycerate mutase. J. Mol. Biol. 276:449–459.
- Zhang, E., Brewer, J. M., Minor, W., Carreira, L. A., and Lebioda, L. 1997. Mechanism of enolase: The crystal structure of asymmetric dimer enolase-2-phospho-D-glycerate/enolase-phosphoenolpyruvate at 2.0 Å resolution. *Biochemistry* 36:12526–12534.
- Mattevi, A., Valentini, G., Rizzi, M., Speranza, M. L., Bolognesi, M., and Coda, A. 1995. Crystal structure of *Escherichia coli* pyruvate kinase type I: Molecular basis of the allosteric transition. *Structure* 3:729–741.
- Hasemann, C. A., Istvan E. S., Uyeda, K., and Deisenhofer, J. 1996. The crystal structure of the bifunctional enzyme 6-phosphofructo-2-kinase/fructose-2,6-biphosphatase reveals distinct domain homologies. *Structure* 4:1017–1029.
- Tari, L. W., Matte, A., Pugazhenthi, U., Goldie, H., and Delbaere, L. T. J. 1996. Snapshot of an enzyme reaction intermediate in the structure of the ATP-Mg²⁺-oxalate ternary complex of *Escherichia coli* PEP carboxykinase. *Nat. Struct. Biol.* 3:355–363.

Catalytic Mechanisms

- Soukri, A., Mougin, A., Corbier, C., Wonacott, A., Branlant, C., and Branlant, G. 1989. Role of the histidine 176 residue in glyceraldehyde-3-phosphate dehydrogenase as probed by site-directed mutagenesis. *Biochemistry* 28:2586–2592.
- Bash, P. A., Field, M. J., Davenport, R. C., Petsko, G. A., Ringe, D., and Karplus, M. 1991. Computer simulation and analysis of the reaction pathway of triosephosphate isomerase. *Biochemistry* 30:5826–5832.
- Knowles, J. R., and Albery, W. J. 1977. Perfection in enzyme catalysis: The energetics of triosephosphate isomerase. Acc. Chem. Res. 10:105–111.
- Rose, I. A. 1981. Chemistry of proton abstraction by glycolytic enzymes (aldolase, isomerases, and pyruvate kinase). *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 293:131–144.

Regulation

- Anderka, O., Boyken, J., Aschenbach, U., Batzer, A., Boscheinen, O., and Schmoll, D. 2008. Biophysical characterization of the interaction between hepatic glucokinase and its regulatory protein: Impact of physiological and pharmacological effectors. J. Biol. Chem. 283:31333–31340.
- Iancu, C. V., Mukund, S., Fromm, H. J., and Honzatko, R. B. 2005. R-state AMP complex reveals initial steps of the quaternary transition of fructose-l,6-bisphosphatase. J. Biol. Chem. 280:19737–19745.
- Wilson, J. E. 2003. Isozymes of mammalian hexokinase: Structure, function and subcellular location. J. Exp. Biol. 206:2049–2057.

SELECTED READINGS

- Lee, Y. H., Li, Y., Uyeda, K., and Hasemann, C. A. 2003. Tissuespecific structure/function differentiation of the five isoforms of 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase. J. Biol. Chem. 278:523–530.
- Depre, C., Rider, M. H., and Hue, L. 1998. Mechanisms of control of heart glycolysis. *Eur. J. Biochem.* 258:277–290.
- Harrington, G. N., and Bush, D. R. 2003. The bifunctional role of hexokinase in metabolism and glucose signaling. *Plant Cell* 15:2493–2496.
- Gleeson, T. T. 1996. Post-exercise lactate metabolism: A comparative review of sites, pathways, and regulation. Annu. Rev. Physiol. 58:556–581.
- Nordlie, R. C., Foster, J. D., and Lange, A. J. 1999. Regulation of glucose production by the liver. Annu. Rev. Nutr. 19:379–406.
- Jitrapakdee, S., and Wallace, J. C. 1999. Structure, function and regulation of pyruvate carboxylase. *Biochem. J.* 340:1–16.
- Pilkis, S. J., and Claus, T. H. 1991. Hepatic gluconeogenesis/glycolysis: Regulation and structure/function relationships of substrate cycle enzymes. Annu. Rev. Nutr. 11:465–515.
- Plaxton, W. C. 1996. The organization and regulation of plant glycolysis. Annu. Rev. Plant Physiol. Plant Mol. Biol. 47:185–214.
- van de Werve, G., Lange, A., Newgard, C., Mechin, M. C., Li, Y., and Berteloot, A. 2000. New lessons in the regulation of glucose metabolism taught by the glucose 6-phosphatase system. *Eur. J. Biochem.* 267:1533–1549.

Sugar Transporters

- Blodgett, D. M., Graybill, C. and Carruthers, A. 2008. Analysis of glucose transporter topology and structural dynamics. J. Biol. Chem. 283: 36416–36424.
- Huang, S., and Czech, M. P. 2007. The GLUT4 glucose transporter. Cell Metab. 5:237–252.
- Czech, M. P., and Corvera, S. 1999. Signaling mechanisms that regulate glucose transport. J Biol. Chem. 274:1865–1868.
- Silverman, M. 1991. Structure and function of hexose transporters. Annu. Rev. Biochem. 60:757-794.
- Thorens, B., Charron, M. J., and Lodish, H. F. 1990. Molecular physiology of glucose transporters. *Diabetes Care* 13:209–218.

Glycolysis and Cancer

- Vander Heiden, M. G., Cantley, L. C., and Thompson, C. B. 2009. Understanding the Warburg effect: The metabolic requirements of cell proliferation. *Science* 324:1029–1033.
- Mathupala, S. P., Ko, Y. H., and Pedersen, P. L. 2009. Hexokinase-2 bound to mitochondria: Cancer's stygian link to the "Warburg effect" and a pivotal target for effective therapy. Sem. Cancer Biol. 19:17–24.
- Kroemer, G. K., and Pouyssegur, J. 2008. Tumor cell metabolism: Cancer's Achilles' heel. *Cancer Cell* 12:472–482.
- Hsu, P. P., and Sabatini, D. M. 2008. Cancer cell metabolism: Warburg and beyond. *Cell* 134:703–707.
- Dang, C. V., and Semenza, G. L. 1999. Oncogenic alterations of metabolism. Trends Biochem. Sci. 24:68-72.

Genetic Diseases

Scriver, C. R., Beaudet, A. L., Valle, D., Sly, W. S., Childs, B., Kinzler, K., and Vogelstein, B. (Eds.). 2001. The Metabolic and Molecular Basis of Inherited Disease (8th ed.). McGraw-Hill.

Evolution

- Dandekar, T., Schuster, S., Snel, B., Huynen, M., and Bork, P. 1999. Pathway alignment: Application to the comparative analysis of glycolytic enzymes. *Biochem. J.* 343:115–124.
- Heinrich, R., Melendez-Hevia, E., Montero, F., Nuno, J. C., Stephani, A., and Waddell, T. G. 1999. The structural design of glycolysis: An evolutionary approach. *Biochem. Soc. Trans.* 27:294–298.

- Walmsley, A. R., Barrett, M. P., Bringaud, F., and Gould, G. W. 1998. Sugar transporters from bacteria, parasites and mammals: Structure-activity relationships. *Trends Biochem. Sci.* 23:476–480.
- Maes, D., Zeelen, J. P., Thanki, N., Beaucamp, N., Alvarez, M., Thi, M. H., Backmann, J., Martial, J. A., Wyns, L., Jaenicke, R., and Wierenga, R. K. 1999. The crystal structure of triosephosphate isomerase (TIM) from *Thermotoga maritima*: A comparative thermostability structural analysis of ten different TIM structures. *Proteins* 37:441–453.

Historical Aspects

- Friedmann, H. C. 2004. From Butyribacterium to E. coli: An essay on unity in biochemistry. Perspect. Biol. Med. 47:47–66.
- Fruton, J. S. 1999. Proteins, Enzymes, Genes: The Interplay of Chemistry and Biology. Yale University Press.
- Kalckar, H. M. (Ed.). 1969. Biological Phosphorylations: Development of Concepts. Prentice Hall.

Chapter 17

Where to Start

- Sugden, M. C., and Holness, M. J. 2003. Recent advances in mechanisms regulating glucose oxidation at the level of the pyruvate dehydrogenase complex by PDKs. Am. J. Physiol. Endocrinol. Metab. 284:E855-E862.
- Owen, O. E., Kalhan, S. C., and Hanson, R. W. 2002. The key role of anaplerosis and cataplerosis for citric acid function. J. Biol. Chem. 277:30409–30412.

Pyruvate Dehydrogenase Complex

- Hiromasa, Y., Fujisawa, T., Aso, Y., and Roche, T. E. 2004. Organization of the cores of the mammalian pyruvate dehydrogenase complex formed by E2 and E2 plus the E3-binding proteins and their capacities to bind the E1 and E3 components. *J. Biol Chem.* 279:6921–6933.
- Izard, T., Ævarsson, A., Allen, M. D., Westphal, A. H., Perham, R. N., De Kok, A., and Hol, W. G. 1999. Principles of quasi-equivalence and Euclidean geometry govern the assembly of cubic and dodecahedral cores of pyruvate dehydrogenase complexes. *Proc. Natl. Acad. Sci. U.S.A.* 96:1240–1245.
- Domingo, G. J., Chauhan, H. J., Lessard, I. A., Fuller, C., and Perham, R. N. 1999. Self-assembly and catalytic activity of the pyruvate dehydrogenase multienzyme complex from *Bacillus stearothermophilus. Eur. J. Biochem.* 266:1136–1146.
- Jones, D. D., Horne, H. J., Reche, P. A., and Perham, R. N. 2000. Structural determinants of post-translational modification and catalytic specificity for the lipoyl domains of the pyruvate dehydrogenase multienzyme complex of *Escherichia coli. J. Mol. Biol.* 295:289–306.

Structure of Citric Acid Cycle Enzymes

- Fraser, M. E. Hayakawa, K., Hume, M. S., Ryan, D. G., and Brownie, E. R. 2006. Interactions of GTP with the ATP-grasp domain of GTPspecific succinyl-CoA synthetase. J. Biol. Chem. 281:11058–11065.
- Yankovskaya, V., Horsefield, R., Törnroth, S., Luna-Chavez, C., Miyoshi, H., Léger, C., Byrne, B., Cecchini, G., and Iowata, S. 2003. Architecture of succinate dehydrogenase and reactive oxygen species generation. *Science* 299:700–704.
- Chapman, A. D., Cortes, A., Dafforn, T. R., Clarke, A. R., and Brady, R. L. 1999. Structural basis of substrate specificity in malate dehydrogenases: Crystal structure of a ternary complex of porcine cytoplasmic malate dehydrogenase, α-ketomalonate and tetrahydoNAD. J. Mol. Biol. 285:703–712.
- Fraser, M. E., James, M. N., Bridger, W. A., and Wolodko, W. T. 1999. A detailed structural description of *Escherichia coli* succinyl-CoA

synthetase. J. Mol. Biol. 285:1633-1653. [Published erratum appears in May 7, 1999, issue of J. Mol. Biol. 288(3):501.]

- Lloyd, S. J., Lauble, H., Prasad, G. S., and Stout, C. D. 1999. The mechanism of aconitase: 1.8 Å resolution crystal structure of the S642a:citrate complex. *Protein Sci.* 8:2655–2662.
- Remington, S. J. 1992. Structure and mechanism of citrate synthase. Curr. Top. Cell. Regul. 33:209–229.
- Rose, I. A. 1998. How fumarase recycles after the malate → fumarate reaction: Insights into the reaction mechanism. *Biochemistry* 37: 17651–17658.

Organization of the Citric Acid Cycle

- Lambeth, D. O., Tews, K. N., Adkins, S., Frohlich, D., and Milavetz, B. I. 2004. Expression of two succinyl-CoA specificities in mammalian tissues. J. Biol. Chem. 279:36621–36624.
- Velot, C., Mixon, M. B., Teige, M., and Srere, P. A. 1997. Model of a quinary structure between Krebs TCA cycle enzymes: A model for the metabolon. *Biochemistry* 36:14271–14276.
- Haggie, P. M., and Brindle, K. M. 1999. Mitochondrial citrate synthase is immobilized in vivo. J. Biol. Chem. 274:3941–3945.
- Morgunov, I., and Srere, P. A. 1998. Interaction between citrate synthase and malate dehydrogenase: Substrate channeling of oxaloacetate. J. Biol. Chem. 273:29540–29544.

Regulation

- Phillips, D., Aponte, A. M., French, S. A., Chess, D. J., and Balaban, R. S. 2009. Succinyl-CoA synthetase is a phosphate target for the activation of mitochondrial metabolism. *Biochemistry* 48: 7140–7149.
- Taylor, A. B., Hu, G., Hart, P. J., and McAlister-Henn, L. 2008. Allosteric motions in structures of yeast NAD⁺-specific isocitrate dehydrogenase. J. Biol. Chem. 283: 10872–10880.
- Green, T., Grigorian, A., Klyuyeva, A., Tuganova, A., Luo, M., and Popov, K. M. 2008. Structural and functional insights into the molecular mechanisms responsible for the regulation of pyruvate dehydrogenase kinase. J. Biol. Chem. 283: 15789–15798.
- Hiromasa, Y., and Roche, T. E. 2003. Facilitated interaction between the pyruvate dehydrogenase kinase isoform 2 and the dihydrolipoyl acetyltransferases. *J. Biol. Chem.* 278:33681–33693.
- Jitrapakdee, S., and Wallace, J. C. 1999. Structure, function and regulation of pyruvate carboxylase. *Biochem. J.* 340:1–16.

The Citric Acid Cycle and Cancer

- Thompson, C. B. 2009. Metabolic enzymes as oncogenes or tumor suppressors. New Engl. J. Med. 360:813–815.
- McFate, T., Mohyeldin, A., Lu, H., Thakar, J., Henriques, J., Halim, N. D., Wu, H., Schell, M. J., Tsang, T. M., Teahan, O., Zhou, S., Califano, J. A., Jeoung, M. N., Harris, R. A., and Verma, A. 2008. Pyruvate dehydrogenase complex activity controls metabolic and malignant phenotype in cancer cells. J. Biol. Chem. 283: 22700–22708.
- Gogvadze, V., Orrenius, S., and Zhivotovsky, B. 2008. Mitochondria in cancer cells: What is so special about them? *Trends Cell Biol.* 18:165–173.

Evolutionary Aspects

- Meléndez-Hevia, E., Waddell, T. G., and Cascante, M. 1996. The puzzle of the Krebs citric acid cycle: Assembling the pieces of chemically feasible reactions, and opportunism in the design of metabolic pathways in evolution. J. Mol. Evol. 43:293–303.
- Baldwin, J. E., and Krebs, H. 1981. The evolution of metabolic cycles. *Nature* 291:381–382.
- Gest, H. 1987. Evolutionary roots of the citric acid cycle in prokaryotes. Biochem. Soc. Symp. 54:3–16.
- Weitzman, P. D. J. 1981. Unity and diversity in some bacterial citric acid cycle enzymes. Adv. Microbiol. Physiol. 22:185–244.

Discovery of the Citric Acid Cycle

- Kornberg, H. 2000. Krebs and his trinity of cycles. Nat. Rev. Mol. Cell. Biol. 1:225–228.
- Krebs, H. A., and Johnson, W. A. 1937. The role of citric acid in intermediate metabolism in animal tissues. *Enzymologia* 4:148–156.
- Krebs, H. A. 1970. The history of the tricarboxylic acid cycle. Perspect. Biol. Med. 14:154–170.
- Krebs, H. A., and Martin, A. 1981. *Reminiscences and Reflections*. Clarendon Press.

Chapter 18

Where to Start

- Guarente, L. 2008. Mitochondria: A nexus for aging, calorie restriction, and sirtuins? Cell 132:171–176.
- Wallace, D. C. 2007. Why do we still have a maternally inherited mitochondrial DNA? Insights from evolutionary medicine. Annu. Rev. Biochem. 76:781–821.
- Brandt, U. 2006. Energy converting NADH:quinone oxidoreductase (Complex I). Annu Rev Biochem. 75:69–92.
- Hosler, J. P., Ferguson-Miller, S., and Mills, D. A. 2006. Energy transduction: Proton transfer through the respiratory complexes. *Annu. Rev. Biochem.* 75:165–187.
- Gray, M. W., Burger, G., and Lang, B. F. 1999. Mitochondrial evolution. *Science* 283:1476–1481.
- Shultz, B. E., and Chan, S. I. 2001. Structures and proton-pumping strategies of mitochondrial respiratory enzymes. *Annu. Rev. Biophys. Biomol. Struct.* 30:23–65.

Books

- Scheffler, I. E. 2007. Mitochondria. Wiley.
- Lane, N. 2005. Power, Sex, Suicide: Mitochondria and the Meaning of Life. Oxford.
- Nicholls, D. G., and Ferguson, S. J. 2002. Bioenergetics 3. Academic Press.

Electron-Transport Chain

- Qin, L., Liu, J., Mills, D. A., Proshlyakov, D. A., Hiser, C., and Ferguson-Miller, S. 2009. Redox-dependent conformational changes in cytochrome c oxidase suggest a gating mechanism for proton uptake. *Biochemistry* 48:5121–5130.
- Lill, R. 2009. Function and biogenesis of iron-sulphur proteins. Nature 460:831–838.
- Cooley, C. W., Lee, D.-W., and Daldal, F. 2009. Across membrane communication between the Q_o and Q_i active sites of cytochrome bc₁. Biochemistry 48:1888–1899.
- Verkhovskaya, M. L., Belevich, N., Euro, L., Wikström, M., and. Verkhovsky, M. I. 2008. Real-time electron transfer in respiratory complex I. Proc. Natl. Acad. Sci. U.S.A. 105:3763–3767.
- Berrisford, J. M., and Sazanov, L. A. 2009. Structural basis for the mechanism of respiratory complex I. J. Biol. Chem. 284:29773–29783.
- Acín-Pérez, R., Fernández-Silva, P., Peleato, M. L., Pérez-Martos, A., and Enriquez, J. A. 2008. Respiratory active mitochondrial supercomplexes. *Molecular Cell* 32:529–539.
- Kruse, S. E., Watt, W. C., Marcinek, D. J., Kapur, R. P., Schenkman, K. A., and Palmiter, R. D. 2008. Mice with mitochondrial Complex I deficiency develop a fatal encephalomyopathy. *Cell Metab.* 7:312–320.
- Belevich, I., Verkhovsky, M. I., and Wikström, M. 2007. Protoncoupled electron transfer drives the proton pump of cytochrome *c* oxidase. *Nature* 440:829–832.
- Sun, F., Huo, X., Zhai, Y., Wang, A., Xu, J., Su, D., Bartlam, M., and Ral, Z. 2005. Crystal structure of mitochondrial respiratory membrane protein complex II. *Cell* 121:1043–1057.
- Crofts, A. R. 2004. The cytochrome bc₁ complex: Function in the context of structure. Annu. Rev. Physiol. 66:689–733.
- Bianchi, C., Genova, M. L., Castelli, G. P., and Lenaz, G. 2004. The mitochondrial respiratory chain is partially organized in a supramolecular complex. J. Biol. Chem. 279:36562–36569.

SELECTED READINGS

- Ugalde, C., Vogel, R., Huijbens, R., van den Heuvel, B., Smeitink, J., and Nijtmans, L. 2004. Human mitochondrial Complex I assembles through a combination of evolutionary conserved modules: A framework to interpret Complex I deficiencies. *Hum. Mol. Genet.* 13:2461–2472.
- Yagi, T., and Matsuno-Yagi, A. 2003. The proton-translocating NADHquinone oxidoreductase in the respiratory chain: The secret unlocked. *Biochemistry* 42:2266–2274.
- Cecchini, G. 2003. Function and structure of Complex II of the respiratory chain. Annu. Rev. Biochem. 72:77–109.
- Lange, C., and Hunte, C. 2002. Crystal structure of the yeast cytochrome bc₁ complex with its bound substrate cytochrome c. Proc. Natl. Acad. Sci. U.S.A. 99:2800–2805.

ATP Synthase

- Wittig, I., and Hermann, S. 2009. Supramolecular organization of ATP synthase and respiratory chain in mitochondrial membranes. *Biochim. Biophys. Acta* 1787:672–680.
- Junge, W., Sielaff, H., and Engelbrecht S. 2009. Torque generation and elastic power transmission in the rotary $F_0F_1\text{-}ATPase.\ Nature\ 459:364–370.$
- von Ballmoos, C., Cook, G. M., and Dimroth, P. 2008. Unique rotary ATP synthase and its biological diversity. Annu. Rev. Biophys. 37:43-64.
- Adachi, K., Oiwa, K., Nishizaka, T., Furuike, S., Noji, H., Itoh, H., Yoshida, M., and Kinosita, K., Jr. 2007. Coupling of rotation and catalysis in F₁-ATPase revealed by single-molecule imaging and manipulation. *Cell* 130:309–321.
- Chen, C., Ko, Y., Delannoy, M., Ludtke, S. J., Chiu, W., and Pedersen, P. L. 2004. Mitochondrial ATP synthasome: Three-dimensional structure by electron microscopy of the ATP synthase in complex formation with the carriers for P_i and ADP/ATP. J. Biol. Chem. 279:31761–31768.
- Noji, H., and Yoshida, M. 2001. The rotary machine in the cell: ATP synthase. J. Biol. Chem. 276:1665–1668.
- Yasuda, R., Noji, H., Kinosita, K., Jr., and Yoshida, M. 1998. F₁-ATPase is a highly efficient molecular motor that rotates with discrete 120 degree steps. *Cell* 93:1117–1124.
- Kinosita, K., Jr., Yasuda, R., Noji, H., Ishiwata, S., and Yoshida, M. 1998.
 F₁-ATPase: A rotary motor made of a single molecule. *Cell* 93:21–24.
- Noji, H., Yasuda, R., Yoshida, M., and Kinosita, K., Jr., 1997. Direct observation of the rotation of F₁-ATPase. *Nature* 386:299–302.
- Tsunoda, S. P., Aggeler, R., Yoshida, M., and Capaldi, R. A. 2001. Rotation of the *c* subunit oligomer in fully functional $F_1 F_0$ ATP synthase. *Proc. Natl. Acad. Sci. U.S.A.* 987:898–902.
- Gibbons, C., Montgomery, M. G., Leslie, A. G. W., and Walker, J. 2000. The structure of the central stalk in F₁-ATPase at 2.4 Å resolution. *Nat. Struct. Biol.* 7:1055–1061.
- Sambongi, Y., Iko, Y., Tanabe, M., Omote, H., Iwamoto-Kihara, A., Ueda, I., Yanagida, T., Wada, Y., and Futai, M. 1999. Mechanical rotation of the *c* subunit oligomer in ATP synthase (F₀F₁): Direct observation. *Science* 286:1722–1724.

Translocators and Channels

- van Marken Lichtenbelt, W. D., Vanhommerig, J. W., Smulders, N. M., Drossaerts, J. M., Kemerink, G. J., Bouvy, N. D., Schrauwen, P., and Teule, G. J. 2009. Cold-activated brown adipose tissue in healthy men. N. Engl. J. Med. 360:1500–1508.
- Cypess, A. M., Sanaz Lehman, S., Gethin Williams, G., Tal, I., Rodman, D., Goldfine, A. B., Kuo, F. C., Palmer, E. L., Tseng, Y.-H., Doria, A., Kolodny, G. M., and Kahn, C. R. 2009. Identification and importance of brown adipose tissue in adult humans. N. Engl. J. Med. 360:1509–1517.
- Virtanen, K. A., Lidell, M. E., Orava, J., Heglind, M., Westergren, R., Niemi, T., Taittonen, M., Laine, J., Savisto, N.-J., Enerbäck, S., and Nuutila, P. 2009. Functional brown adipose tissue in healthy adults. N. Engl. J. Med. 360:1518–1525.

- Bayrhuber, M., Meins, T., Habeck, M., Becker, S., Giller, K., Villinger, S., Vonrhein, C., Griesinger, C., Zweckstetter, M.,and Zeth, K. 2008. Structure of the human voltage-dependent anion channel. *Proc. Natl. Acad. Sci. U.S.A.* 105:15370–15375.
- Bamber, L., Harding, M., Monné, M., Slotboom, D.-J. and Kunji, E. R. 2007. The yeast mitochoondrial ADP/ATP carrier functions as a monomer in mitochondrial membranes. *Proc. Natl. Acad. Sci.* U. S. A. 10:10830–10843.
- Pebay-Peyroula, E., Dahout, C., Kahn, R., Trézéguet, V., Lauquin, G. J.-M., and Brandolin, G. 2003. Structure of mitochondrial ADP/ATP carrier in complex with carboxyatractyloside. *Nature* 246: 39–44.
- Nicholls, D. G., and Rial, E. 1999. A history of the first uncoupling protein, UCP1. J. Bioenerg. Biomembr. 31:399-406.
- Ricquier, D., and Bouillaud, F. 2000. The uncoupling protein homologues: UCP1, UCP2, UCP3, StUCP and AtUCP. *Biochem.* J. 345:161–179.

Reactive Oxygen Species, Superoxide Dismutase, and Catalase

Forman, H. J., Maiorino, M., and Ursini, F. 2010. Signaling functions of reactive oxygen species. *Biochemistry* 49:835–842.

- Murphy, M. P. 2009. How mitochondria produce reactive oxygen species. *Biochem. J.* 417:1–13.
- Leitch, J. M., Yick, P. J., and Culotta, V. V. 2009. The right to choose: Multiple pathways for activating copper, zinc superoxide dismutase. J. Biol. Chem. 284: 24679–24683.
- Winterbourn, C. C. 2008. Reconciling the chemistry and biology of reactive oxygen species. Nat. Chem. Biol. 4:278–286.
- Veal, E. A., Day, A. M., and Morgan, B. A. 2007. Hydrogen peroxide sensing and signaling. *Mol. Cell* 26:1–14.
- Stone, J. R., and Yang, S. 2006. Hydrogen peroxide: A signaling messenger. Antioxid. Redox Signal. 8(3-4):243-270.
- Acker, H. 2005. The oxygen sensing signal cascade under the influence of reactive oxygen species. *Phil. Trans. R. Soc. B* 360:2201–2210.
- Valentine, J. S., Doucette, P. A., and Potter S. Z. 2005. Copper-zinc superoxide dismutase and amyotrophic lateral sclerosis. Annu. Rev. Biochem. 74:563–593.
- Culotta, V. C. 2000. Superoxide dismutase, oxidative stress, and cell metabolism. *Curr. Top. Cell Regul.* 36:117-132.
- Morrison, B. M., Morrison, J. H., and Gordon, J. W. 1998. Superoxide dismutase and neurofilament transgenic models of amyotrophic lateral sclerosis. J. Exp. Zool. 282:32–47.
- Tainer, J. A., Getzoff, E. D., Richardson, J. S., and Richardson, D. C. 1983. Structure and mechanism of copper, zinc superoxide dismutase. *Nature* 306:284–287.
- Reid, T. J., Murthy, M. R., Sicignano, A., Tanaka, N., Musick, W. D., and Rossmann, M. G. 1981. Structure and heme environment of beef liver catalase at 2.5 Å resolution. *Proc. Natl. Acad. Sci.* U. S. A. 78:4767–4771.
- Stallings, W. C., Pattridge, K. A., Strong, R. K., and Ludwig, M. L. 1984. Manganese and iron superoxide dismutases are structural homologs. J. Biol. Chem. 259:10695–10699.
- Hsieh, Y., Guan, Y., Tu, C., Bratt, P. J., Angerhofer, A., Lepock, J. R., Hickey, M. J., Tainer, J. A., Nick, H. S., and Silverman, D. N. 1998. Probing the active site of human manganese superoxide dismutase: The role of glutamine 143. *Biochemistry* 37:4731–4739.

Mitochondrial Diseases

- Mitochondria Disease. 2009. A compendium of nine articles on mitochondrial diseases. *Biochem. Biophys. Acta Mol. Basis Disease* 1792:1095–1167.
- Cicchetti, F., Drouin-Ouellet, J., and Gross, R. E. 2009. Environmental toxins and Parkinson's disease: What have we learned from pesticide-induced animal models? *Trends Pharm. Sci.* 30:475–483.
- DiMauro, S., and Schon, E. A. 2003. Mitochondrial respiratory-chain disease. New Engl. J. Med. 348:2656–2668.

- Smeitink, J., van den Heuvel, L., and DiMauro, S. 2001. The genetics and pathology of oxidative phosphorylation. *Nat. Rev. Genet.* 2:342–352.
- Wallace, D. C. 1999. Mitochondrial diseases in man and mouse. *Science* 283:1482–1488.

Apoptosis

- Qi, S., Pang, Y., Hu, Q., Liu, Q., Li, H., Zhou, Y., He, T., Liang, Q., Liu, Y., Yuan, X., Luo, G., Li, H., Wang, J., Yan, N., and Shi, Y. 2010.Crystal structure of the *Caenorhabditis elegans* apoptosome reveals an octameric assembly of CED-4. *Cell* 141:446–457.
- Chan, D. C. 2006. Mitochondria: Dynamic organelles in disease, aging, and development. Cell 125:1241–1252.
- Green, D. R. 2005. Apoptotic pathways: Ten minutes to dead. *Cell* 121:671–674.
- Joza, N., Susin, S. A., Daugas, E., Stanford, W. L., Cho, S. K., Li, C. Y. J., Sasaki, T., Elia, A. J., Cheng, H.-Y. M., Ravagnan, L., Ferri, K. F., Zamzami, N., Wakeham, A., Hakem, R., Yoshida, H., Kong, Y.-Y., Mak, T. W., Zúñiga-Pflücker, J. C., Kroemer, G., and Penninger, J. M. 2001. Essential role of the mitochondrial apoptosis-inducing factor in programmed cell death. *Nature* 410:549–554.
- Desagher, S., and Martinou, J. C. 2000. Mitochondria as the central control point of apoptosis. *Trends Cell Biol.* 10:369–377.
- Hengartner, M. O. 2000. The biochemistry of apoptosis. Nature 407:770–776.

Historical Aspects

- Prebble, J., and Weber, B. 2003. Wandering in the Gardens of the Mind: Peter Mitchell and the Making of Glynn. Oxford.
- Mitchell, P. 1979. Keilin's respiratory chain concept and its chemiosmotic consequences. Science 206:1148–1159.
- Preeble, J. 2002. Peter Mitchell and the ox phos wars. *Trends Biochem*. Sci. 27:209–212.
- Mitchell, P. 1976. Vectorial chemistry and the molecular mechanics of chemiosmotic coupling: Power transmission by proticity. *Biochem. Soc. Trans.* 4:399–430.
- Racker, E. 1980. From Pasteur to Mitchell: A hundred years of bioenergetics. Fed. Proc. 39:210–215.
- Kalckar, H. M. 1991. Fifty years of biological research: From oxidative phosphorylation to energy requiring transport and regulation. *Annu. Rev. Biochem.* 60:1–37.

Chapter 19

Where to Start

- Huber, R. 1989. A structural basis of light energy and electron transfer in biology. EMBO J. 8:2125–2147.
- Deisenhofer, J., and Michel, H. 1989. The photosynthetic reaction centre from the purple bacterium *Rhodopseudomonas viridis*. *EMBO J.* 8:2149–2170.
- Barber, J., and Andersson, B. 1994. Revealing the blueprint of photosynthesis. *Nature* 370:31–34.

Books and General Reviews

- Nelson, N., and Yocum, C. 2006. Structure and functions of photosystems I and II. Annu. Rev. Plant Biol. 57:521–565.
- Merchant, S., and Sawaya, M. R. 2005. The light reactions: A guide to recent acquisitions for the picture gallery. *Plant Cell* 17:648–663.
- Blankenship, R. E. 2002. Molecular Mechanisms of Photosynthesis. Wiley-Blackwell.
- Raghavendra, A. S. 2000. Photosynthesis: A Comprehensive Treatise. Cambridge University Press.
- Nicholls, D. G., and Ferguson, S. J. 2002. *Bioenergetics* (3d ed.). Academic Press.

Electron-Transfer Mechanisms

- Beratan, D., and Skourtis, S. 1998. Electron transfer mechanisms. Curr. Opin. Chem. Biol. 2:235–243.
- Moser, C. C., Keske, J. M., Warncke, K., Farid, R. S., and Dutton, P. L. 1992. Nature of biological electron transfer. *Nature* 355: 796–802.
- Boxer, S. G. 1990. Mechanisms of long-distance electron transfer in proteins: Lessons from photosynthetic reaction centers. Annu. Rev. Biophys. Biophys. Chem. 19:267–299.

Photosystem II

- Kirchhoff, H., Tremmel, I., Haase, W., and Kubitscheck, U. 2004. Supramolecular photosystem II organization in grana of thylakoid membranes: Evidence for a structured arrangement. *Biochemistry* 43:9204–9213.
- Diner, B. A., and Rappaport, F. 2002. Structure, dynamics, and energetics of the primary photochemistry of photosystem II of oxygenic photosynthesis. Annu. Rev. Plant Biol. 54:551–580.
- Zouni, A., Witt, H. T., Kern, J., Fromme, P., Krauss, N., Saenger, W., and Orth, P. 2001. Crystal structure of photosystem II from *Synechococcus elongatus* at 3.8 Å resolution. *Nature* 409:739–743.
- Rhee, K. H. 2001. Photosystem II: The solid structural era. Annu. Rev. Biophys. Biomolec. Struct. 30:307–328.
- Deisenhofer, J., and Michel, H. 1991. High-resolution structures of photosynthetic reaction centers. Annu. Rev. Biophys. Biophys. Chem. 20:247–266.

Oxygen Evolution

- Barber, J. 2008. Crystal structure of the oxygen-evolving complex of photosystem II. *Inorg. Chem.* 47:1700–1710.
- Pushkar, Y., Yano, J., Sauer, K., Boussac, A., and Yachandra, V. K. 2008. Structural changes in the Mn₄Ca cluster and the mechanism of photosynthetic water splitting. *Proc. Natl. Acad. Sci. U.S.A.* 105:1879–1884.
- Renger, G. 2007. Oxidative photosynthetic water splitting: Energetics, kinetics and mechanism. *Photosynth. Res.* 92:407–425.
- Renger, G., and Kühn, P. 2007. Reaction pattern and mechanism of light induced oxidative water splitting in photosynthesis. *Biochim. Biophys. Acta* 1767: 458-471.
- Ferreira, K. N., Iverson, T. M., Maghlaoui, K., Barber, J., and Iwata, S. 2004. Architecture of the photosynthetic oxygen-evolving center. *Science* 303:1831–1838.
- Hoganson, C. W., and Babcock, G. T. 1997. A metalloradical mechanism for the generation of oxygen from water in photosynthesis. *Science* 277:1953–1956.
- Yamachandra, V. K., DeRose, V. J., Latimer, M. J., Mukerji, I., Sauer, K., and Klein, M. P. 1993. Where plants make oxygen: A structural model for the photosynthetic oxygen-evolving manganese complex. *Science* 260:675–679.
- Peloquin, J. M., and Britt, R. D. 2001. EPR/ENDOR characterization of the physical and electronic structure of the OEC Mn cluster. *Biochim. Biophys. Acta* 1503:96–111.

Photosystem I and Cytochrome bf

- Iwai, M., Takizawa, K., Tokutsu, R., Okamuro, A., Takahashi, Y., and Minagawa, J. 2010. Isolation of the elusive supercomplex that drives cyclic electron flow in photosynthesis. *Nature* 464:1210–1214.
- Amunts, A., Drory, O., and Nelson, N. 2007. The structure of photosystem I supercomplex at 3.4 Å resolution. *Nature* 447:58–63.
- Cramer, W. A., Zhang, H., Yan, J., Kurisu, G., and Smith, J. L. 2004. Evolution of photosynthesis: Time-independent structure of the cytochrome b₆ f complex. *Biochemistry* 43:5921–5929.
- Kargul, J., Nield, J., and Barber, J. 2003. Three-dimensional reconstruction of a light-harvesting complex I-photosystem I (LHCI-PSI) supercomplex from the green alga *Chlamydomonas reinhardtii*. J. Biol. Chem. 278:16135–16141.

- Schubert, W. D., Klukas, O., Saenger, W., Witt, H. T., Fromme, P., and Krauss, N. 1998. A common ancestor for oxygenic and anoxygenic photosynthetic systems: A comparison based on the structural model of photosystem I. J. Mol. Biol. 280:297–314.
- Fotiadis, D., Muller, D. J., Tsiotis, G., Hasler, L., Tittmann, P., Mini, T., Jeno, P., Gross, H., and Engel, A. 1998. Surface analysis of the photosystem I complex by electron and atomic force microscopy. *J. Mol. Biol.* 283:83–94.
- Klukas, O., Schubert, W. D., Jordan, P., Krauss, N., Fromme, P., Witt, H. T., and Saenger, W. 1999. Photosystem I, an improved model of the stromal subunits PsaC, PsaD, and PsaE. J. Biol. Chem. 274:7351-7360.
- Jensen, P. E., Gilpin, M., Knoetzel, J., and Scheller, H. V. 2000. The PSI-K subunit of photosystem I is involved in the interaction between light-harvesting complex I and the photosystem I reaction center core. J. Biol. Chem. 275:24701–24708.
- Kitmitto, A., Mustafa, A. O., Holzenburg, A., and Ford, R. C. 1998. Three-dimensional structure of higher plant photosystem I determined by electron crystallography. J. Biol. Chem. 273:29592–29599.
- Krauss, N., Hinrichs, W., Witt, I., Fromme, P., Pritzkow, W., Dauter, Z., Betzel, C., Wilson, K. S., Witt, H. T., and Saenger, W. 1993. Three-dimensional structure of system I photosynthesis at 6 Å resolution. *Nature* 361:326–331.
- Malkin, R. 1992. Cytochrome *bc*₁ and *b*₆*f* complexes of photosynthetic membranes. *Photosynth. Res.* 33:121–136.
- Karplus, P. A., Daniels, M. J., and Herriott, J. R. 1991. Atomic structure of ferredoxin-NADP⁺ reductase: Prototype for a structurally novel flavoenzyme family. *Science* 251:60–66.

ATP Synthase

- Vollmar, M., Schlieper, D., Winn, D., Büchner, C., and Groth, G. 2009. Structure of the c14 rotor ring of the proton translocating chloroplast ATP synthase. J. Biol. Chem. 284:18228–18235.
- Varco-Merth, B., Fromme, R., Wang, M., and Fromme, P. 2008. Crystallization of the c14-rotor of the chloroplast ATP synthase reveals that it contains pigments. *Biochim. Biophys. Acta* 1777: 605–612.
- Richter, M. L., Hein, R., and Huchzermeyer, B. 2000. Important subunit interactions in the chloroplast ATP synthase. *Biochim. Biophys. Acta* 1458:326–329.
- Oster, G., and Wang, H. 1999. ATP synthase: Two motors, two fuels. Structure 7:R67–R72.
- Weber, J., and Senior, A. E. 2000. ATP synthase: What we know about ATP hydrolysis and what we do not know about ATP synthesis. *Biochim. Biophys. Acta* 1458:300–309.

Light-Harvesting Assemblies

- Melkozernov, A. N., Barber, J., and Blankenship, R. E. 2006. Light harvesting in photosystem I supercomplexes. *Biochemistry* 45:331–345.
- Conroy, M. J., Westerhuis, W. H., Parkes-Loach, P. S., Loach, P. A., Hunter, C. N., and Williamson, M. P. 2000. The solution structure of *Rhodobacter sphaeroides* LH1β reveals two helical domains separated by a more flexible region: Structural consequences for the LH1 complex. J. Mol. Biol. 298:83–94.
- Koepke, J., Hu, X., Muenke, C., Schulten, K., and Michel, H. 1996. The crystal structure of the light-harvesting complex II (B800–850) from *Rhodospirillum molischianum*. Structure 4:581–597.
- Grossman, A. R., Bhaya, D., Apt, K. E., and Kehoe, D. M. 1995. Light-harvesting complexes in oxygenic photosynthesis: Diversity, control, and evolution. *Annu. Rev. Genet.* 29:231–288.
- Kühlbrandt, W., Wang, D.-N., and Fujiyoshi, Y. 1994. Atomic model of plant light-harvesting complex by electron crystallography. *Nature* 367:614–621.

Evolution

Chen, M., and Zhang, Y. 2008. Tracking the molecular evolution of photosynthesis through characterization of atomic contents of the photosynthetic units. *Photosynth. Res.* 97:255–261.

- Iverson, T. M. 2006. Evolution and unique bioenergetic mechanisms in oxygenic photosynthesis. *Curr. Opin. Chem. Biol.* 10(2):91–100.
- Cavalier-Smith, T. 2002. Chloroplast evolution: Secondary symbiogenesis and multiple losses. *Curr. Biol.* 12:R62–64.
- Nelson, N., and Ben-Shem, A. 2005. The structure of photosystem I and evolution of photosynthesis. *BioEssays* 27:914–922.
- Green, B. R. 2001. Was "molecular opportunism" a factor in the evolution of different photosynthetic light-harvesting pigment systems? *Proc. Natl. Acad. Sci. U.S.A.* 98:2119–2121.
- Dismukes, G. C., Klimov, V. V., Baranov, S. V., Nozlov, Y. N., Das Gupta, J., and Tyryshkin, A. 2001. The origin of atmospheric oxygen on Earth: The innovation of oxygenic photosynthesis. *Proc. Natl. Acad. Sci. U.S.A.* 98:2170–2175.
- Moreira, D., Le Guyader, H., and Phillippe, H. 2000. The origin of red algae and the evolution of chloroplasts. *Nature* 405:69–72.
- Cavalier-Smith, T. 2000. Membrane heredity and early chloroplast evolution. *Trends Plant Sci.* 5:174–182.
- Blankenship, R. E., and Hartman, H. 1998. The origin and evolution of oxygenic photosynthesis. *Trends Biochem. Sci.* 23:94–97.

Chapter 20

Where to Start

- Ellis, R. J. 2010. Tackling unintelligent design. Nature 463: 164-165.
- Gutteridge, S., and Pierce, J. 2006. A unified theory for the basis of the limitations of the primary reaction of photosynthetic CO₂ fixation: Was Dr. Pangloss right? *Proc. Natl. Acad. Sci. U.S.A.* 103: 7203–7204.
- Horecker, B. L. 1976. Unravelling the pentose phosphate pathway. In *Reflections on Biochemistry* (pp. 65–72), edited by A. Kornberg, L. Cornudella, B. L. Horecker, and J. Oro. Pergamon.
- Levi, P. 1984. Carbon. In The Periodic Table. Random House.

Books and General Reviews

- Parry, M. A. J., Andralojc, P. J., Mitchell, R. A. C., Madgwick, P. J., and Keys, A. J. 2003. Manipulation of rubisco: The amount, activity, function and regulation. J. Exp. Bot. 54:1321–1333.
- Spreitzer, R. J., and Salvucci, M. E. 2002. Rubisco: Structure, regulatory interactions, and possibilities for a better enzyme. Annu. Rev. Plant Biol. 53:449–475.
- Wood, T. 1985. The Pentose Phosphate Pathway. Academic Press.
- Buchanan, B. B., Gruissem, W., and Jones, R. L. 2000. Biochemistry and Molecular Biology of Plants. American Society of Plant Physiologists.

Enzymes and Reaction Mechanisms

- Harrison, D. H., Runquist, J. A., Holub, A., and Miziorko, H. M. 1998. The crystal structure of phosphoribulokinase from *Rhodobacter sphaeroides* reveals a fold similar to that of adenylate kinase. *Biochemistry* 37:5074–5085.
- Miziorko, H. M. 2000. Phosphoribulokinase: Current perspectives on the structure/function basis for regulation and catalysis. Adv. Enzymol. Relat. Areas Mol. Biol. 74:95–127.
- Thorell, S., Gergely, P., Jr., Banki, K., Perl, A., and Schneider, G. 2000. The three-dimensional structure of human transaldolase. *FEBS Lett.* 475:205–208.

Carbon Dioxide Fixation and Rubisco

- Satagopan, S., Scott, S. S., Smith, T. G., and Tabita, F. R. 2009. A rubisco mutant that confers growth under a normally "inhibitory" oxygen concentration. *Biochemistry* 48: 9076–9083.
- Tcherkez, G. G. B., Farquhar, G. D., and Andrews, J. T. 2006. Despite slow catalysis and confused substrate specificity, all ribulose bisphosphate carboxylases may be nearly perfectly optimized. *Proc. Natl. Acad. Sci. U.S.A.* 103: 7246–7251.
- Sugawara, H., Yamamoto, H., Shibata, N., Inoue, T., Okada, S., Miyake, C., Yokota, A., and Kai, Y. 1999. Crystal structure of car-

boxylase reaction-oriented ribulose 1,5-bisphosphate carboxylase/ oxygenase from a thermophilic red alga, *Galdieria partita. J. Biol. Chem.* 274:15655–15661.

- Hansen, S., Vollan, V. B., Hough, E., and Andersen, K. 1999. The crystal structure of rubisco from *Alcaligenes eutrophus* reveals a novel central eight-stranded β-barrel formed by β-strands from four subunits. J. Mol. Biol. 288:609–621.
- Knight, S., Andersson, I., and Branden, C. I. 1990. Crystallographic analysis of ribulose 1,5-bisphosphate carboxylase from spinach at 2.4 Å resolution: Subunit interactions and active site. J. Mol. Biol. 215:113–160.
- Taylor, T. C., and Andersson, I. 1997. The structure of the complex between rubisco and its natural substrate ribulose 1,5-bisphosphate. J. Mol. Biol. 265:432–444.
- Cleland, W. W., Andrews, T. J., Gutteridge, S., Hartman, F. C., and Lorimer, G. H. 1998. Mechanism of rubisco: The carbamate as general base. *Chem. Rev.* 98:549–561.
- Buchanan, B. B. 1992. Carbon dioxide assimilation in oxygenic and anoxygenic photosynthesis. *Photosynth. Res.* 33:147–162.
- Hatch, M. D. 1987. C₄ photosynthesis: A unique blend of modified biochemistry, anatomy, and ultrastructure. *Biochim. Biophys. Acta* 895:81–106.

Regulation

- Lebreton, S., Andreescu, S., Graciet, E., and Gontero, B. 2006. Mapping of the interaction site of CP12 with glyceraldehyde-3phosphate dehydrogenase from *Chlamydomonas reinhardtii*. Functional consequences for glyceraldehyde-3-phosphate dehydrogenase. *FEBS J.* 273:3358–3369.
- Graciet, E., Lebreton, S., and Gontero, B. 2004. The emergence of new regulatory mechanisms in the Benson-Calvin pathway via proteinprotein interactions: A glyceraldehyde-3-phosphate dehydrogenase/ CP12/phosphoribulokinase complex. J. Exp. Bot. 55:1245–1254.
- Balmer, Y., Koller, A., del Val, G., Manieri, W., Schürmann, P., and Buchanan, B. B. 2003. Proteomics gives insight into the regulatory function of chloroplast thioredoxins. *Proc. Natl. Acad. Sci. U.S.A.* 100:370–375.
- Rokka, A., Zhang, L., and Aro, E.-M. 2001. Rubisco activase: An enzyme with a temperature-dependent dual function? *Plant J.* 25:463–472.
- Zhang, N., and Portis, A. R., Jr. 1999. Mechanism of light regulation of rubisco: A specific role for the larger rubisco activase isoform involving reductive activation by thioredoxin-f. *Proc. Natl. Acad. Sci. U.S.A.* 96:9438–9443.
- Wedel, N., Soll, J., and Paap, B. K. 1997. CP12 provides a new mode of light regulation of Calvin cycle activity in higher plants. *Proc. Natl. Acad. Sci. U.S.A.* 94:10479–10484.
- Avilan, L., Lebreton, S., and Gontero, B. 2000. Thioredoxin activation of phosphoribulokinase in a bi-enzyme complex from *Chlamydomonas reinhardtii* chloroplasts. J. Biol. Chem. 275:9447–9451.
- Irihimovitch, V., and Shapira, M. 2000. Glutathione redox potential modulated by reactive oxygen species regulates translation of rubisco large subunit in the chloroplast. J. Biol. Chem. 275:16289–16295.

Glucose 6-phosphate Dehydrogenase

- Wang, X.-T., and Engel, P. C. 2009. Clinical mutants of human glucose 6-phosphate dehydrogenase: Impairment of NADP⁺ binding affects both folding and stability. *Biochim. Biophys. Acta* 1792: 804–809.
- Au, S. W., Gover, S., Lam, V. M., and Adams, M. J. 2000. Human glucose-6-phosphate dehydrogenase: The crystal structure reveals a structural NADP(+) molecule and provides insights into enzyme deficiency. *Struct. Fold. Des.* 8:293–303.
- Salvemini, F., Franze, A., Iervolino, A., Filosa, S., Salzano, S., and Ursini, M. V. 1999. Enhanced glutathione levels and oxidoresistance mediated by increased glucose-6-phosphate dehydrogenase expression. J. Biol. Chem. 274:2750–2757.
- Tian, W. N., Braunstein, L. D., Apse, K., Pang, J., Rose, M., Tian, X., and Stanton, R. C. 1999. Importance of glucose-6-phos-

phate dehydrogenase activity in cell death. Am. J. Physiol. 276:C1121-C1131.

- Tian, W. N., Braunstein, L. D., Pang, J., Stuhlmeier, K. M., Xi, Q. C., Tian, X., and Stanton, R. C. 1998. Importance of glucose-6-phosphate dehydrogenase activity for cell growth. J. Biol. Chem. 273:10609–10617.
- Ursini, M. V., Parrella, A., Rosa, G., Salzano, S., and Martini, G. 1997. Enhanced expression of glucose-6-phosphate dehydrogenase in human cells sustaining oxidative stress. *Biochem. J.* 323:801–806.

Evolution

- Deschamps, P., Haferkamp, I., d'Hulst, C., Neuhaus, H. E., and Ball, S. G. 2008. The relocation of starch metabolism to chloroplasts: when, why and how. *Trends Plant Sci.* 13: 574–582.
- Coy, J. F., Dubel, S., Kioschis, P., Thomas, K., Micklem, G., Delius, H., and Poustka, A. 1996. Molecular cloning of tissue-specific transcripts of a transketolase-related gene: Implications for the evolution of new vertebrate genes. *Genomics* 32:309–316.
- Schenk, G., Layfield, R., Candy, J. M., Duggleby, R. G., and Nixon, P. F. 1997. Molecular evolutionary analysis of the thiamine-diphosphatedependent enzyme, transketolase. J. Mol. Evol. 44:552–572.
- Notaro, R., Afolayan, A., and Luzzatto L. 2000. Human mutations in glucose 6-phosphate dehydrogenase reflect evolutionary history. *FASEB J.* 14:485–494.
- Wedel, N., and Soll, J. 1998. Evolutionary conserved light regulation of Calvin cycle activity by NADPH-mediated reversible phosphoribulokinase/CP12/glyceraldehyde-3-phosphate dehydrogenase complex dissociation. Proc. Natl. Acad. Sci. U.S.A. 95:9699–9704.
- Martin, W., and Schnarrenberger, C. 1997. The evolution of the Calvin cycle from prokaryotic to eukaryotic chromosomes: A case study of functional redundancy in ancient pathways through endosymbiosis. *Curr. Genet.* 32:1–18.
- Ku, M. S., Kano-Murakami, Y., and Matsuoka, M. 1996. Evolution and expression of C₄ photosynthesis genes. *Plant Physiol.* 111:949–957.
- Pereto, J. G., Velasco, A. M., Becerra, A., and Lazcano, A. 1999. Comparative biochemistry of CO₂ fixation and the evolution of autotrophy. *Int. Microbiol.* 2:3–10.

Chapter 21

Where to Start

- Krebs, E. G. 1993. Protein phosphorylation and cellular regulation I. Biosci. Rep. 13:127–142.
- Fischer, E. H. 1993. Protein phosphorylation and cellular regulation II. Angew. Chem. Int. Ed. 32:1130–1137.
- Johnson, L. N. 1992. Glycogen phosphorylase: Control by phosphorylation and allosteric effectors. FASEB J. 6:2274–2282.
- Browner, M. F., and Fletterick, R. J. 1992. Phosphorylase: A biological transducer. Trends Biochem. Sci. 17:66–71.

Books and General Reviews

- Agius, L. 2008. Glucokinase and molecular aspects of liver glycogen metabolism. *Biochem. J.* 414:1–18.
- Greenberg, C. C., Jurczak, M. J., Danos, A. M., and Brady, M. J. 2006. Glycogen branches out: New perspectives on the role of glycogen metabolism in the integration of metabolic pathways. Am. J. Physiol. Endocrinol. Metab. 291:E1–E8.
- Shulman, R. G., and Rothman, D. L. 1996. Enzymatic phosphorylation of muscle glycogen synthase: A mechanism for maintenance of metabolic homeostasis. *Proc. Natl. Acad. Sci. U.S.A.* 93:7491–7495.
- Shulman, G. I., and Landau, B. R. 1992. Pathways of glycogen repletion. *Physiol. Rev.* 72:1019–1035.

X-ray Crystallographic Studies

Horcajada, C., Guinovart, J. J., Fita, I., and Ferrer, J. C. 2006. Crystal structure of an archaeal glycogen synthase: Insights into oligomerization and substrate binding of eukaryotic glycogen synthases. *J. Biol. Chem.* 281:2923–2931.

- Buschiazzo, A., Ugalde, J. E., Guerin, M. E., Shepard, W., Ugalde, R. A., and Alzari, P. M. 2004. Crystal structure of glycogen synthase: Homologous enzymes catalyze glycogen synthesis and degradation. *EMBO J.* 23:3196–3205.
- Gibbons, B. J., Roach, P. J., and Hurley, T. D. 2002. Cyrstal structure of the autocatalytic initiator of glycogen biosynthesis, glycogenin. J. Mol. Biol. 319:463–477.
- Sprang, S. R., Withers, S. G., Goldsmith, E. J., Fletterick, R. J., and Madsen, N. B. 1991. Structural basis for the activation of glycogen phosphorylase b by adenosine monophosphate. Science 254:1367–1371.
- Johnson, L. N., and Barford, D. 1990. Glycogen phosphorylase: The structural basis of the allosteric response and comparison with other allosteric proteins. J. Biol. Chem. 265:2409–2412.
- Browner, M. F., Fauman, E. B., and Fletterick, R. J. 1992. Tracking conformational states in allosteric transitions of phosphorylase. *Biochemistry* 31:11297–11304.
- Martin, J. L., Johnson, L. N., and Withers, S. G. 1990. Comparison of the binding of glucose and glucose 1-phosphate derivatives to T-state glycogen phosphorylase b. Biochemistry 29:10745–10757.

Priming of Glycogen Synthesis

- Lomako, J., Lomako, W. M., and Whelan, W. J. 2004. Glycogenin: The primer for mammalian and yeast glycogen synthesis. *Biochim. Biophys. Acta* 1673:45–55.
- Lin, A., Mu, J., Yang, J., and Roach, P. J. 1999. Self-glucosylation of glycogenin, the initiator of glycogen biosynthesis, involves an intersubunit reaction. Arch. Biochem. Biophys. 363:163–170.
- Roach, P. J., and Skurat, A. V. 1997. Self-glucosylating initiator proteins and their role in glycogen biosynthesis. Prog. Nucleic Acid Res. Mol. Biol. 57:289–316.
- Smythe, C., and Cohen, P. 1991. The discovery of glycogenin and the priming mechanism for glycogen biogenesis. *Eur. J. Biochem.* 200:625–631.

Catalytic Mechanisms

- Skamnaki, V. T., Owen, D. J., Noble, M. E., Lowe, E. D., Lowe, G., Oikonomakos, N. G., and Johnson, L. N. 1999. Catalytic mechanism of phosphorylase kinase probed by mutational studies. *Biochemistry* 38:14718–14730.
- Buchbinder, J. L., and Fletterick, R. J. 1996. Role of the active site gate of glycogen phosphorylase in allosteric inhibition and substrate binding. J. Biol. Chem. 271:22305–22309.
- Palm, D., Klein, H. W., Schinzel, R., Buehner, M., and Helmreich, E. J. M. 1990. The role of pyridoxal 5'-phosphate in glycogen phosphorylase catalysis. *Biochemistry* 29:1099–1107.

Regulation of Glycogen Metabolism

- Boulatnikov, I. G., Peters, J. L., Nadeau, O. W., Sage, J. M., Daniels, P. J., Kumar, P., Walsh, D. A., and Carlson, G. M. 2009. Expressed phosphorylase b kinase and its αγδ subcomplex as regulatory models for the rabbit skeletal muscle holoenzyme. *Biochemistry* 48:10183–10191.
- Ros, S., García-Rocha, M., Domínguez, J., Ferrer, J. C., and Guinovart, J. J. 2009. Control of liver glycogen synthase activity and intracellular distribution by phosphorylation. J. Biol. Chem. 284:6370–6378.
- Danos, A. M., Osmanovic, S., and Brady, M. J. 2009. Differential regulation of glycogenolysis by mutant protein phosphatase-1 glycogentargeting subunits. J. Biol. Chem. 284:19544–19553.
- Pautsch, A., Stadler, N., Wissdorf, O., Langkopf, E., Moreth, M., and Streicher, R. 2008. Molecular recognition of the protein phosphatase 1 glycogen targeting subunit by glycogen phosphorylase. J. Biol. Chem. 283:8913–8918.
- Jope, R. S., and Johnson, G. V. W. 2004. The glamour and gloom of glycogen synthase kinase-3. Trends Biochem. Sci. 29:95–102.
- Doble, B. W., and Woodgett, J. R. 2003. GSK-3: Tricks of the trade for a multi-tasking kinase. J. Cell Sci. 116:1175–1186.

- Pederson, B. A., Cheng, C., Wilson, W. A., and Roach, P. J. 2000. Regulation of glycogen synthase: Identification of residues involved in regulation by the allosteric ligand glucose-6-P and by phosphorylation. J. Biol. Chem. 275:27753–27761.
- Melendez, R., Melendez-Hevia, E., and Canela, E. I. 1999. The fractal structure of glycogen: A clever solution to optimize cell metabolism. *Biophys. J.* 77:1327–1332.
- Franch, J., Aslesen, R., and Jensen, J. 1999. Regulation of glycogen synthesis in rat skeletal muscle after glycogen-depleting contractile activity: Effects of adrenaline on glycogen synthesis and activation of glycogen synthase and glycogen phosphorylase. *Biochem. J.* 344(pt.1):231–235.
- Aggen, J. B., Nairn, A. C., and Chamberlin, R. 2000. Regulation of protein phosphatase-1. *Chem. Biol.* 7:R13–R23.
- Egloff, M. P., Johnson, D. F., Moorhead, G., Cohen, P. T., Cohen, P., and Barford, D. 1997. Structural basis for the recognition of regulatory subunits by the catalytic subunit of protein phosphatase 1. *EMBO J.* 16:1876–1887.
- Wu, J., Liu, J., Thompson, I., Oliver, C. J., Shenolikar, S., and Brautigan, D. L. 1998. A conserved domain for glycogen binding in protein phosphatase-1 targeting subunits. *FEBS Lett.* 439:185–191.

Genetic Diseases

- Nyhan, W. L., Barshop, B. A., and Ozand, P. T. 2005. Atlas of Metabolic Diseases. (2d ed., pp. 373–408). Hodder Arnold.
- Chen, Y.-T. 2001. Glycogen storage diseases. In *The Metabolic and Molecular Bases of Inherited Diseases* (8th ed., pp. 1521–1552), edited by C. R. Scriver., W. S. Sly, B. Childs, A. L. Beaudet, D. Valle, K. W. Kinzler, and B. Vogelstein. McGraw-Hill.
- Burchell, A., and Waddell, I. D. 1991. The molecular basis of the hepatic microsomal glucose-6-phosphatase system. *Biochim. Biophys. Acta* 1092:129–137.
- Lei, K. J., Shelley, L. L., Pan, C. J., Sidbury, J. B., and Chou, J. Y. 1993. Mutations in the glucose-6-phosphatase gene that cause glycogen storage disease type Ia. *Science* 262:580–583.
- Ross, B. D., Radda, G. K., Gadian, D. G., Rocker, G., Esiri, M., and Falconer-Smith, J. 1981. Examination of a case of suspected McArdle's syndrome by ³¹P NMR. N. Engl. J. Med. 304:1338–1342.

Evolution

- Holm, L., and Sander, C. 1995. Evolutionary link between glycogen phosphorylase and a DNA modifying enzyme. EMBO J. 14:1287–1293.
- Hudson, J. W., Golding, G. B., and Crerar, M. M. 1993. Evolution of allosteric control in glycogen phosphorylase. J. Mol. Biol. 234:700-721.
- Rath, V. L., and Fletterick, R. J. 1994. Parallel evolution in two homologues of phosphorylase. *Nat. Struct. Biol.* 1:681–690.
- Melendez, R., Melendez-Hevia, E., and Cascante, M. 1997. How did glycogen structure evolve to satisfy the requirement for rapid mobilization of glucose? A problem of physical constraints in structure building. J. Mol. Evol. 45:446–455.
- Rath, V. L., Lin, K., Hwang, P. K., and Fletterick, R. J. 1996. The evolution of an allosteric site in phosphorylase. *Structure* 4:463–473.

Chapter 22

Where to Start

- Rinaldo, P., Matern, D., and Bennet, M. J. 2002. Fatty acid oxidation disorders. Annu. Rev. Physiol. 64:477–502.
- Rasmussen, B. B., and Wolfe, R. R. 1999. Regulation of fatty acid oxidation in skeletal muscle. Annu. Rev. Nutr. 19:463–484.
- Semenkovich, C. F. 1997. Regulation of fatty acid synthase (FAS). Prog. Lipid Res. 36:43–53.
- Sul, H. S., Smas, C. M., Wang, D., and Chen, L. 1998. Regulation of fat synthesis and adipose differentiation. *Prog. Nucleic Acid Res. Mol. Biol.* 60:317–345.

Wolf, G. 1996. Nutritional and hormonal regulation of fatty acid synthase. Nutr. Rev. 54:122–123.

Books

- Vance, D. E., and Vance, J. E. (Eds.). 2008. Biochemistry of Lipids, Lipoproteins, and Membranes. Elsevier.
- Stipanuk, M. H. (Ed.). 2006. Biochemical and Physiological Aspects of Human Nutrition. Saunders.

Fatty Acid Oxidation

- Ahmadian, M., Duncan, R. E., and Sul, H. S. 2009. The skinny on fat: Lipolysis and fatty acid utilization in adipocytes. *Trends Endocrinol. Metab.* 20:424–428.
- Farese, R. V., Jr., and Walther, T. C. 2009. Lipid droplets finally get a little R-E-S-P-E-C-T. Cell 139:855–860.
- Goodman, J. L. 2008. The gregarious lipid droplet. J. Biol. Chem. 283:28005–28009.
- Saha, P. K., Kojima, H., Marinez-Botas, J., Sunehag, A. L., and Chan, L. 2004. Metabolic adaptations in absence of perilipin. J. Biol. Chem. 279:35150–35158.
- Barycki, J. J., O'Brien, L. K., Strauss, A. W., and Banaszak, L. J. 2000. Sequestration of the active site by interdomain shifting: Crystallographic and spectroscopic evidence for distinct conformations of L-3-hydroxyacyl-CoA dehydrogenase. J. Biol. Chem. 275:27186–27196.
- Ramsay, R. R. 2000. The carnitine acyltransferases: Modulators of acyl-CoA-dependent reactions. *Biochem. Soc. Trans.* 28:182–186.
- Eaton, S., Bartlett, K., and Pourfarzam, M. 1996. Mammalian mitochondrial β-oxidation. *Biochem. J.* 320:345–357.
- Thorpe, C., and Kim, J. J. 1995. Structure and mechanism of action of the acyl-CoA dehydrogenases. *FASEB J.* 9:718–725.

Fatty Acid Synthesis

- Maier, T., Leibundgut, M., and Ban, N. 2008. The crystal structure of a mammalian fatty acid synthase. *Science* 321:1315–1322.
- Kuhajda, F. P. 2006. Fatty acid synthase and cancer: New application of an old pathway. *Cancer Res.* 66:5977–5980.
- Ming, D., Kong, Y., Wakil, S. J., Brink, J., and Ma, J. 2002. Domain movements in human fatty acid synthase by quantized elastic deformational model. *Proc. Natl. Acad. Sci. U.S.A.* 99:7895– 7899.
- Zhang, Y.-M., Rao, M. S., Heath, R. J., Price, A. C., Olson, A. J., Rock, C. O., and White, S. W. 2001. Identification and analysis of the acyl carrier protein (ACP) docking site on β-ketoacyl-ACP synthase III. J. Biol. Chem. 276:8231–8238.
- Davies, C., Heath, R. J., White, S. W., and Rock, C. O. 2000. The 1.8 Å crystal structure and active-site architecture of β-ketoacyl-acyl carrier protein synthase III (FabH) from *Escherichia coli*. Struct. Fold. Design 8:185–195.
- Denton, R. M., Heesom, K. J., Moule, S. K., Edgell, N. J., and Burnett, P. 1997. Signalling pathways involved in the stimulation of fatty acid synthesis by insulin. *Biochem. Soc. Trans.* 25:1238–1242.
- Loftus, T. M., Jaworsky, D. E., Frehywot, G. L., Townsend, C. A., Ronnett, G. V., Lane, M. D., and Kuhajda, F. P. 2000. Reduced food intake and body weight in mice treated with fatty acid synthase inhibitors. *Science* 288:2379–2381.

Acetyl CoA Carboxylase

- Brownsey, R. W., Boone, A. N., Elliott, J. E., Kulpa, J. E., and Lee, W. M. 2006. Regulation of acetyl-CoA carboxylase. *Biochem. Soc. Trans.* 34:223–227.
- Munday, M. R. 2002. Regulation of acetyl CoA carboxylase. Biochem. Soc. Trans. 30: 1059–1064.
- Thoden, J. B., Blanchard, C. Z., Holden, H. M., and Waldrop, G. L. 2000. Movement of the biotin carboxylase B-domain as a result of ATP binding. J. Biol. Chem. 275:16183–16190.

Eicosanoids

- Harizi, H., Corcuff, J.-B., and Gualde, N. 2008. Arachidonic-acidderived eicosanoids: Roles in biology and immunopathology. *Trends Mol. Med.* 14:461–469.
- Nakamura, M. T., and Nara, T. Y. 2004. Structure, function, and dietary regulation of $\Delta 6$, $\Delta 5$, and $\Delta 9$ desaturases. *Annu. Rev. Nutr.* 24:345–376.
- Malkowski, M. G., Ginell, S. L., Smith, W. L., and Garavito, R. M. 2000. The productive conformation of arachidonic acid bound to prostaglandin synthase. *Science* 289:1933–1937.
- Smith, T., McCracken, J., Shin, Y.-K., and DeWitt, D. 2000. Arachidonic acid and nonsteroidal anti-inflammatory drugs induce conformational changes in the human prostaglandin endoperoxide H₂ synthase-2 (cyclooxygenase-2). J. Biol. Chem. 275:40407–40415.
- Kalgutkar, A. S., Crews, B. C., Rowlinson, S. W., Garner, C., Seibert, K., and Marnett L. J. 1998. Aspirin-like molecules that covalently inactivate cyclooxygenase-2. *Science* 280:1268–1270.
- Lands, W. E. 1991. Biosynthesis of prostaglandins. Annu. Rev. Nutr. 11:41–60.
- Sigal, E. 1991. The molecular biology of mammalian arachidonic acid metabolism. Am. J. Physiol. 260:L13–L28.
- Weissmann, G. 1991. Aspirin. Sci. Am. 264(1):84-90.
- Vane, J. R., Flower, R. J., and Botting, R. M. 1990. History of aspirin and its mechanism of action. *Stroke* (12 suppl.):IV12–IV23.

Genetic Diseases

- Nyhan, W. L., Barshop, B. A., and Ozand, P. T. 2005. Atlas of Metabolic Diseases (2d ed., pp. 339–300). Hodder Arnold.
- Roe, C. R., and Coates, P. M. 2001. Mitochondrial fatty acid oxidation disorders. In *The Metabolic and Molecular Bases of Inherited Diseases* (8th ed., pp. 2297–2326), edited by C. R. Scriver., W. S. Sly, B. Childs, A. L. Beaudet, D. Valle, K. W. Kinzler, and B. Vogelstein. McGraw-Hill.
- Brivet, M., Boutron, A., Slama, A., Costa, C., Thuillier, L., Demaugre, F., Rabier, D., Saudubray, J. M., and Bonnefont, J. P. 1999. Defects in activation and transport of fatty acids. *J. Inherit. Metab. Dis.* 22:428–441.
- Wanders, R. J., van Grunsven, E. G., and Jansen, G. A. 2000. Lipid metabolism in peroxisomes: Enzymology, functions and dysfunctions of the fatty acid α-and β-oxidation systems in humans. *Biochem. Soc. Trans.* 28:141–149.
- Wanders, R. J., Vreken, P., den Boer, M. E., Wijburg, F. A., van Gennip, A. H., and Ijist, L. 1999. Disorders of mitochondrial fatty acyl-CoA β-oxidation. J. Inherit. Metab. Dis. 22:442–487.
- Kerner, J., and Hoppel, C. 1998. Genetic disorders of carnitine metabolism and their nutritional management. Annu. Rev. Nutr. 18:179–206.
- Bartlett, K., and Pourfarzam, M. 1998. Recent developments in the detection of inherited disorders of mitochondrial β-oxidation. *Biochem. Soc. Trans.* 26:145–152.
- Pollitt, R. J. 1995. Disorders of mitochondrial long-chain fatty acid oxidation. J. Inherit. Metab. Dis. 18:473–490.

Chapter 23

Where to Start

- Ubiquitin-Mediated Protein Regulation. 2009. Annu. Rev. Biochem. 78: A series of reviews on the various roles of ubiquitin.
- Torchinsky, Y. M. 1989. Transamination: Its discovery, biological and chemical aspects. Trends Biochem. Sci. 12:115–117.
- Eisensmith, R. C., and Woo, S. L. C. 1991. Phenylketonuria and the phenylalanine hydroxylase gene. *Mol. Biol. Med.* 8:3–18.
- Schwartz, A. L., and Ciechanover, A. 1999. The ubiquitin-proteasome pathway and pathogenesis of human diseases. *Annu. Rev. Med.* 50:57–74.
- Watford, M. 2003. The urea cycle. Biochem. Mol. Biol. Ed. 31:289-297.

SELECTED READINGS

Books

- Bender, D. A. 1985. Amino Acid Metabolism (2d ed.). Wiley.
- Lippard, S. J., and Berg, J. M. 1994. Principles of Bioinorganic Chemistry. University Science Books.
- Walsh, C. 1979. Enzymatic Reaction Mechanisms. W. H. Freeman and Company.
- Christen, P., and Metzler, D. E. 1985. Transaminases. Wiley.

Ubiquitin and the Proteasome

- Greer, P. L., Hanayama, R., Bloodgood, B. L., Mardinly, A. R., Lipton, D. M., Flavell, S.W., Kim, T.-K., Griffith, E. C., Waldon, Z., Maehr, R., Ploegh, H. L., Chowdhury, S., Worley, P. F., Steen, J., and Greenberg, M. E. 2010. The Angelman syndrome protein Ube3A regulates synapse development by ubiquitinating Arc. *Cell* 140: 704–716.
- Peth, A., Besche, H. C., and Goldberg A. L. 2009. Ubiquitinated proteins activate the proteasome by binding to Usp14/Ubp6, which causes 20S gate opening. *Mol. Cell* 36: 794–804.
- Cheng, Y. 2009. Toward an atomic model of the 26S proteasome. Curr. Opin. Struct. Biol. 19:203–208.
- Lin, G., Li, D., Carvalho, L. P. S., Deng, H., Tao, H., Vogt, G., Wu, K., Schneider, J., Chidawanyika, T., Warren, J. D., Li, H., and Nathan, C. 2009. Inhibitors selective for mycobacterial versus human proteasomes. *Nature* 461: 621–626.
- Wang, K. H., Roman-Hernandez, G., Grant, R. A., Sauer, R. T., and Baker, T. A. 2008. The molecular basis of N-end rule recognition. *Mol. Cell* 32: 406–414.
- da Fonseca, P. C. A., and Morris, E. P. 2008. Structure of the human 26S proteasome: Subunit radial displacements open the gate into the proteolytic core. *J. Biol. Chem.* 283: 23305–23314.
- Cooper, E. M., Hudson, A. W., Amos, J., Wagstaff, J., and Howley, P. M. 2004. Biochemical analysis of Angelman syndrome-associated mutation in the E3 ubiquitin ligase E6-associated protein. J. Biol. Chem. 279:41208–41217.
- Giasson, B. I. and Lee, V. M.-Y. 2003. Are ubiquitination pathways central to Parkinson's disease? *Cell* 114:1–8.
- Pagano, M., and Benmaamar, R. 2003. When protein destruction runs amok, malignancy is on the loose. *Cancer Cell* 4:251–256.
- Thrower, J. S., Hoffman, L., Rechsteiner, M., and Pickart, C. M. 2000. Recognition of the polyubiquitin proteolytic signal. *EMBO J.* 19:94–102.
- Hochstrasser, M. 2000. Evolution and function of ubiquitin-like protein-conjugation systems. Nat. Cell Biol. 2:E153–E157.

Pyridoxal Phosphate-Dependent Enzymes

- Eliot, A. C., and Kirsch, J. F. 2004. Pyridoxal phosphate enzymes: Mechanistic, structural, and evolutionary considerations. *Annu. Rev. Biochem.* 73:383–415.
- Mehta, P. K., and Christen, P. 2000. The molecular evolution of pyridoxal-5'-phosphate-dependent enzymes. Adv. Enzymol. Relat. Areas Mol. Biol. 74:129–184.
- Schneider, G., Kack, H., and Lindqvist, Y. 2000. The manifold of vitamin B₆ dependent enzymes. Structure Fold Des. 8:R1–R6.

Urea Cycle Enzymes

- Morris, S. M., Jr. 2002. Regulation of enzymes of the urea cycle and arginine metabolism. *Annu. Rev. Nutr.* 22:87–105.
- Huang, X., and Raushel, F. M. 2000. Restricted passage of reaction intermediates through the ammonia tunnel of carbamoyl phosphate synthetase. J. Biol. Chem. 275:26233–26240.
- Lawson, F. S., Charlebois, R. L., and Dillon, J. A. 1996. Phylogenetic analysis of carbamoylphosphate synthetase genes: Complex evolutionary history includes an internal duplication within a gene which can root the tree of life. *Mol. Biol. Evol.* 13:970–977.
- McCudden, C. R., and Powers-Lee, S. G. 1996. Required allosteric effector site for N-acetylglutamate on carbamoyl-phosphate synthetase I. J. Biol. Chem. 271:18285–18294.

Amino Acid Degradation

- Li, M., Smith, C. J., Walker, M. T., and Smith, T. J. 2009. Novel inhibitors complexed with glutamate dehydrogenase: allosteric regulation by control of protein dynamics. J. Biol. Chem. 284:22988–23000.
- Smith, T. J., and Stanley, C. A. 2008. Untangling the glutamate dehydrogenase allosteric nightmare. *Trends Biochem. Sci.* 33: 557–564.
- Fusetti, F., Erlandsen, H., Flatmark, T., and Stevens, R. C. 1998. Structure of tetrameric human phenylalanine hydroxylase and its implications for phenylketonuria. J. Biol. Chem. 273:16962–16967.
- Titus, G. P., Mueller, H. A., Burgner, J., Rodriguez De Cordoba, S., Penalva, M. A., and Timm, D. E. 2000. Crystal structure of human homogentisate dioxygenase. *Nat. Struct. Biol.* 7:542–546.
- Erlandsen, H., and Stevens, R. C. 1999. The structural basis of phenylketonuria. *Mol. Genet. Metab.* 68:103–125.

Genetic Diseases

- Jayakumar, A. R., Liu, M., Moriyama, M. Ramakrishnan, R., Forbush III, B., Reddy, P. V. V., and Norenberg, M.D. 2008. Na-K-Cl cotransporter-1 in the mechanism of ammonia-induced astrocyte swelling. J. Biol. Chem. 283: 33874–33882.
- Scriver, C. R., and Sly, W. S. (Eds.), Childs, B., Beaudet, A. L., Valle, D., Kinzler, K. W., and Vogelstein, B. 2001. *The Metabolic Basis of Inherited Disease* (8th ed.). McGraw-Hill.

Historical Aspects and the Process of Discovery

- Cooper, A. J. L., and Meister, A. 1989. An appreciation of Professor Alexander E. Braunstein: The discovery and scope of enzymatic transamination. *Biochimie* 71:387–404.
- Garrod, A. E. 1909. *Inborn Errors in Metabolism*. Oxford University Press (reprinted in 1963 with a supplement by H. Harris).
- Childs, B. 1970. Sir Archibald Garrod's conception of chemical individuality: A modern appreciation. N. Engl. J. Med. 282:71–78.
- Holmes, F. L. 1980. Hans Krebs and the discovery of the ornithine cycle. *Fed. Proc.* 39:216–225.

Chapter 24

Where to Start

- Kim, J., and Rees, D. C. 1989. Nitrogenase and biological nitrogen fixation. *Biochemistry* 33:389–397.
- Christen, P., Jaussi, R., Juretic, N., Mehta, P. K., Hale, T. I., and Ziak, M. 1990. Evolutionary and biosynthetic aspects of aspartate aminotransferase isoenzymes and other aminotransferases. Ann. N. Y. Acad. Sci. 585:331–338.
- Schneider, G., Kack, H., and Lindqvist, Y. 2000. The manifold of vitamin B6 dependent enzymes. *Structure Fold Des.* 8:R1–R6.
- Rhee, S. G., Chock, P. B., and Stadtman, E. R. 1989. Regulation of Escherichia coli glutamine synthetase. Adv. Enzymol. Mol. Biol. 62:37–92.
- Shemin, D. 1989. An illustration of the use of isotopes: The biosynthesis of porphyrins. *Bioessays* 10:30–35.

Books

- Bender, D. A. 1985. Amino Acid Metabolism (2d ed.). Wiley.
- Jordan, P. M. (Ed.). 1991. Biosynthesis of Tetrapyrroles. Elsevier.
- Scriver, C. R. (Ed.), Sly, W. S. (Ed.), Childs, B., Beaudet, A. L., Valle, D., Kinzler, K. W., and Vogelstein, B. 2001. *The Metabolic Basis of Inherited Disease* (8th ed.). McGraw-Hill.
- Meister, A. 1965. *Biochemistry of the Amino Acids* (vols. 1 and 2, 2d ed.). Academic Press.
- McMurry, J. E., and Begley, T. P. 2005. The Organic Chemistry of Biological Pathways. Roberts and Company.
- Blakley, R. L., and Benkovic, S. J. 1989. Folates and Pterins (vol. 2). Wiley.
- Walsh, C. 1979. Enzymatic Reaction Mechanisms. W. H. Freeman and Company.

Selected Readings

Nitrogen Fixation

- Seefeldt, L. C., Hoffman, B. M., and Dean, D. R. 2009. Mechanism of Mo-dependent nitrogenase. Annu. Rev. Biochem. 79:701–722.
- Halbleib, C. M., and Ludden, P. W. 2000. Regulation of biological nitrogen fixation. J. Nutr. 130:1081–1084.
- Einsle, O., Tezcan, F. A., Andrade, S. L., Schmid, B., Yoshida, M., Howard, J. B., and Rees, D. C. 2002. Nitrogenase MoFe-protein at 1.16 Å resolution: A central ligand in the FeMo-cofactor. *Science* 297:1696–1700.
- Benton, P. M., Laryukhin, M., Mayer, S. M., Hoffman, B. M., Dean, D. R., and Seefeldt, L. C. 2003. Localization of a substrate binding site on the FeMo-cofactor in nitrogenase: Trapping propargyl alcohol with an α-70-substituted MoFe protein. *Biochemistry* 42: 9102–9109.
- Peters, J. W., Fisher, K., and Dean, D. R. 1995. Nitrogenase structure and function: A biochemical-genetic perspective. Annu. Rev. Microbiol. 49:335–366.
- Leigh, G. J. 1995. The mechanism of dinitrogen reduction by molybdenum nitrogenases. Eur. J. Biochem. 229:14–20.
- Georgiadis, M. M., Komiya, H., Chakrabarti, P., Woo, D., Kornuc, J. J., and Rees, D. C. 1992. Crystallographic structure of the nitrogenase iron protein from Azotobacter vinelandii. Science 257:1653–1659.

Regulation of Amino Acid Biosynthesis

- Eisenberg, D., Gill, H. S., Pfluegl, G. M., and Rotstein, S. H. 2000. Structure-function relationships of glutamine synthetases. *Biochim. Biophys. Acta* 1477:122–145.
- Purich, D. L. 1998. Advances in the enzymology of glutamine synthesis. Adv. Enzymol. Relat. Areas Mol. Biol. 72:9–42.
- Yamashita, M. M., Almassy, R. J., Janson, C. A., Cascio, D., and Eisenberg, D. 1989. Refined atomic model of glutamine synthetase at 3.5 Å resolution. J. Biol. Chem. 264:17681–17690.
- Schuller, D. J., Grant, G. A., and Banaszak, L. J. 1995. The allosteric ligand site in the V_{max}-type cooperative enzyme phosphoglycerate dehydrogenase. *Nat. Struct. Biol.* 2:69–76.
- Rhee, S. G., Park, R., Chock, P. B., and Stadtman, E. R. 1978. Allosteric regulation of monocyclic interconvertible enzyme cascade systems: Use of *Escherichia coli* glutamine synthetase as an experimental model. *Proc. Natl. Acad. Sci. U.S.A.* 75:3138–3142.
- Wessel, P. M., Graciet, E., Douce, R., and Dumas, R. 2000. Evidence for two distinct effector-binding sites in threonine deaminase by site-directed mutagenesis, kinetic, and binding experiments. *Biochemistry* 39:15136–15143.
- James, C. L., and Viola, R. E. 2002. Production and characterization of bifunctional enzymes: Domain swapping to produce new bifunctional enzymes in the aspartate pathway. *Biochemistry* 41: 3720–3725.
- Xu, Y., Carr, P. D., Huber, T., Vasudevan, S. G., and Ollis, D. L. 2001. The structure of the P_{II}-ATP complex. *Eur. J. Biochem.* 268: 2028–2037.
- Krappmann, S., Lipscomb, W. N., and Braus, G. H. 2000. Coevolution of transcriptional and allosteric regulation at the chorismate metabolic branch point of *Saccharomyces cerevisiae*. Proc. Natl. Acad. Sci. U.S.A. 97:13585–13590.

Aromatic Amino Acid Biosynthesis

- Brown, K. A., Carpenter, E. P., Watson, K. A., Coggins, J. R., Hawkins, A. R., Koch, M. H., and Svergun, D. I. 2003. Twists and turns: A tale of two shikimate-pathway enzymes. *Biochem. Soc. Trans.* 31:543–547.
- Pan, P., Woehl, E., and Dunn, M. F. 1997. Protein architecture, dynamics and allostery in tryptophan synthase channeling. *Trends Biochem. Sci.* 22:22–27.
- Sachpatzidis, A., Dealwis, C., Lubetsky, J. B., Liang, P. H., Anderson, K. S., and Lolis, E. 1999. Crystallographic studies of phosphonatebased α-reaction transition-state analogues complexed to tryptophan synthase. *Biochemistry* 38:12665–12674.
- Weyand, M., and Schlichting, I. 1999. Crystal structure of wild-type tryptophan synthase complexed with the natural substrate indole-3-glycerol phosphate. *Biochemistry* 38:16469–16480.

- Crawford, I. P. 1989. Evolution of a biosynthetic pathway: The tryptophan paradigm. *Annu. Rev. Microbiol.* 43:567–600.
- Carpenter, E. P., Hawkins, A. R., Frost, J. W., and Brown, K. A. 1998. Structure of dehydroquinate synthase reveals an active site capable of multistep catalysis. *Nature* 394:299–302.
- Schlichting, I., Yang, X. J., Miles, E. W., Kim, A. Y., and Anderson, K. S. 1994. Structural and kinetic analysis of a channel-impaired mutant of tryptophan synthase. J. Biol. Chem. 269:26591–26593.

Glutathione

- Edwards, R., Dixon, D. P., and Walbot, V. 2000. Plant glutathione S-transferases: Enzymes with multiple functions in sickness and in health. *Trends Plant Sci.* 5:193–198.
- Lu, S. C. 2000. Regulation of glutathione synthesis. Curr. Top. Cell Regul. 36:95–116.
- Schulz, J. B., Lindenau, J., Seyfried, J., and Dichgans, J. 2000. Glutathione, oxidative stress and neurodegeneration. *Eur. J. Biochem.* 267:4904–4911.
- Lu, S. C. 1999. Regulation of hepatic glutathione synthesis: Current concepts and controversies. FASEB J. 13:1169–1183.
- Salinas, A. E., and Wong, M. G. 1991. Glutathione S-transferases: A review. Curr. Med. Chem. 6:279–309.

Ethylene and Nitric Oxide

- Nisoli, E., Falcone, S., Tonello, C., Cozzi, V., Palomba, L., Fiorani, M., Pisconti, A., Brunelli, S., Cardile, A., Francolini, M., Cantoni, O., Carruba, M. O., Moncada, S., and Clementi, E. 2004. Mitochondrial biogenesis by NO yields functionally active mitochondria in mammals. *Proc. Natl. Acad. Sci U.S.A.* 101:16507–16512.
- Bretscher, L. E., Li, H., Poulos, T. L. and Griffith, O. W. 2003. Structural characterization and kinetics of nitric oxide synthase inhibition by novel N^5 -(iminoalkyl)- and N^5 -(iminoalkenyl)ornithines. J. Biol. Chem. 278:46789–46797.
- Haendeler, J., Zeiher, A. M., and Dimmeler, S. 1999. Nitric oxide and apoptosis. Vitam. Horm. 57:49–77.
- Capitani, G., Hohenester, E., Feng, L., Storici, P., Kirsch, J. F., and Jansonius, J. N. 1999. Structure of 1-aminocyclopropane-1carboxylate synthase, a key enzyme in the biosynthesis of the plant hormone ethylene. J. Mol. Biol. 294:745–756.
- Hobbs, A. J., Higgs, A., and Moncada, S. 1999. Inhibition of nitric oxide synthase as a potential therapeutic target. Annu. Rev. Pharmacol. Toxicol. 39:191–220.
- Stuehr, D. J. 1999. Mammalian nitric oxide synthases. Biochim. Biophys. Acta 1411:217–230.
- Chang, C., and Shockey, J. A. 1999. The ethylene-response pathway: Signal perception to gene regulation. *Curr. Opin. Plant Biol.* 2:352–358.
- Theologis, A. 1992. One rotten apple spoils the whole bushel: The role of ethylene in fruit ripening. *Cell* 70:181–184.

Biosynthesis of Porphyrins

- Kaasik, K. and Lee, C. C. 2004. Reciprocal regulation of haem biosynthesis and the circadian clock in mammals. *Nature* 430:467–471.
- Leeper, F. J. 1989. The biosynthesis of porphyrins, chlorophylls, and vitamin B₁₂. Nat. Prod. Rep. 6:171–199.
- Porra, R. J., and Meisch, H.-U. 1984. The biosynthesis of chlorophyll. Trends Biochem. Sci. 9:99–104.

Chapter 25

Where to Start

- Sutherland, J. D. 2010. Ribonucleotides. Cold Spring Harb. Perspect. Biol. 2:a005439.
- Ordi, J., Alonso, P. L., de Zulueta, J., Esteban, J., Velasco, M., Mas, E., Campo, E., and Fernández, P. L. 2006. The severe gout of Holy Roman Emperor Charles V. N. Eng. J. Med. 355: 516–520.

- Kappock, T. I., Ealick, S. E., and Stubbe, J. 2000. Modular evolution of the purine biosynthetic pathway. Curr. Opin. Chem. Biol. 4:567-572.
- Jordan, A., and Reichard, P. 1998. Ribonucleotide reductases. Annu. Rev. Biochem. 67:71-98.

Pyrimidine Biosynthesis

- Raushel, F. M., Thoden, J. B., Reinhart, G. D., and Holden, H. M. 1998. Carbamoyl phosphate synthetase: A crooked path from substrates to products. Curr. Opin. Chem. Biol. 2:624-632.
- Huang, X., Holden, H. M., and Raushel, F. M. 2001. Channeling of substrates and intermediates in enzyme-catalyzed reactions. Annu. Rev. Biochem. 70:149-180.
- Begley, T. P., Appleby, T. C., and Ealick, S. E. 2000. The structural basis for the remarkable proficiency of orotidine 5'-monophosphate decarboxylase. Curr. Opin. Struct. Biol. 10:711-718.
- Traut, T. W., and Temple, B. R. 2000. The chemistry of the reaction determines the invariant amino acids during the evolution and divergence of orotidine 5'-monophosphate decarboxylase. J. Biol. Chem. 275:28675-28681.

Purine Biosynthesis

- An, S., Kyoung, M., Allen, J. J., Shokat, K. M., and Benkovic, S. J. 2010. Dynamic regulation of a metabolic multi-enzyme complex by protein kinase CK2. J. Biol. Chem. 285: 11093-11099.
- An, S., Kumar, R., Sheets, E. D., and Benkovic, S. J. 2008. Reversible compartmentalization of de novo purine biosynthetic complexes in living cells. Science 320: 103-106.
- Thoden, J. B., Firestine, S., Nixon, A., Benkovic, S. J., and Holden, H. M. 2000. Molecular structure of Escherichia coli PurT-encoded glycinamide ribonucleotide transformylase. Biochemistry 39:8791-8802.
- McMillan, F. M., Cahoon, M., White, A., Hedstrom, L., Petsko, G. A., and Ringe, D. 2000. Crystal structure at 2.4 Å resolution of Borrelia burgdorferi inosine 5'-monophosphate dehydrogenase: Evidence of a substrate-induced hinged-lid motion by loop 6. Biochemistry 39:4533-4542.
- Levdikov, V. M., Barynin, V. V., Grebenko, A. I., Melik-Adamyan, W. R., Lamzin, V. S., and Wilson, K. S. 1998. The structure of SAICAR synthase: An enzyme in the de novo pathway of purine nucleotide biosynthesis. Structure 6:363-376.
- Smith, J. L., Zaluzec, E. J., Wery, J. P., Niu, L., Switzer, R. L., Zalkin, H., and Satow, Y. 1994. Structure of the allosteric regulatory enzyme of purine biosynthesis. Science 264:1427-1433.
- Weber, G., Nagai, M., Natsumeda, Y., Ichikawa, S., Nakamura, H., Eble, J. N., Jayaram, H. N., Zhen, W. N., Paulik, E., and Hoffman, R. 1991. Regulation of de novo and salvage pathways in chemotherapy. Adv. Enzyme Regul. 31:45-67.

Ribonucleotide Reductases

- Cotruvo, Jr., J. A., and Stubbe, J. 2010. An active dimanganese(III)tyrosyl radical cofactor in Escherichia coli class Ib ribonucleotide reductase. Biochemistry 49:1297-1309.
- Avval, F. Z., and Holmgren, A. 2009. Molecular mechanisms of thioredoxin and glutaredoxin as hydrogen donors for mammalian S phase ribonucleotide reductase. J. Biol. Chem. 284: 8233-8240.
- Rofougaran, R., Crona M., Vodnala, M., Sjöberg, B. M., and Hofer, A. 2008. Oligomerization status directs overall activity regulation of the Escherichia coli class Ia ribonucleotide reductase. J. Biol. Chem. 283: 35310-35318.
- Nordlund, P., and Reichard, P. 2006. Ribonucleotide reductases. Annu. Rev. Biochem. 75: 681-706.
- Eklund, H., Uhlin, U., Farnegardh, M., Logan, D. T. and Nordlund, P. 2001. Structure and function of the radical enzyme ribonucleotide reductase. Prog. Biophys. Mol. Biol. 77:177-268.
- Reichard, P. 1997. The evolution of ribonucleotide reduction. Trends Biochem. Sci. 22:81-85.
- Stubbe, J. 2000. Ribonucleotide reductases: The link between an RNA and a DNA world? Curr. Opin. Struct. Biol. 10:731-736.

- Logan, D. T., Andersson, I., Sjoberg, B. M., and Nordlund, P. 1999, A glycyl radical site in the crystal structure of a class III ribonucleotide reductase. Science 283:1499-1504.
- Tauer, A., and Benner, S. A. 1997. The B₁₂-dependent ribonucleotide reductase from the archaebacterium Thermoplasma acidophila: An evolutionary solution to the ribonucleotide reductase conundrum. Proc. Natl. Acad. Sci. U.S.A. 94:53-58.
- Stubbe, J., Nocera, D. G., Yee, C. S. and Chang, M. C. 2003. Radical initiation in the class I ribonucleotide reductase: Long-range proton-coupled electron transfer? Chem. Rev. 103:2167-2201.
- Stubbe, J., and Riggs-Gelasco, P. 1998. Harnessing free radicals: Formation and function of the tyrosyl radical in ribonucleotide reductase. Trends Biochem. Sci. 23:438-443.

Thymidylate Synthase and Dihydrofolate Reductase

- Abali, E. E., Skacel, N. E., Celikkaya, H., and Hsieh, Y.-C. 2008. Regulation of human dihydrofolate reductase activity and expression. Vitam. Horm. 79:267-292.
- Schnell, J. R., Dyson, H. J., and Wright, P. E. 2004. Structure, dynamics, and catalytic function of dihydrofolate reductase. Annu. Rev. Biophys. Biomol. Struct. 33:119-140.
- Li, R., Sirawaraporn, R., Chitnumsub, P., Sirawaraporn, W., Wooden, J., Athappilly, F., Turley, S., and Hol, W. G. 2000. Threedimensional structure of M. tuberculosis dihydrofolate reductase reveals opportunities for the design of novel tuberculosis drugs. I. Mol. Biol. 295:307-323.
- Liang, P. H., and Anderson, K. S. 1998. Substrate channeling and domain-domain interactions in bifunctional thymidylate synthasedihydrofolate reductase. Biochemistry 37:12195-12205.
- Miller, G. P., and Benkovic, S. J. 1998. Stretching exercises: Flexibility in dihydrofolate reductase catalysis. Chem. Biol. 5:R105-R113.
- Carreras, C. W., and Santi, D. V. 1995. The catalytic mechanism and structure of thymidylate synthase. Annu. Rev. Biochem. 64:721-762.
- Schweitzer, B. I., Dicker, A. P., and Bertino, J. R. 1990. Dihydrofolate reductase as a therapeutic target. FASEB J. 4:2441-2452.

Defects in Nucleotide Biosynthesis

- Aiuti, A., Cattaneo, F., Galimberti, S., Benninghoff, U., et al. 2009. Gene therapy for immunodeficiency due to adenosine deaminase deficiency. N. Engl. J. Med. 360:447-58.
- Jurecka, A. 2009. Inborn errors of purine and pyrimidine metabolism. J. Inherit. Metab. Dis. 32:247-263.
- Nyhan, W. L., Barshop, B. A., and Ozand, P. T. 2005. Atlas of Metabolic Diseases. (2d ed., pp. 429-462). Hodder Arnold.
- Scriver, C. R., Sly, W. S., Childs, B., Beaudet, A. L., Valle, D., Kinzler, K. W., and Vogelstein, B. (Eds.). 2001. The Metabolic and Molecular Bases of Inherited Diseases (8th ed., pp. 2513-2704). McGraw-Hill.
- Nyhan, W. L. 1997. The recognition of Lesch-Nyhan syndrome as an inborn error of purine metabolism. J. Inherited Metab. Dis. 20: 171-178.
- Wong, D. F., Harris, J. C., Naidu, S., Yokoi, F., Marenco, S., Dannals, R. F., Ravert, H. T., Yaster, M., Evans, A., Rousset, O., Bryan, R. N., Gjedde, A., Kuhar, M. J., and Breese, G. R. 1996. Dopamine transporters are markedly reduced in Lesch-Nyhan disease in vivo. Proc. Natl. Acad. Sci. U.S.A. 93:5539-5543.
- Resta, R., and Thompson, L. F. 1997. SCID: The role of adenosine deaminase deficiency. Immunol. Today 18:371-374.
- Neychev, V. K., and Mitev, V. I. 2004. The biochemical basis of the neurobehavioral abnormalities in the Lesch-Nyhan syndrome: A hypothesis. Med. Hypotheses 63:131-134.

Chapter 26 Where to Start

Brown, M. S., and Goldstein, J. L. 2009. Cholesterol feedback: From Schoenheimer's bottle to Scap's MELADL. J. Lipid Res. 50:S15-S27.

- Gimpl, G., Burger, K., and Fahrenholz, F. 2002. A closer look at the cholesterol sensor. *Trends Biochem. Sci.* 27:595–599.
- Oram, J. F. 2002. Molecular basis of cholesterol homeostasis: Lessons from Tangier disease and ABCA1. Trends Mol. Med. 8:168–173.
- Vance, D. E., and Van den Bosch, H. 2000. Cholesterol in the year 2000. Biochim. Biophys. Acta 1529:1–8.
- Endo, A. 1992. The discovery and development of HMG-CoA reductase inhibitors. *J. Lipid Res.* 33:1569–1582.

Books

- Vance, J. E., and Vance, D. E. (Eds.). 2008. Biochemistry of Lipids, Lipoproteins and Membranes. Elsevier.
- Nyhan, W. L., Barshop, B. A., and Ozand, P. T. 2005. Atlas of Metabolic Diseases. (2d ed., pp. 567–696). Hodder Arnold.
- Scriver, C. R., Sly, W. S., Childs, B., Beaudet, A. L., Valle, D., Kinzler, K. W., and Vogelstein, B. (Eds.). 2001. *The Metabolic and Molecular Bases of Inherited Diseases* (8th ed., pp. 2707–2960). McGraw-Hill.

Phospholipids and Sphingolipids

- Carman, G. M., and Han, G.-S. 2009. Phosphatidic acid phosphatase, a key enzyme in the regulation of lipid synthesis. J. Biol. Chem. 284:2593–2597.
- Bartke, N., and Hannun, Y. A. 2009. Bioactive sphingolipids: Metabolism and function. J. Lipid Res. 50:S91–S96.
- Lee, J., Johnson, J., Ding, Z., Paetzel, M., and Cornell, R. B. 2009. Crystal structure of a mammalian CTP: Phosphocholine cytidylyltransferase catalytic domain reveals novel active site residues within a highly conserved nucleotidyltransferase fold. *J. Biol. Chem.* 284:33535–33548.
- Nye, C. K., Hanson, R. W., and Kalhan, S. C. 2008. Glyceroneogenesis is the dominant pathway for triglyceride glycerol synthesis in vivo in the rat. J. Biol. Chem. 283:27565–27574.
- Kent, C. 1995. Eukaryotic phospholipid biosynthesis. Annu. Rev. Biochem. 64:315–343.

Biosynthesis of Cholesterol and Steroids

- Radhakrishnan, A., Goldstein, J. L., McDonald, J. G., and Brown, M. S. 2008. Switch-like control of SREBP-2 transport triggered by small changes in ER cholesterol: A delicate balance. *Cell Metab.* 8:512–521.
- DeBose-Boyd, R. A. 2008. Feedback regulation of cholesterol synthesis: Sterol-accelerated ubiquitination and degradation of HMG CoA reductase. *Cell Res.* 18:609–621.
- Hampton, R. Y. 2002. Proteolysis and sterol regulation. Annu. Rev. Cell Dev. Biol. 18:345–378.
- Kelley, R. I., and Herman, G. E. 2001. Inborn errors of sterol biosynthesis. Annu. Rev. Genom. Hum. Genet. 2:299-341.
- Istvan, E. S., and Deisenhofer, J. 2001. Structural mechanism for statin inhibition of HMG-CoA reductase. *Science* 292:1160–1164.
- Ness, G. C., and Chambers, C. M. 2000. Feedback and hormonal regulation of hepatic 3-hydroxy-3-methylglutaryl coenzyme A reductase: The concept of cholesterol buffering capacity. *Proc. Soc. Exp. Biol. Med.* 224:8–19.
- Libby, P., Aikawa, M., and Schonbeck, U. 2000. Cholesterol and atherosclerosis. *Biochim. Biophys. Acta* 1529:299–309.
- Yokoyama, S. 2000. Release of cellular cholesterol: Molecular mechanism for cholesterol homeostasis in cells and in the body. *Biochim. Biophys. Acta* 1529:231–244.
- Cronin, S. R., Khoury, A., Ferry, D. K., and Hampton, R. Y. 2000. Regulation of HMG-CoA reductase degradation requires the P-type ATPase Cod1p/Spf1p. J. Cell Biol. 148:915–924.
- Edwards, P. A., Tabor, D., Kast, H. R., and Venkateswaran, A. 2000. Regulation of gene expression by SREBP and SCAP. *Biochim. Biophys. Acta* 1529:103–113.
- Istvan, E. S., Palnitkar, M., Buchanan, S. K., and Deisenhofer, J. 2000. Crystal structure of the catalytic portion of human HMG-CoA reductase: Insights into regulation of activity and catalysis. *EMBO* J. 19:819–830.

Jeon, H., Meng, W., Takagi, J., Eck, M. J., Springer, T. A., and Blacklow, S. C. 2001. Implications for familial hypercholesterolemia from the structure of the LDL receptor YWTD-EGF domain pair. *Nat. Struct. Biol.* 8:499–504.

Lipoproteins and Their Receptors

- Rye, K-A., Bursill, C. A., Lambert, G., Tabet, F., and Barter, P. J. 2009. The metabolism and anti-atherogenic properties of HDL. J. Lipid Res. 50:S195–S200.
- Rader, D. J., Alexander, E. T., Weibel, G. L., Billheimer, J., and Rothblat, G. H. 2009. The role of reverse cholesterol transport in animals and humans and relationship to atherosclerosis. J. Lipid Res. 50:S189–S194.
- Tall, A. R., Yvan-Charvet, L., Terasaka, N., Pagler, T., and Wang, N. 2008. HDL, ABC transporters, and cholesterol efflux: Implications for the treatment of atherosclerosis. *Cell Metab.* 7:365–375.
- Jeon, H., and Blacklow, S. C. 2005. Structure and physiologic function of the low-density lipoprotein receptor. Annu. Rev. Biochem. 74:535–562.
- Beglova, N., and Blacklow, S. C. 2005. The LDL receptor: How acid pulls the trigger. *Trends Biochem. Sci.* 30:309–316.
- Brouillette, C. G., Anantharamaiah, G. M., Engler, J. A., and Borhani, D. W. 2001. Structural models of human apolipoprotein A-I: A critical analysis and review. *Biochem. Biophys. Acta* 1531:4–46.
- Hevonoja, T., Pentikainen, M. O., Hyvonen, M. T., Kovanen, P. T., and Ala-Korpela, M. 2000. Structure of low density lipoprotein (LDL) particles: Basis for understanding molecular changes in modified LDL. Biochim. Biophys. Acta 1488:189–210.
- Silver, D. L., Jiang, X. C., Arai, T., Bruce, C., and Tall, A. R. 2000. Receptors and lipid transfer proteins in HDL metabolism. Ann. N. Y. Acad. Sci. 902:103–111.
- Nimpf, J., and Schneider, W. J. 2000. From cholesterol transport to signal transduction: Low density lipoprotein receptor, very low density lipoprotein receptor, and apolipoprotein E receptor-2. *Biochim. Biophys. Acta* 1529:287–298.

Oxygen Activation and P450 Catalysis

- Stiles, A. R., McDonald, J. G., Bauman, D. R., and Russell, D. W. 2009. CYP7B1: One cytochrome P450, two human genetic diseases, and multiple physiological functions. *J. Biol. Chem.* 284:28485–28489.
- Zhou, S.-F., Liu, J.-P., and Chowbay, B. 2009. Polymorphism of human cytochrome P450 enzymes and its clinical impact. *Drug Metab. Rev.* 4:89–295.
- Williams, P. A., Cosme, J., Vinkovic, D. M., Ward, A., Angove, H. C., Day, P. J., Vonrhein, C., Tickle, I. J., and Jhoti, H. 2004. Crystal structure of human cytochrome P450 3A4 bound to metyrapone and progesterone. *Science* 305:683–686.
- Ingelmn-Sundberg, M., Oscarson, M., and McLellan, R. A. 1999. Polymorphic human cytochrome P450 enzymes: An opportunity for individualized drug treatment. *Trends Pharmacol. Sci.* 20:342–349.

Chapter 27

Books

- Wrangham, R. 2009. Catching Fire: How Cooking Made Us Human. Basic Books.
- Stipanuk, M. H. (Ed.). 2006. Biochemical, Physiological, & Molecular Aspects of Human Nutrition. Saunders-Elsevier.
- Fell, D. 1997. Understanding the Control of Metabolism. Portland Press.
- Frayn, K. N. 1996. Metabolic Regulation: A Human Perspective. Portland Press.
- Poortmans, J. R. (Ed.). 2004. Principles of Exercise Biochemistry. Karger.
- Harris, R. A., and Crabb, D. W. 2006. Metabolic interrelationships. In Textbook of Biochemistry with Clinical Correlations (pp. 849–890), edited by T. M. Devlin. Wiley-Liss.

SELECTED READINGS

Caloric Homeostasis

- Woods, S. C. 2009. The control of food intake: Behavioral versus molecular perspectives. *Cell Metab.* 9:489–498.
- Figlewicz, D. P., and Benoit, S. C. 2009. Insulin, leptin, and food reward: Update 2008. Am. J. Physiol. Integr. Comp. Physiol. 296:R9-R19.
- Israel, D., and Chua, S. Jr. 2009. Leptin receptor modulation of adiposity and fertility. *Trends Endocrinol. Metab.* 21:10–16.
- Meyers, M. G., Cowley, M. A., and Münzberg, H. 2008. Mechanisms of leptin action and leptin resistance. Annu. Rev. Physiol. 70:537–556.
- Sowers, J. R. 2008. Endocrine functions of adipose tissue: Focus on adiponectin. *Clin. Cornerstone* 9:32–38.
- Brehma, B. J., and D'Alessio, D. A. 2008. Benefits of high-protein weight loss diets: Enough evidence for practice? Curr. Opin. Endocrinol., Diabetes, Obesity 15:416–421.
- Coll, A. P., Farooqi, I. S., and O'Rahillt, S. O. 2007. The hormonal control of food intake. *Cell* 129:251–262.
- Muoio, D. M., and Newgard, C. B. 2006. Obesity-related derangements in metabolic regulation. Annu. Rev. Biochem. 75:367–401.

Diabetes Mellitus

- Zhang, B. B., Zhou, G., and Li, C. 2009. AMPK: An emerging drug target for diabetes and the metabolic syndrome. *Cell Metab.* 9:407–416.
- Magkos, F., Yannakoulia, M., Chan, J. L., and Mantzoros, C. S. 2009. Management of the metabolic syndrome and type 2 diabetes through lifestyle modification. *Annu. Rev. Nutr.* 29:8.1–8.34.
- Muoio, D. M., and Newgard, C. B. 2008. Molecular and metabolic mechanisms of insulin resistance and β-cell failure in type 2 diabetes. Nat. Rev. Mol. Cell. Biol. 9:193–205.
- Leibiger, I. B., Leibiger, B., and Berggren, P.-O. 2008. Insulin signaling in the pancreatic β-cell. Annu. Rev. Nutr. 28:233–251.
- Doria, A., Patti, M. E., and Kahn, C. R. 2008. The emerging architecture of type 2 diabetes. *Cell Metab.* 8:186–200.
- Croker, B. A., Kiu, H., and Nicholson, S. E. 2008. SOCS regulation of the JAK/STAT signalling pathway. Semin. Cell Dev. Biol. 19:414–422.
- Eizirik, D. L., Cardozo, A. K., and Cnop, M. 2008. The role of endoplasmic reticulum stress in diabetes mellitus. *Endocrinol. Rev.* 29:42–61.
- Howard, J. K., and Flier, J. S. 2006. Attenuation of leptin and insulin signaling by SOCS proteins. *Trends Endocrinol. Metab.* 9:365–371.
- Lowel, B. B., and Shulman, G. 2005. Mitochondrial dysfunction and type 2 diabetes. Science 307:384–387.
- Taylor, S. I. 2001. Diabetes mellitus. In *The Metabolic Basis of Inherited Diseases* (8th ed., pp. 1433–1469), edited by C. R. Scriver, W. S. Sly, B. Childs, A. L. Beaudet, D. Valle, K. W. Kinzler, and B. Vogelstein. McGraw-Hill.

Exercise Metabolism

- Hood, D. A. 2001. Contractile activity-induced mitochondrial biogenesis in skeletal muscle. J. Appl. Physiol. 90:1137–1157.
- Shulman, R. G., and Rothman, D. L. 2001. The "glycogen shunt" in exercising muscle: A role for glycogen in muscle energetics and fatigue. Proc. Natl. Acad. Sci. U.S.A. 98:457–461.
- Gleason, T. 1996. Post-exercise lactate metabolism: A comparative review of sites, pathways, and regulation. Annu. Rev. Physiol. 58:556–581.
- Holloszy, J. O., and Kohrt, W. M. 1996. Regulation of carbohydrate and fat metabolism during and after exercise. *Annu. Rev. Nutr.* 16:121–138.
- Hochachka, P. W., and McClelland, G. B. 1997. Cellular metabolic homeostasis during large-scale change in ATP turnover rates in muscles. J. Exp. Biol. 200:381–386.
- Horowitz, J. F., and Klein, S. 2000. Lipid metabolism during endurance exercise. Am. J. Clin. Nutr. 72:558S–563S.
- Wagenmakers, A. J. 1999. Muscle amino acid metabolism at rest and during exercise. *Diabetes Nutr. Metab.* 12:316–322.

Metabolic Adaptations in Starvation

- Baverel, G., Ferrier, B., and Martin, M. 1995. Fuel selection by the kidney: Adaptation to starvation. *Proc. Nutr. Soc.* 54:197–212.
- MacDonald, I. A., and Webber, J. 1995. Feeding, fasting and starvation: Factors affecting fuel utilization. Proc. Nutr. Soc. 54:267–274.
- Cahill, G. F., Jr. 1976. Starvation in man. Clin. Endocrinol. Metab. 5:397–415.
- Sugden, M. C., Holness, M. J., and Palmer, T. N. 1989. Fuel selection and carbon flux during the starved-to-fed transition. *Biochem.* J. 263:313–323.

Ethanol Metabolism

- Nagy, L. E. 2004. Molecular aspects of alcohol metabolism: Transcription factors involved in early-induced liver injury. Annu. Rev. Nutr. 24:55–78.
- Molotkov, A., and Duester, G. 2002. Retinol/ethanol drug interaction during acute alcohol intoxication involves inhibition of retinol metabolism to retinoic acid by alcohol dehydrogenase. J. Biol. Chem. 277:22553–22557.
- Stewart, S., Jones, D., and Day, C. P. 2001. Alcoholic liver disease: New insights into mechanisms and preventive strategies. *Trends Mol. Med.* 7:408–413.
- Lieber, C. S. 2000. Alcohol: Its metabolism and interaction with nutrients. Annu. Rev. Nutr. 20:395–430.
- Niemela, O. 1999. Aldehyde-protein adducts in the liver as a result of ethanol-induced oxidative stress. *Front. Biosci.* 1:D506–D513.
- Riveros-Rosas, H., Julian-Sanchez, A., and Pina, E. 1997. Enzymology of ethanol and acetaldehyde metabolism in mammals. Arch. Med. Res. 28:453–471.

Chapter 28

Where to Start

- Johnson, A., and O'Donnell, M. 2005. Cellular DNA replicases: Components and dynamics at the replication fork. *Annu. Rev. Biochem.* 74:283-315.
- Kornberg, A. 1988. DNA replication. J. Biol. Chem. 263:1-4.
- Wang, J. C. 1982. DNA topoisomerases. Sci. Am. 247(1):94-109.
- Lindahl, T. 1993. Instability and decay of the primary structure of DNA. Nature 362:709-715.
- Greider, C. W., and Blackburn, E. H. 1996. Telomeres, telomerase, and cancer. Sci. Am. 274(2):92-97.

Books

- Kornberg, A., and Baker, T. A. 1992. *DNA Replication* (2d ed.). W. H. Freeman and Company.
- Bloomfield, V. A., Crothers, D., Tinoco, I., and Hearst, J. 2000. Nucleic Acids: Structures, Properties and Functions. University Science Books.
- Friedberg, E. C., Walker, G. C., and Siede, W. 1995. DNA Repair and Mutagenesis. American Society for Microbiology.
- Cozzarelli, N. R., and Wang, J. C. (Eds.). 1990. DNA Topology and Its Biological Effects. Cold Spring Harbor Laboratory Press.

DNA Topology and Topoisomerases

- Graille, M., Cladiere, L., Durand, D., Lecointe, F., Gadelle, D., Quevillon-Cheruel, S., Vachette, P., Forterre, P., and van Tilbeurgh, H. 2008. Crystal structure of an intact type II DNA topoisomerase: Insights into DNA transfer mechanisms. *Structure* 16:360–370.
- Charvin, G., Strick, T. R., Bensimon, D., and Croquette, V. 2005. Tracking topoisomerase activity at the single-molecule level. *Annu. Rev. Biophys. Biomol. Struct.* 34:201–219.
- Sikder, D., Unniraman, S., Bhaduri, T., and Nagaraja, V. 2001. Functional cooperation between topoisomerase I and single strand DNA-binding protein. J. Mol. Biol. 306:669-679.
- Yang, Z., and Champoux, J. J. 2001. The role of histidine 632 in catalysis by human topoisomerase I. J. Biol. Chem. 276:677-685.
B29

- Fortune, J. M., and Osheroff, N. 2000. Topoisomerase II as a target for anticancer drugs: When enzymes stop being nice. Prog. Nucleic Acid Res. Mol. Biol. 64:221-253.
- Isaacs, R. J., Davies, S. L., Sandri, M. I., Redwood, C., Wells, N. J., and Hickson, I. D. 1998. Physiological regulation of eukaryotic topoisomerase II. *Biochim. Biophys. Acta* 1400:121–137.
- Wang, J. C. 1996. DNA topoisomerases. Annu. Rev. Biochem. 65:635-692.
- Wang, J. C. 1998. Moving one DNA double helix through another by a type II DNA topoisomerase: The story of a simple molecular machine. Q. Rev. Biophys. 31:107-144.
- Baird, C. L., Harkins, T. T., Morris, S. K., and Lindsley, J. E. 1999. Topoisomerase II drives DNA transport by hydrolyzing one ATP. *Proc. Natl. Acad. Sci. U.S.A.* 96:13685–13690.
- Vologodskii, A. V., Levene, S. D., Klenin, K. V., Frank, K. M., and Cozzarelli, N. R. 1992. Conformational and thermodynamic properties of supercoiled DNA. J. Mol. Biol. 227:1224-1243.
- Fisher, L. M., Austin, C. A., Hopewell, R., Margerrison, M., Oram, M., Patel, S., Wigley, D. B., Davies, G. J., Dodson, E. J., Maxwell, A., and Dodson, G. 1991. Crystal structure of an N-terminal fragment of the DNA gyrase B protein. *Nature* 351:624–629.

Mechanism of Replication

- Davey, M. J., and O'Donnell, M. 2000. Mechanisms of DNA replication. Curr. Opin. Chem. Biol. 4:581-586.
- Keck, J. L., and Berger, J. M. 2000. DNA replication at high resolution. *Chem. Biol.* 7:R63–R71.
- Kunkel, T. A., and Bebenek, K. 2000. DNA replication fidelity. Annu. Rev. Biochem. 69:497-529.
- Waga, S., and Stillman, B. 1998. The DNA replication fork in eukaryotic cells. Annu. Rev. Biochem. 67:721-751.
- Marians, K. J. 1992. Prokaryotic DNA replication. Annu. Rev. Biochem. 61:673-719.

DNA Polymerases and Other Enzymes of Replication

- Singleton, M. R., Sawaya, M. R., Ellenberger, T., and Wigley, D. B. 2000. Crystal structure of T7 gene 4 ring helicase indicates a mechanism for sequential hydrolysis of nucleotides. *Cell* 101:589–600.
- Donmez, I., and Patel, S. S. 2006. Mechanisms of a ring shaped helicase. Nucleic Acids Res. 34:4216–4224.
- Johnson, D. S., Bai, L., Smith, B. Y., Patel, S. S., and Wang, M. D. 2007. Single-molecule studies reveal dynamics of DNA unwinding by the ring-shaped T7 helicase. *Cell* 129:1299–1309.
- Lee, S. J., Qimron, U., and Richardson, C. C. 2008. Communication between subunits critical to DNA binding by hexameric helicase of bacteriophage T7. Proc. Natl. Acad. Sci. U.S.A. 105:8908-8913.
- Toth, E. A., Li, Y., Sawaya, M. R., Cheng, Y., and Ellenberger, T. 2003. The crystal structure of the bifunctional primase-helicase of bacteriophage T7. *Mol. Cell* 12:1113–1123.
- Hubscher, U., Maga, G., and Spadari, S. 2002. Eukaryotic DNA polymerases. Annu. Rev. Biochem. 71:133-163.
- Doublié, S., Tabor, S., Long, A. M., Richardson, C. C., and Ellenberger, T. 1998. Crystal structure of a bacteriophage T7 DNA replication complex at 2.2 Å resolution. *Nature* 391:251–258.
- Arezi, B., and Kuchta, R. D. 2000. Eukaryotic DNA primase. Trends Biochem. Sci. 25:572-576.
- Jager, J., and Pata, J. D. 1999. Getting a grip: Polymerases and their substrate complexes. Curr. Opin. Struct. Biol. 9:21-28.
- Steitz, T. A. 1999. DNA polymerases: Structural diversity and common mechanisms. J. Biol. Chem. 274:17395–17398.
- Beese, L. S., Derbyshire, V., and Steitz, T. A. 1993. Structure of DNA polymerase I Klenow fragment bound to duplex DNA. *Science* 260:352-355.
- McHenry, C. S. 1991. DNA polymerase III holoenzyme: Components, structure, and mechanism of a true replicative complex. J. Biol. Chem. 266:19127-19130.

- Kong, X. P., Onrust, R., O'Donnell, M., and Kuriyan, J. 1992. Threedimensional structure of the β subunit of *E. coli* DNA polymerase III holoenzyme: A sliding DNA clamp. *Cell* 69:425–437.
- Polesky, A. H., Steitz, T. A., Grindley, N. D., and Joyce, C. M. 1990. Identification of residues critical for the polymerase activity of the Klenow fragment of DNA polymerase I from *Escherichia coli*. J. Biol. Chem. 265:14579-14591.
- Lee, J. Y., Chang, C., Song, H. K., Moon, J., Yang, J. K., Kim, H. K., Kwon, S. T., and Suh, S. W. 2000. Crystal structure of NAD⁺ dependent DNA ligase: Modular architecture and functional implications. *EMBO J.* 19:1119–1129.
- Timson, D. J., and Wigley, D. B. 1999. Functional domains of an NAD⁺-dependent DNA ligase. *J. Mol. Biol.* 285:73–83.
- Doherty, A. J., and Wigley, D. B. 1999. Functional domains of an ATPdependent DNA ligase. J. Mol. Biol. 285:63-71.
- von Hippel, P. H., and Delagoutte, E. 2001. A general model for nucleic acid helicases and their "coupling" within macromolecular machines. *Cell* 104:177-190.
- Tye, B. K., and Sawyer, S. 2000. The hexameric eukaryotic MCM helicase: Building symmetry from nonidentical parts. J. Biol. Chem. 275:34833-34836.
- Marians, K. J. 2000. Crawling and wiggling on DNA: Structural insights to the mechanism of DNA unwinding by helicases. *Struct. Fold. Des.* 5:R227–R235.
- Soultanas, P., and Wigley, D. B. 2000. DNA helicases: "Inching forward." Curr. Opin. Struct. Biol. 10:124-128.
- de Lange, T. 2009. How telomeres solve the end-protection problem. Science 326:948-952.
- Bachand, F., and Autexier, C. 2001. Functional regions of human telomerase reverse transcriptase and human telomerase RNA required for telomerase activity and RNA-protein interactions. *Mol. Cell Biol.* 21:1888–1897.
- Bryan, T. M., and Cech, T. R. 1999. Telomerase and the maintenance of chromosome ends. *Curr. Opin. Cell Biol.* 11:318-324.
- Griffith, J. D., Comeau, L., Rosenfield, S., Stansel, R. M., Bianchi, A., Moss, H., and de Lange, T. 1999. Mammalian telomeres end in a large duplex loop. *Cell* 97:503–514.
- McEachern, M. J., Krauskopf, A., and Blackburn, E. H. 2000. Telomeres and their control. *Annu. Rev. Genet.* 34:331–358.

Mutations and DNA Repair

- Yang, W. 2003. Damage repair DNA polymerases Y. Curr. Opin. Struct. Biol. 13:23–30.
- Wood, R. D., Mitchell, M., Sgouros, J., and Lindahl, T. 2001. Human DNA repair genes. *Science* 291:1284–1289.
- Shin, D. S., Chahwan, C., Huffman, J. L., and Tainer, J. A. 2004. Structure and function of the double-strand break repair machinery. DNA Repair (Amst.) 3:863-873.
- Michelson, R. J., and Weinert, T. 2000. Closing the gaps among a web of DNA repair disorders. *Bioessays* 22:966–969.
- Aravind, L., Walker, D. R., and Koonin, E. V. 1999. Conserved domains in DNA repair proteins and evolution of repair systems. *Nucleic Acids Res.* 27:1223–1242.
- Mol, C. D., Parikh, S. S., Putnam, C. D., Lo, T. P., and Tainer, J. A. 1999. DNA repair mechanisms for the recognition and removal of damaged DNA bases. Annu. Rev. Biophys. Biomol. Struct. 28:101–128.
- Parikh, S. S., Mol, C. D., and Tainer, J. A. 1997. Base excision repair enzyme family portrait: Integrating the structure and chemistry of an entire DNA repair pathway. *Structure* 5:1543-1550.
- Vassylyev, D. G., and Morikawa, K. 1997. DNA-repair enzymes. Curr. Opin. Struct. Biol. 7:103-109.
- Verdine, G. L., and Bruner, S. D. 1997. How do DNA repair proteins locate damaged bases in the genome? *Chem. Biol.* 4:329-334.
- Bowater, R. P., and Wells, R. D. 2000. The intrinsically unstable life of DNA triplet repeats associated with human hereditary disorders. *Prog. Nucleic Acid Res. Mol. Biol.* 66:159–202.

SELECTED READINGS

Cummings, C. J., and Zoghbi, H. Y. 2000. Fourteen and counting: Unraveling trinucleotide repeat diseases. *Hum. Mol. Genet.* 9:909-916.

Defective DNA Repair and Cancer

- Berneburg, M., and Lehmann, A. R. 2001. Xeroderma pigmentosum and related disorders: Defects in DNA repair and transcription. *Adv. Genet.* 43:71–102.
- Lambert, M. W., and Lambert, W. C. 1999. DNA repair and chromatin structure in genetic diseases. Prog. Nucleic Acid Res. Mol. Biol. 63:257-310.
- Buys, C. H. 2000. Telomeres, telomerase, and cancer. N. Engl. J. Med. 342:1282–1283.
- Urquidi, V., Tarin, D., and Goodison, S. 2000. Role of telomerase in cell senescence and oncogenesis. Annu. Rev. Med. 51:65-79.
- Lynch, H. T., Smyrk, T. C., Watson, P., Lanspa, S. J., Lynch, J. F., Lynch, P. M., Cavalieri, R. J., and Boland, C. R. 1993. Genetics, natural history, tumor spectrum, and pathology of hereditary nonpolyposis colorectal cancer: An updated review. *Gastroenterology* 104:1535-1549.
- Fishel, R., Lescoe, M. K., Rao, M. R. S., Copeland, N. G., Jenkins, N. A., Garber, J., Kane, M., and Kolodner, R. 1993. The human mutator gene homolog *MSH2* and its association with hereditary nonpolyposis colon cancer. *Cell* 75:1027-1038.
- Ames, B. N., and Gold, L. S. 1991. Endogenous mutagens and the causes of aging and cancer. *Mutat. Res.* 250:3–16.
- Ames, B. N. 1979. Identifying environmental chemicals causing mutations and cancer. Science 204:587-593.

Recombination and Recombinases

- Singleton, M. R., Dillingham, M. S., Gaudier, M., Kowalczykowski, S. C., and Wigley, D. B. 2004. Crystal structure of RecBCD enzyme reveals a machine for processing DNA breaks. *Nature* 432:187-193.
- Spies, M., Bianco, P. R., Dillingham, M. S., Handa, N., Baskin, R. J., and Kowalczykowski, S. C. 2003. A molecular throttle: The recombination hotspot chi controls DNA translocation by the RecBCD helicase. *Cell* 114:647-654.
- Kowalczykowski, S. C. 2000. Initiation of genetic recombination and recombination-dependent replication. *Trends Biochem. Sci.* 25:156-165.
- Prevost, C., and Takahashi, M. 2003. Geometry of the DNA strands within the RecA nucleofilament: Role in homologous recombination. Q. Rev. Biophys. 36:429-453.
- Van Duyne, G. D. 2001. A structural view of Cre-loxP site-specific recombination. Annu. Rev. Biophys. Biomol. Struct. 30:87-104.
- Chen, Y., Narendra, U., Iype, L. E., Cox, M. M., and Rice, P. A. 2000. Crystal structure of a Flp recombinase-Holliday junction complex: Assembly of an active oligomer by helix swapping. *Mol. Cell* 6:885-897.
- Craig, N. L. 1997. Target site selection in transposition. Annu. Rev. Biochem. 66:437-474.
- Gopaul, D. N., Guo, F., and Van Duyne, G. D. 1998. Structure of the Holliday junction intermediate in Cre-loxP site-specific recombination. *EMBO J.* 17:4175–4187.
- Gopaul, D. N., and Duyne, G. D. 1999. Structure and mechanism in site-specific recombination. Curr. Opin. Struct. Biol. 9:14-20.

Chapter 29

Where to Start

- Kornberg, R. D. 2007. The molecular basis of eukaryotic transcription. Proc. Natl. Acad. Sci. U.S.A. 104:12955–12961.
- Woychik, N. A. 1998. Fractions to functions: RNA polymerase II thirty years later. Cold Spring Harbor Symp. Quant. Biol. 63:311-317.

- Losick, R. 1998. Summary: Three decades after sigma. Cold Spring Harbor Symp. Quant. Biol. 63:653–666.
- Ast, G. 2005. The alternative genome. Sci. Am. 292(4):40-47.
- Sharp, P. A. 1994. Split genes and RNA splicing (Nobel Lecture). Angew. Chem. Int. Ed. Engl. 33:1229–1240.
- Cech, T. R. 1990. Nobel lecture: Self-splicing and enzymatic activity of an intervening sequence RNA from *Tetrahymena*. Biosci. Rep. 10:239–261.
- Villa, T., Pleiss, J. A., and Guthrie, C. 2002. Spliceosomal snRNAs: Mg²⁺ dependent chemistry at the catalytic core? *Cell* 109:149–152.

Books

- Lewin, B. 2007. Genes (9th ed.). Jones and Bartlett.
- Kornberg, A., and Baker, T. A. 1992. *DNA Replication* (2d ed.). W. H. Freeman and Company.
- Lodish, H., Berk, A., Matsudaira, P., Krieger, M., Kaiser, C. A., Scott, M. P., Bretscher, A., Plough, H., and Darnell, J. 2008. *Molecular Cell Biology* (6th ed.). W. H. Freeman and Company.
- Watson, J. D., Baker, T. A., Bell, S. P., Gann, A., Levine, M., and Losick, R. 2004. *Molecular Biology of the Gene* (5th ed.). Pearson/ Benjamin Cummings.
- Gesteland, R. F., Cech, T., and Atkins, J. F. 2006. The RNA World: The Nature of Modern RNA Suggests a Prebiotic RNA (3d ed.) Cold Spring Harbor Laboratory Press.

RNA Polymerases

- Liu, X., Bushnell, D. A., Wang, D., Calero, G., and Kornberg, R. D. 2010. Structure of an RNA polymerase II-TFIIB complex and the transcription initiation mechanism. *Science* 327:206–209.
- Wang, D., Bushnell, D. A., Huang, X., Westover, K. D., Levitt, M., and Kornberg, R. D. 2009. Structural basis of transcription: Backtracked RNA polymerase II at 3.4 angstrom resolution. *Science* 324:1203–1206.
- Darst, S. A. 2001. Bacterial RNA polymerase. Curr. Opin. Struct. Biol. 11:155–162.
- Ross, W., Gosink, K. K., Salomon, J., Igarashi, K., Zou, C., Ishihama, A., Severinov, K., and Gourse, R. L. 1993. A third recognition element in bacterial promoters: DNA binding by the alpha subunit of RNA polymerase. *Science* 262:1407–1413.
- Cramer, P., Bushnell, D. A., and Kornberg, R. D. 2001. Structural basis of transcription: RNA polymerase II at 2.8 Å resolution. *Science* 292:1863–1875.
- Gnatt, A. L., Cramer, P., Fu, J., Bushnell, D. A., and Kornberg, R. D. 2001. Structural basis of transcription: An RNA polymerase II elongation complex at 3.3 Å resolution. *Science* 292:1876–1882.
- Zhang, G., Campbell, E. A., Minakhin, L., Richter, C., Severinov, K., and Darst, S. A. 1999. Crystal structure of *Thermus aquaticus* core RNA polymerase at 3.3 Å resolution. *Cell* 98:811–824.
- Campbell, E. A., Korzheva, N., Mustaev, A., Murakami, K., Nair, S., Goldfarb, A., and Darst, S. A. 2001. Structural mechanism for rifampicin inhibition of bacterial RNA polymerase. *Cell* 104:901–912.
- Darst, S. A. 2004. New inhibitors targeting bacterial RNA polymerase. Trends Biochem. Sci. 29:159–160.
- Cheetham, G. M., and Steitz, T. A. 1999. Structure of a transcribing T7 RNA polymerase initiation complex. *Science* 286:2305–2309.
- Ebright, R. H. 2000. RNA polymerase: Structural similarities between bacterial RNA polymerase and eukaryotic RNA polymerase II. *J. Mol. Biol.* 304:687–698.
- Paule, M. R., and White, R. J. 2000. Survey and summary: Transcription by RNA polymerases I and III. Nucleic Acids Res. 28:1283–1298.

Initiation and Elongation

- Murakami, K. S., and Darst, S. A. 2003. Bacterial RNA polymerases: The whole story. *Curr. Opin. Struct. Biol.* 13:31–39.
- Buratowski, S. 2000. Snapshots of RNA polymerase II transcription initiation. Curr. Opin. Cell Biol. 12:320–325.

- Conaway, J. W., and Conaway, R. C. 1999. Transcription elongation and human disease. Annu. Rev. Biochem. 68:301–319.
- Conaway, J. W., Shilatifard, A., Dvir, A., and Conaway, R. C. 2000. Control of elongation by RNA polymerase II. *Trends Biochem. Sci.* 25:375–380.
- Korzheva, N., Mustaev, A., Kozlov, M., Malhotra, A., Nikiforov, V., Goldfarb, A., and Darst, S. A. 2000. A structural model of transcription elongation. *Science* 289:619–625.
- Reines, D., Conaway, R. C., and Conaway, J. W. 1999. Mechanism and regulation of transcriptional elongation by RNA polymerase II. *Curr. Opin. Cell Biol.* 11:342–346.

Promoters, Enhancers, and Transcription Factors

- Merika, M., and Thanos, D. 2001. Enhanceosomes. Curr. Opin. Genet. Dev. 11:205–208.
- Park, J. M., Gim, B. S., Kim, J. M., Yoon, J. H., Kim, H. S., Kang, J. G., and Kim, Y. J. 2001. *Drosophila* mediator complex is broadly utilized by diverse gene-specific transcription factors at different types of core promoters. *Mol. Cell. Biol.* 21:2312–2323.
- Smale, S. T., and Kadonaga, J. T. 2003. The RNA polymerase II core promoter. Annu. Rev. Biochem. 72:449–479.
- Gourse, R. L., Ross, W., and Gaal, T. 2000. Ups and downs in bacterial transcription initiation: The role of the alpha subunit of RNA polymerase in promoter recognition. *Mol. Microbiol.* 37:687–695.
- Fiering, S., Whitelaw, E., and Martin, D. I. 2000. To be or not to be active: The stochastic nature of enhancer action. *Bioessays* 22:381–387.
- Hampsey, M., and Reinberg, D. 1999. RNA polymerase II as a control panel for multiple coactivator complexes. *Curr. Opin. Genet. Dev.* 9:132–139.
- Chen, L. 1999. Combinatorial gene regulation by eukaryotic transcription factors. Curr. Opin. Struct. Biol. 9:48–55.
- Muller, C. W. 2001. Transcription factors: Global and detailed views. Curr. Opin. Struct. Biol. 11:26–32.
- Reese, J. C. 2003. Basal transcription factors. Curr. Opin. Genet. Dev. 13:114–118.
- Kadonaga, J. T. 2004. Regulation of RNA polymerase II transcription by sequence-specific DNA binding factors. *Cell* 116:247–257.
- Harrison, S. C. 1991. A structural taxonomy of DNA-binding domains. *Nature* 353:715–719.
- Sakurai, H., and Fukasawa, T. 2000. Functional connections between mediator components and general transcription factors of Saccharomyces cerevisiae. J. Biol. Chem. 275:37251–37256.
- Droge, P., and Muller-Hill, B. 2001. High local protein concentrations at promoters: Strategies in prokaryotic and eukaryotic cells. *Bioessays* 23:179–183.
- Smale, S. T., Jain, A., Kaufmann, J., Emami, K. H., Lo, K., and Garraway, I. P. 1998. The initiator element: A paradigm for core promoter heterogeneity within metazoan protein-coding genes. *Cold Spring Harbor Symp. Quant. Biol.* 63:21–31.
- Kim, Y., Geiger, J. H., Hahn, S., and Sigler, P. B., 1993. Crystal structure of a yeast TBP/TATA-box complex. *Nature* 365:512–520.
- Kim, J. L., Nikolov, D. B., and Burley, S. K., 1993. Co-crystal structure of TBP recognizing the minor groove of a TATA element. *Nature* 365:520–527.
- White, R. J., and Jackson, S. P., 1992. The TATA-binding protein: A central role in transcription by RNA polymerases I, II and III. *Trends Genet.* 8:284–288.
- Martinez, E. 2002. Multi-protein complexes in eukaryotic gene transcription. Plant Mol. Biol. 50:925–947.
- Meinhart, A., Kamenski, T., Hoeppner, S., Baumli, S., and Cramer, P. 2005. A structural perspective of CTD function. *Genes Dev.* 19:1401–1415.
- Palancade, B., and Bensaude, O. 2003. Investigating RNA polymerase II carboxyl-terminal domain (CTD) phosphorylation. *Eur.* J. Biochem. 270:3859–3870.

Termination

- Burgess, B. R., and Richardson, J. P. 2001. RNA passes through the hole of the protein hexamer in the complex with *Escherichia coli* Rho factor. J. Biol. Chem. 276:4182–4189.
- Yu, X., Horiguchi, T., Shigesada, K., and Egelman, E. H. 2000. Threedimensional reconstruction of transcription termination factor rho: Orientation of the N-terminal domain and visualization of an RNA-binding site. J. Mol. Biol. 299:1279–1287.
- Stitt, B. L. 2001. Escherichia coli transcription termination factor Rho binds and hydrolyzes ATP using a single class of three sites. Biochemistry 40:2276–2281.
- Henkin, T. M. 2000. Transcription termination control in bacteria. Curr. Opin. Microbiol. 3:149–153.
- Gusarov, I., and Nudler, E. 1999. The mechanism of intrinsic transcription termination. *Mol. Cell* 3:495–504.

Riboswitches

- Barrick, J. E., and Breaker, R. R. 2007. The distributions, mechanisms, and structures of metabolite-binding riboswitches. *Genome Biol.* 8:R239.
- Cheah, M. T., Wachter, A., Sudarsan, N., and Breaker, R. R. 2007. Control of alternative RNA splicing and gene expression by eukaryotic riboswitches. *Nature* 447:497–500.
- Serganov, A., Huang, L., and Patel, D. J. 2009. Coenzyme recognition and gene regulation by a flavin mononucleotide riboswitch. *Nature* 458:233–237.

Noncoding RNA

- Peculis, B. A. 2002. Ribosome biogenesis: Ribosomal RNA synthesis as a package deal. Curr. Biol. 12:R623–R624.
- Decatur, W. A., and Fournier, M. J. 2002. rRNA modifications and ribosome function. *Trends Biochem. Sci.* 27:344–351.
- Hopper, A. K., and Phizicky, E. M. 2003. tRNA transfers to the limelight. Genes Dev. 17:162–180.
- Weiner, A. M. 2004. tRNA maturation: RNA polymerization without a nucleic acid template. Curr. Biol. 14:R883–R885.

5'-Cap Formation and Polyadenylation

- Shatkin, A. J., and Manley, J. L. 2000. The ends of the affair: Capping and polyadenylation. Nat. Struct. Biol. 7:838-842.
- Bentley, D. L. 2005. Rules of engagement: Co-transcriptional recruitment of pre-mRNA processing factors. Curr. Opin. Cell Biol. 17:251–256.
- Aguilera, A. 2005. Cotranscriptional mRNP assembly: From the DNA to the nuclear pore. Curr. Opin. Cell Biol. 17:242–250.
- Ro-Choi, T. S. 1999. Nuclear snRNA and nuclear function (discovery of 5' cap structures in RNA). Crit. Rev. Eukaryotic Gene Expr. 9:107–158.
- Bard, J., Zhelkovsky, A. M., Helmling, S., Earnest, T. N., Moore, C. L., and Bohm, A. 2000. Structure of yeast poly(A) polymerase alone and in complex with 3'-dATP. *Science* 289:1346–1349.
- Martin, G., Keller, W., and Doublie, S. 2000. Crystal structure of mammalian poly(A) polymerase in complex with an analog of ATP. *EMBO J.* 19:4193–4203.
- Zhao, J., Hyman, L., and Moore, C. 1999. Formation of mRNA 3' ends in eukaryotes: Mechanism, regulation, and interrelationships with other steps in mRNA synthesis. *Microbiol. Mol. Biol. Rev.* 63:405–445.
- Minvielle-Sebastia, L., and Keller, W. 1999. mRNA polyadenylation and its coupling to other RNA processing reactions and to transcription. *Curr. Opin. Cell Biol.* 11:352–357.

Small Regulatory RNAs

- Winter, J., Jung, S., Keller, S., Gregory, R. I., and Diederichs, S. 2009. Many roads to maturity: MicroRNA biogenesis pathways and their regulation. *Nat. Cell Biol.* 11:228–234.
- Ruvkun, G., Wightman, B., and Ha, I. 2004. The 20 years it took to recognize the importance of tiny RNAs. *Cell* 116:S93–S96.

B 3 2

SELECTED READINGS

RNA Editing

- Gott, J. M., and Emeson, R. B. 2000. Functions and mechanisms of RNA editing. Annu. Rev. Genet. 34:499–531.
- Simpson, L., Thiemann, O. H., Savill, N. J., Alfonzo, J. D., and Maslov, D. A. 2000. Evolution of RNA editing in trypanosome mitochondria. Proc. Natl. Acad. Sci. U.S.A. 97:6986–6993.
- Chester, A., Scott, J., Anant, S., and Navaratnam, N. 2000. RNA editing: Cytidine to uridine conversion in apolipoprotein B mRNA. *Biochim. Biophys. Acta* 1494:1–3.
- Maas, S., and Rich, A. 2000. Changing genetic information through RNA editing. *Bioessays* 22:790–802.

Splicing of mRNA Precursors

- Caceres, J. F., and Kornblihtt, A. R. 2002. Alternative splicing: Multiple control mechanisms and involvement in human disease. *Trends Genet*. 18:186–193.
- Faustino, N. A., and Cooper, T. A. 2003. Pre-mRNA splicing and human disease. *Genes Dev.* 17:419–437.
- Lou, H., and Gagel, R. F. 1998. Alternative RNA processing: Its role in regulating expression of calcitonin/calcitonin gene-related peptide. J. Endocrinol. 156:401–405.
- Matlin, A. J., Clark, F., and Smith, C. W. 2005. Understanding alternative splicing: Towards a cellular code. Nat. Rev. Mol. Cell Biol. 6:386–398.
- McKie, A. B., McHale, J. C., Keen, T. J., Tarttelin, E. E., Goliath, R., et al. 2001. Mutations in the pre-mRNA splicing factor gene PRPC8 in autosomal dominant retinitis pigmentosa (RP13). *Hum. Mol. Genet.* 10:1555–1562.
- Nilsen, T. W. 2003. The spliceosome: The most complex macromolecular machine in the cell? *Bioessays* 25:1147–1149.
- Rund, D., and Rachmilewitz, E. 2005. β-Thalassemia. N. Engl. J. Med. 353:1135–1146.
- Patel, A. A., and Steitz, J. A. 2003. Splicing double: Insights from the second spliceosome. Nat. Rev. Mol. Cell Biol. 4:960–970.
- Sharp, P. A. 2005. The discovery of split genes and RNA splicing. Trends Biochem. Sci. 30:279–281.
- Valadkhan, S., and Manley, J. L. 2001. Splicing-related catalysis by protein-free snRNAs. *Nature* 413:701–707.
- Zhou, Z., Licklider, L. J., Gygi, S. P., and Reed, R. 2002. Comprehensive proteomic analysis of the human spliceosome. *Nature* 419:182–185.
- Stark, H., Dube, P., Luhrmann, R., and Kastner, B. 2001. Arrangement of RNA and proteins in the spliceosomal U1 small nuclear ribonucleoprotein particle. *Nature* 409:539–542.
- Strehler, E. E., and Zacharias, D. A. 2001. Role of alternative splicing in generating isoform diversity among plasma membrane calcium pumps. *Physiol. Rev.* 81:21–50.
- Graveley, B. R. 2001. Alternative splicing: Increasing diversity in the proteomic world. *Trends Genet.* 17:100–107.
- Newman, A. 1998. RNA splicing. Curr. Biol. 8:R903-R905.
- Reed, R. 2000. Mechanisms of fidelity in pre-mRNA splicing. Curr. Opin. Cell Biol. 12:340–345.
- Sleeman, J. E., and Lamond, A. I. 1999. Nuclear organization of premRNA splicing factors. *Curr. Opin. Cell Biol.* 11:372–377.
- Black, D. L. 2000. Protein diversity from alternative splicing: A challenge for bioinformatics and post-genome biology. *Cell* 103:367–370.
- Collins, C. A., and Guthrie, C. 2000. The question remains: Is the spliceosome a ribozyme? *Nat. Struct. Biol.* 7:850–854.

Self-Splicing and RNA Catalysis

- Adams, P. L., Stanley, M. R., Kosek, A. B., Wang, J., and Strobel, S. A. 2004. Crystal structure of a self-splicing group I intron with both exons. *Nature* 430:45–50.
- Adams, P. L., Stanley, M. R., Gill, M. L., Kosek, A. B., Wang, J., and Strobel, S. A. 2004. Crystal structure of a group I intron splicing intermediate. *RNA* 10:1867–1887.
- Stahley, M. R., and Strobel, S. A. 2005. Structural evidence for a two-metalion mechanism of group I intron splicing. *Science* 309:1587–1590.

- Carola, C., and Eckstein, F. 1999. Nucleic acid enzymes. Curr. Opin. Chem. Biol. 3:274–283.
- Doherty, E. A., and Doudna, J. A. 2000. Ribozyme structures and mechanisms. Annu. Rev. Biochem. 69:597–615.
- Fedor, M. J. 2000. Structure and function of the hairpin ribozyme. J. Mol. Biol. 297:269–291.
- Hanna, R., and Doudna, J. A. 2000. Metal ions in ribozyme folding and catalysis. *Curr. Opin. Chem. Biol.* 4:166–170.
- Scott, W. G. 1998. RNA catalysis. Curr. Opin. Struct. Biol. 8:720-726.

Chapter 30

Where to Start

- Williamson, J. R. 2009. The ribosome at atomic resolution. Cell 139:1041–1043.
- Noller, H. F. 2005. RNA structure: Reading the ribosome. Science 309:1508–1514.
- Dahlberg, A. E. 2001. Ribosome structure: The ribosome in action. Science 292:868–869.
- Ibba, M., Curnow, A. W., and Söll, D. 1997. Aminoacyl-tRNA synthesis: Divergent routes to a common goal. *Trends Biochem.* Sci. 22:39-42.
- Koonin, E. V., and Novozhilov, A. S. 2009. Origin and evolution of the genetic code: The universal enigma. *IUBMB Life* 61:99–111.
- Schimmel, P., and Ribas de Pouplana, L. 2000. Footprints of aminoacyl-tRNA synthetases are everywhere. *Trends Biochem. Sci.* 25:207–209.

Books

- Cold Spring Harbor Symposia on Quantitative Biology. 2001. Volume 66, *The Ribosome*. Cold Spring Harbor Laboratory Press.
- Gesteland, R. F., Atkins, J. F., and Cech, T. (Eds.). 2005. *The RNA World*, 3d ed. Cold Spring Harbor Laboratory Press.
- Garrett, R., Douthwaite, S. R., Liljas, A., Matheson, A. T, Moore, P. B., and Noller, H. F. 2000. The Ribosome: Structure, Function, Antibiotics, and Cellular Interactions. The American Society for Microbiology.

Aminoacyl-tRNA Synthetases

- Kaminska, M., Havrylenko, S., Decottignies, P., Le Maréchal, P., Negrutskii, B., and Mirande, M. 2009. Dynamic organization of aminoacyl-tRNA synthetase complexes in the cytoplasm of human cells. J. Biol. Chem. 284:13746–13754.
- Park, S. G., Schimmel, P., and Kim, S. 2008. Aminoacyl tRNA synthetases and their connections to disease. Proc. Natl. Acad. Sci. U.S.A. 105:11043–11049.
- Ibba, M., and Söll, D. 2000. Aminoacyl-tRNA synthesis. Annu. Rev. Biochem. 69:617–650.
- Sankaranarayanan, R., Dock-Bregeon, A. C., Rees, B., Bovee, M., Caillet, J., Romby, P., Francklyn, C. S., and Moras, D. 2000. Zinc ion mediated amino acid discrimination by threonyl-tRNA synthetase. *Nat. Struct. Biol.* 7:461–465.
- Sankaranarayanan, R., Dock-Bregeon, A. C., Romby, P., Caillet, J., Springer, M., Rees, B., Ehresmann, C., Ehresmann, B., and Moras, D. 1999. The structure of threonyl-tRNA synthetase-tRNA^{Thr} complex enlightens its repressor activity and reveals an essential zinc ion in the active site. *Cell* 97:371–381.
- Dock-Bregeon, A., Sankaranarayanan, R., Romby, P., Caillet, J., Springer, M., Rees, B., Francklyn, C. S., Ehresmann, C., and Moras, D. 2000. Transfer RNA-mediated editing in threonyltRNA synthetase: The class II solution to the double discrimination problem. *Cell* 103:877–884.
- de Pouplana, L. R., and Schimmel, P. 2000. A view into the origin of life: Aminoacyl-tRNA synthetases. *Cell. Mol. Life Sci.* 57:865–870.

Transfer RNA

- Ibba, M., Becker, H. D., Stathopoulos, C., Tumbula, D. L., and Söll, D. 2000. The adaptor hypothesis revisited. *Trends Biochem. Sci.* 25:311–316.
- Weisblum, B. 1999. Back to Camelot: Defining the specific role of tRNA in protein synthesis. *Trends Biochem. Sci.* 24:247–250.

Ribosomes and Ribosomal RNAs

- Jin, H., Kelley, A. C., Loakes, D., and Ramakrishnan, V. 2010. Structure of the 70S ribosome bound to release factor 2 and a substrate analog provides insights into catalysis of peptide release. *Proc. Natl. Acad. Sci. U.S.A.* 107:8593–8598.
- Rodnina, M. V., and Wintermeyer, W. 2009. Recent mechanistic insights into eukaryotic ribosomes. Curr. Opin. Cell Biol. 21:435–443.
- Dinman, J. D. 2008. The eukaryotic ribosome: Current status and challenges. J. Biol. Chem. 284:11761–11765.
- Wen, J.-D., Lancaster, L., Hodges, C., Zeri, A.-C., Yoshimura, S. H., Noller, H. F., Bustamante, C., and Tinoco, I., Jr. 2008. Following translation by single ribosomes one codon at a time. *Nature* 452:598–603.
- Korostelev, A., and Noller, H. F. 2007. The ribosome in focus: New structures bring insights. *Trends Biochem. Sci.* 32:434–441.
- Brandt, F., Etchells, S. A., Ortiz, J. O., Elcock, A. H., Hartl, F. U., and Baumeister, W. 2009. The native 3D organization of bacterial polysomes. *Cell* 136:261–271.
- Schuwirth, B. S., Borovinskaya, M. A., Hau, C. W., Zhang, W., Vila-Sanjurjo, A., Holton, J. M., and Cate, J. H. 2005. Structures of the bacterial ribosome at 3.5 Å resolution. *Science* 310:827–834.
- Yonath, A., and Franceschi, F. 1998. Functional universality and evolutionary diversity: Insights from the structure of the ribosome. *Structure* 6:679–684.
- Ban, N., Nissen, P., Hansen, J., Moore, P. B., and Steitz, T. A. 2000. The complete atomic structure of the large ribosomal subunit at 2.4 Å resolution. Science 289:905–920.
- Carter, A. P., Clemons, W. M., Brodersen, D. E., Morgan-Warren, R. J., Wimberly, B. T., and Ramakrishnan, V. 2000. Functional insights from the structure of the 30S ribosomal subunit and its interactions with antibiotics. *Nature* 407:340–348.
- Wimberly, B. T., Brodersen, D. E., Clemons, W. M., Morgan-Warren, R. J., Carter, A. P., Vonrhein, C., Hartsch, T., and Ramakrishnan, V. 2000. Structure of the 30S ribosomal subunit. *Nature* 407:327–339.

Initiation Factors

- Søgaard, B., Sørensen, H. P., Mortensen, K. K., and Sperling-Petersen, H. U. 2005. Initiation of protein synthesis in bacteria. *Microbiol. Mol. Biol. Rev.* 69:101–123.
- Carter, A. P., Clemons, W. M., Jr., Brodersen, D. E., Morgan-Warren, R. J., Hartsch, T., Wimberly, B. T., and Ramakrishnan, V. 2001. Crystal structure of an initiation factor bound to the 30S ribosomal subunit. Science 291:498–501.
- Guenneugues, M., Caserta, E., Brandi, L., Spurio, R., Meunier, S., Pon, C. L., Boelens, R., and Gualerzi, C. O. 2000. Mapping the fMet-tRNA_f^{Met} binding site of initiation factor IF2. *EMBO J.* 19:5233–5240.
- Meunier, S., Spurio, R., Czisch, M., Wechselberger, R., Guenneugues, M., Gualerzi, C. O., and Boelens, R. 2000. Structure of the fMettRNA_f^{Met}-binding domain of *B. stearothermophilus* initiation factor IF2. *EMBO J.* 19:1918–1926.

Elongation Factors

- Schuette, J.-C., Murphy, F.V., IV, Kelley, A. C., Weir, J. R., Giesebrecht, J., Connell, S. R., Loerke, J., Mielke, T., Zhang, W., Penczek, P. A., Ramakrishnan, V., and Spahn, C. M. T. 2009. GTPase activation of elongation factor EF-Tu by the ribosome during decoding. *EMBO* J. 28:755–765.
- Stark, H., Rodnina, M. V., Wieden, H. J., van Heel, M., and Wintermeyer, W. 2000. Large-scale movement of elongation factor

G and extensive conformational change of the ribosome during translocation. *Cell* 100:301–309.

- Baensch, M., Frank, R., and Kohl, J. 1998. Conservation of the aminoterminal epitope of elongation factor Tu in Eubacteria and Archaea. *Microbiology* 144:2241–2246.
- Krasny, L., Mesters, J. R., Tieleman, L. N., Kraal, B., Fucik, V., Hilgenfeld, R., and Jonak, J. 1998. Structure and expression of elongation factor Tu from *Bacillus stearothermophilus*. J. Mol. Biol. 283:371–381.
- Pape, T., Wintermeyer, W., and Rodnina, M. V. 1998. Complete kinetic mechanism of elongation factor Tu-dependent binding of aminoacyl-tRNA to the A site of the *E. coli* ribosome. *EMBO J.* 17:7490–7497.
- Piepenburg, O., Pape, T., Pleiss, J. A., Wintermeyer, W., Uhlenbeck, O. C., and Rodnina, M. V. 2000. Intact aminoacyl-tRNA is required to trigger GTP hydrolysis by elongation factor Tu on the ribosome. *Biochemistry* 39:1734–1738.

Peptide-Bond Formation and Translocation

- Uemura, S., Aitken, C. E., Korlach, J., Flusberg, B. A., Turner, S. W., and Puglisi, J. D. 2010. Real-time tRNA transit on single translating ribosomes at codon resolution. *Nature* 464:1012–1018.
- Beringer, M., and. Rodnina, M. V. 2007. The ribosomal peptidyl transferase. *Mol. Cell* 26:311–321.
- Yarus, M., and Welch, M. 2000. Peptidyl transferase: Ancient and exiguous. *Chem. Biol.* 7:R187–R190.
- Vladimirov, S. N., Druzina, Z., Wang, R., and Cooperman, B. S. 2000. Identification of 50S components neighboring 23S rRNA nucleotides A2448 and U2604 within the peptidyl transferase center of *Escherichia coli* ribosomes. *Biochemistry* 39:183–193.
- Frank, J., and Agrawal, R. K. 2000. A ratchet-like inter-subunit reorganization of the ribosome during translocation. *Nature* 406:318–322.

Termination

- Weixlbaumer, A., Jin, H., Neubauer, C., Voorhees, R. M., Petry, S., Kelley, A. C., and Ramakrishnan, V. 2008. Insights into translational termination from the structure of RF2 bound to the ribosome. *Science* 322:953–956.
- Trobro, S., and Aqvist, S. 2007. A model for how ribosomal release factors induce peptidyl-tRNA cleavage in termination of protein synthesis. *Mol. Cell* 27:758–766.
- Korosteleva, A., Asaharaa, H., Lancastera, L., Laurberga, M., Hirschia, A., Zhua, J., Trakhanova, S., Scotta, W. G., and Noller, H. F. 2008. Crystal structure of a translation termination complex formed with release factor RF2. *Proc. Natl. Acad. Sci. U.S.A.* 105:19684– 19689.
- Wilson, D. N., Schluenzen, F., Harms, J. M., Yoshida, T., Ohkubo, T., Albrecht, A., Buerger, J., Kobayashi, Y., and Fucini, P. 2005. X-ray crystallography study on ribosome recycling: The mechanism of binding and action of RRF on the 50S ribosomal subunit. *EMBO J.* 24:251–260.
- Kisselev, L. L., and Buckingham, R. H. 2000. Translational termination comes of age. *Trends Biochem. Sci.* 25:561–566.

Fidelity and Proofreading

- Zaher, H. S., and Green, R. 2009. Quality control by the ribosome following peptide bond formation. *Nature* 457:161–166.
- Zaher, H. S., and Green, R. 2009. Fidelity at the molecular level: Lessons from protein synthesis. *Cell* 136:746–762.
- Ogle, J. M., and Ramakrishnan, V. 2005. Structural insights into translational fidelity. Annu. Rev. Biochem. 74:129–177.
- Ibba, M., and Söll, D. 1999. Quality control mechanisms during translation. Science 286:1893–1897.
- Rodnina, M. V., and Wintermeyer, W. 2001. Ribosome fidelity: tRNA discrimination, proofreading and induced fit. *Trends Biochem. Sci.* 26:124–130.

B34

SELECTED READINGS

Eukaryotic Protein Synthesis

- Rhoads, R. E. 2009. eIF4E: New family members, new binding partners, new roles. J. Biol. Chem. 284:16711–16715.
- Marintchev, A., Edmonds, K. A., Marintcheva, B., Hendrickson, E., Oberer, M., Suzuki, C., Herdy, B., Sonenberg, N., and Wagner, G. 2009. Topology and regulation of the human eIF4A/4G/4H helicase complex in translation initiation. *Cell* 136:447–460.
- Fitzgerald, K. D., and Semler, B. L. 2009. Bridging IRES elements in mRNAs to the eukaryotic translation apparatus. *Biochim. Biophys. Acta* 1789:518–528.
- Mitchell, S. F., and Lorsch, J. R. 2008. Should I stay or should I go? Eukaryotic translation initiation factors 1 and 1A control start codon recognition. J. Biol. Chem. 283:27345–27349.
- Amrani, A., Ghosh, S., Mangus, D. A., and Jacobson, A. 2008. Translation factors promote the formation of two states of the closed-loop mRNP. *Nature* 453:1276–1280.
- Sachs, A. B., and Varani, G. 2000. Eukaryotic translation initiation: There are (at least) two sides to every story. *Nat. Struct. Biol.* 7:356–361.
- Kozak, M. 1999. Initiation of translation in prokaryotes and eukaryotes. Gene 234:187–208.
- Bushell, M., Wood, W., Clemens, M. J., and Morley, S. J. 2000. Changes in integrity and association of eukaryotic protein synthesis initiation factors during apoptosis. *Eur. J. Biochem.* 267:1083–1091.
- Das, S., Ghosh, R., and Maitra, U. 2001. Eukaryotic translation initiation factor 5 functions as a GTPase-activating protein. J. Biol. Chem. 276:6720-6726.
- Lee, J. H., Choi, S. K., Roll-Mecak, A., Burley, S. K., and Dever, T. E. 1999. Universal conservation in translation initiation revealed by human and archaeal homologs of bacterial translation initiation factor IF2. Proc. Natl. Acad. Sci. U.S.A. 96:4342–4347.
- Pestova, T. V., and Hellen, C. U. 2000. The structure and function of initiation factors in eukaryotic protein synthesis. *Cell. Mol. Life Sci.* 57:651–674.

Antibiotics and Toxins

- Belova, L., Tenson, T., Xiong, L., McNicholas, P. M., and Mankin, A. S. 2001. A novel site of antibiotic action in the ribosome: Interaction of evernimicin with the large ribosomal subunit. *Proc. Natl. Acad. Sci. U.S.A.* 98:3726–3731.
- Brodersen, D. E., Clemons, W. M., Jr., Carter, A. P., Morgan-Warren, R. J., Wimberly, B. T., and Ramakrishnan, V. 2000. The structural basis for the action of the antibiotics tetracycline, pactamycin, and hygromycin B on the 30S ribosomal subunit. *Cell* 103:1143–1154.
- Porse, B. T., and Garrett, R. A. 1999. Ribosomal mechanics, antibiotics, and GTP hydrolysis. *Cell* 97:423–426.
- Lord, M. J., Jolliffe, N. A., Marsden, C. J., Pateman, C. S., Smith, D. S., Spooner, R. A., Watson, P. D., and Roberts, L. M. 2003. Ricin: Mechanisms of toxicity. *Toxicol. Rev.* 22:53–64.

Protein Transport Across Membranes

- Janda, C. Y., Li, J., Oubridge, C., Hernández, H., Robinson, C. V., and Nagai, K. 2010. Recognition of a signal peptide by the signal recognition particle. *Nature* 465:507–510.
- Cross, B. C. S., Sinning, I., Luirink, J., and High, S. 2009. Delivering proteins for export from the cytosol. Nat. Rev. Mol. Cell. Biol. 10:255–264.
- Shan, S., Schmid, S. L., and Zhang, X. 2009. Signal recognition particle (SRP) and SRP receptor: A new paradigm for multistate regulatory GTPases. *Biochemistry* 48:6696–6704.
- Johnson, A. E. 2009. The structural and functional coupling of two molecular machines, the ribosome and the translocon. J. Cell Biol. 185:765–767.
- Pool, R. P. 2009. A trans-membrane segment inside the ribosome exit tunnel triggers RAMP4 recruitment to the Sec61p translocase. J. Cell Biol. 185:889–902.

- Egea, P. F., Stroud, R. M., and Walter, P. 2005. Targeting proteins to membranes: Structure of the signal recognition particle. *Curr. Opin. Struct. Biol.* 15:213–220.
- Halic, M., and Beckmann, R. 2005. The signal recognition particle and its interactions during protein targeting. *Curr. Opin. Struct. Biol*.15:116–125.
- Doudna, J. A., and Batey, R. T. 2004. Structural insights into the signal recognition particle. Annu. Rev. Biochem. 73:539–557.
- Schnell, D. J., and Hebert, D. N. 2003. Protein translocons: Multifunctional mediators of protein translocation across membranes. *Cell* 112:491–505.

Chapter 31

Where to Start

- Pabo, C. O., and Sauer, R. T. 1984. Protein–DNA recognition. Annu. Rev. Biochem. 53:293–321.
- Ptashne, M., Johnson, A. D., and Pabo, C. O. 1982. A genetic switch in a bacterial virus. Sci. Am. 247:128–140.
- Ptashne, M., Jeffrey, A., Johnson, A. D., Maurer, R., Meyer, B. J., Pabo, C. O., Roberts, T. M., and Sauer, R. T. 1980. How the lambda repressor and Cro work. *Cell* 19:1–11.

Books

- Ptashne, M. 2004. A Genetic Switch: Phage λ Revisited (3d ed.). Cold Spring Harbor Laboratory Press.
- McKnight, S. L., and Yamamoto, K. R. (Eds.). 1992. Transcriptional Regulation (vols. 1 and 2). Cold Spring Harbor Laboratory Press.
- Lodish, H., Berk, A., Matsudaira, P., Kaiser, C. A., Krieger, M., Scott, M. P., Zipursky, S. L., and Darnell, J., 2008. *Molecular Cell Biology* (6th ed.). W. H. Freeman and Company.

DNA-Binding Proteins

- Balaeff, A., Mahadevan, L. and Schulten, K. 2004. Structural basis for cooperative DNA binding by CAP and *lac* repressor. *Structure* 12:123–132.
- Bell, C. E., and Lewis, M. 2001. The Lac repressor: A second generation of structural and functional studies. *Curr. Opin. Struct. Biol.* 11:19–25.
- Lewis, M., Chang, G., Horton, N. C., Kercher, M. A., Pace, H. C., Schumacher, M. A., Brennan, R. G., and Lu, P. 1996. Crystal structure of the lactose operon repressor and its complexes with DNA and inducer. *Science* 271:1247–1254.
- Niu, W., Kim, Y., Tau, G., Heyduk, T., and Ebright, R. H. 1996. Transcription activation at class II CAP-dependent promoters: Two interactions between CAP and RNA polymerase. *Cell* 87:1123–1134.
- Schultz, S. C., Shields, G. C., and Steitz, T. A. 1991. Crystal structure of a CAP-DNA complex: The DNA is bent by 90 degrees. *Science* 253:1001–1007.
- Parkinson, G., Wilson, C., Gunasekera, A., Ebright, Y. W., Ebright, R. E., and Berman, H. M. 1996. Structure of the CAP-DNA complex at 2.5 Å resolution: A complete picture of the protein–DNA interface. J. Mol. Biol. 260:395–408.
- Busby, S., and Ebright, R. H. 1999. Transcription activation by catabolite activator protein (CAP). J. Mol. Biol. 293:199–213.
- Somers, W. S., and Phillips, S. E. 1992. Crystal structure of the met repressor-operator complex at 2.8 Å resolution reveals DNA recognition by β-strands. *Nature* 359:387–393.

Gene-Regulatory Circuits

- Johnson, A. D., Poteete, A. R., Lauer, G., Sauer, R. T., Ackers, G. K., and Ptashne, M. 1981. Lambda repressor and Cro: Components of an efficient molecular switch. *Nature* 294:217–223.
- Stayrook, S., Jaru-Ampornpan, P., Ni, J., Hochschild, A., and Lewis, M. 2008. Crystal structure of the lambda repressor and a model for pairwise cooperative operator binding. *Nature* 452:1022–1025.

Arkin, A., Ross, J., and McAdams, H. H. 1998. Stochastic kinetic analysis of developmental pathway bifurcation in phage lambdainfected *Escherichia coli* cells. *Genetics* 149:1633–1648.

Posttranscriptional Regulation

- Kolter, R., and Yanofsky, C. 1982. Attenuation in amino acid biosynthetic operons. Annu. Rev. Genet. 16:113–134.
- Yanofsky, C. 1981. Attenuation in the control of expression of bacterial operons. Nature 289:751–758.
- Miller, M. B., and Bassler, B. L. 2001. Quorum sensing in bacteria. Annu. Rev. Microbiol. 55:165–199.
- Zhang, R. G., Pappas, T., Brace, J. L., Miller, P. C., Oulmassov, T., Molyneaux, J. M., Anderson, J. C., Bashkin, J. K., Winans, S. C., and Joachimiak, A. 2002. Structure of a bacterial quorum-sensing transcription factor complexed with pheromone and DNA. *Nature* 417:971–974.
- Soberon-Chavez, G., Aguirre-Ramirez, M., and Ordonez, L. 2005. Is Pseudomonas aeruginosa only "sensing quorum"? Crit. Rev. Microbiol. 31:171–182.

Historical Aspects

- Lewis, M. 2005. The lac repressor. C. R. Biol. 328:521-548.
- Jacob, F., and Monod, J. 1961. Genetic regulatory mechanisms in the synthesis of proteins. J. Mol. Biol. 3:318–356.
- Ptashne, M., and Gilbert, W. 1970. Genetic repressors. Sci. Am. 222(6):36-44.
- Lwoff, A., and Ullmann, A. (Eds.). 1979. Origins of Molecular Biology: A Tribute to Jacques Monod. Academic Press.
- Judson, H. 1996. The Eighth Day of Creation: Makers of the Revolution in Biology. Cold Spring Harbor Laboratory Press.

Chapter 32

Where to Start

- Kornberg, R. D. 2007. The molecular basis of eukaryotic transcription. Proc. Natl. Acad. Sci. U.S.A. 104:12955–12961.
- Pabo, C. O., and Sauer, R. T. 1984. Protein–DNA recognition. Annu. Rev. Biochem. 53:293–321.
- Struhl, K. 1989. Helix-turn-helix, zinc-finger, and leucine-zipper motifs for eukaryotic transcriptional regulatory proteins. *Trends Biochem. Sci.* 14:137–140.
- Struhl, K. 1999. Fundamentally different logic of gene regulation in eukaryotes and prokaryotes. *Cell* 98:1–4.
- Korzus, E., Torchia, J., Rose, D. W., Xu, L., Kurokawa, R., McInerney, E. M., Mullen, T. M., Glass, C. K., and Rosenfeld, M. G. 1998. Transcription factor-specific requirements for coactivators and their acetyltransferase functions. *Science* 279:703–707.
- Aalfs, J. D., and Kingston, R. E. 2000. What does "chromatin remodeling" mean? Trends Biochem. Sci. 25:548-555.

Books

- McKnight, S. L., and Yamamoto, K. R. (Eds.). 1992. Transcriptional Regulation (vols. 1 and 2). Cold Spring Harbor Laboratory Press.
- Latchman, D. S. 2004. *Eukaryotic Transcription Factors* (4th ed.). Academic Press.
- Wolffe, A. 1992. Chromatin Structure and Function. Academic Press.
- Lodish, H., Berk, A., Matsudaira, P., Kaiser, C. A., Krieger, M., Scott, M. P., Zipursky, S. L., and Darnell, J., 2008. *Molecular Cell Biology* (6th ed.). W. H. Freeman and Company.

Chromatin and Chromatin Remodeling

- Lorch, Y., Maier-Davis, B., and Kornberg, R. D. 2010. Mechanism of chromatin remodeling. *Proc. Natl. Acad. Sci. U.S.A.* 107:3458–3462.
- Tang, L., Nogales, E., and Ciferri, C. 2010. Structure and function of SWI/SNF chromatin remodeling complexes and mechanistic

implications for transcription. Prog. Biophys. Mol. Biol. 102: 122-128.

- Jenuwein, T., and Allis, C. D. 2001. Translating the histone code. Science 293:1074–1080.
- Jiang, C., and Pugh, B. F. 2009. Nucleosome positioning and gene regulation: Advances through genomics. Nat. Rev. Genet. 10:161–172.
- Barski, A., Cuddapah, S., Cui, K., Roh, T. Y., Schones, D. E., Wang, Z., Wei, G., Chepelev, I., and Zhao, K. 2007. High-resolution profiling of histone methylations in the human genome. *Cell* 129:823–837.
- Weintraub, H., Larsen, A., and Groudine, M. 1981. β-Globin-gene switching during the development of chicken embryos: Expression and chromosome structure. *Cell* 24:333–344.
- Ren, B., Robert, F., Wyrick, J. J., Aparicio, O., Jennings, E. G., Simon, I., Zeitlinger, J., Schreiber, J., Hannett, N., Kanin, E., Volkert, T. L., Wilson, C. J., Bell, S. P., and Young, R. A. 2000. Genome-wide location and function of DNA-binding proteins. *Science* 290:2306–2309.
- Goodrich, J. A., and Tjian, R. 1994. TBP-TAF complexes: Selectivity factors for eukaryotic transcription. *Curr. Opin. Cell. Biol.* 6:403–409.
- Bird, A. P., and Wolffe, A. P. 1999. Methylation-induced repression: Belts, braces, and chromatin. *Cell* 99:451–454.
- Cairns, B. R. 1998. Chromatin remodeling machines: Similar motors, ulterior motives. *Trends Biochem. Sci.* 23:20–25.
- Albright, S. R., and Tjian, R. 2000. TAFs revisited: More data reveal new twists and confirm old ideas. *Gene* 242:1–13.
- Urnov, F. D., and Wolffe, A. P. 2001. Chromatin remodeling and transcriptional activation: The cast (in order of appearance). Oncogene 20:2991–3006.
- Luger, K., Mader, A. W., Richmond, R. K., Sargent, D. F., and Richmond, T. J. 1997. Crystal structure of the nucleosome core particle at 2.8 Å resolution. *Nature* 389:251–260.
- Arents, G., and Moudrianakis, E. N. 1995. The histone fold: A ubiquitous architectural motif utilized in DNA compaction and protein dimerization. *Proc. Natl. Acad. Sci. U.S.A.* 92:11170–11174.
- Baxevanis, A. D., Arents, G., Moudrianakis, E. N., and Landsman, D. 1995. A variety of DNA-binding and multimeric proteins contain the histone fold motif. *Nucleic Acids Res.* 23:2685–2691.

Transcription Factors

- Green, M. R. 2005. Eukaryotic transcription activation: Right on target. Mol. Cell 18:399–402.
- Kornberg, R. D. 2005. Mediator and the mechanism of transcriptional activation. Trends Biochem. Sci. 30:235–239.
- Clements, A., Rojas, J. R., Trievel, R. C., Wang, L., Berger, S. L., and Marmorstein, R. 1999. Crystal structure of the histone acetyltransferase domain of the human PCAF transcriptional regulator bound to coenzyme A. *EMBO J.* 18:3521–3532.
- Deckert, J., and Struhl, K. 2001. Histone acetylation at promoters is differentially affected by specific activators and repressors. *Mol. Cell. Biol.* 21:2726–2735.
- Dutnall, R. N., Tafrov, S. T., Sternglanz, R., and Ramakrishnan, V. 1998. Structure of the histone acetyltransferase Hat1: A paradigm for the GCN5-related N-acetyltransferase superfamily. *Cell* 94:427–438.
- Finnin, M. S., Donigian, J. R., Cohen, A., Richon, V. M., Rifkind, R. A., Marks, P. A., Breslow, R., and Pavletich, N. P. 1999. Structures of a histone deacetylase homologue bound to the TSA and SAHA inhibitors. *Nature* 401:188–193.
- Finnin, M. S., Donigian, J. R., and Pavletich, N. P. 2001. Structure of the histone deacetylase SIR2. Nat. Struct. Biol. 8:621–625.
- Jacobson, R. H., Ladurner, A. G., King, D. S., and Tjian, R. 2000. Structure and function of a human TAFII250 double bromodomain module. *Science* 288:1422–1425.
- Rojas, J. R., Trievel, R. C., Zhou, J., Mo, Y., Li, X., Berger, S. L., Allis, C. D., and Marmorstein, R. 1999. Structure of *Tetrahymena* GCN5 bound to coenzyme A and a histone H3 peptide. *Nature* 401:93–98.

B36

SELECTED READINGS

Induced Pluripotent Stem Cells

- Takahashi, K., Tanabe, K., Ohnuki, M., Narita, M., Ichisaka, T., Tomoda, K., and Yamanaka, S. 2007. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell* 131:861–872.
- Takahashi, K., and Yamanaka, S. 2006. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 126:663–676.
- Park, I. H., Arora, N., Huo, H., Maherali, N., Ahfeldt, T., Shimamura, A., Lensch, M. W., Cowan, C., Hochedlinger, K., and Daley, G. Q. 2008. Disease-specific induced pluripotent stem cells. *Cell* 134:877–886.

Yamanaka, S. 2009. A fresh look at iPS cells. Cell 137:13-17.

Yu, J., Hu, K., Smuga-Otto, K., Tian, S., Stewart, R., Slukvin, I. I., and Thomson, J. A. 2009. Human induced pluripotent stem cells free of vector and transgene sequences. *Science* 324:797–801.

Nuclear Hormone Receptors

- Downes, M., Verdecia, M. A., Roecker, A. J., Hughes, R., Hogenesch, J. B., Kast-Woelbern, H. R., Bowman, M. E., Ferrer, J. L., Anisfeld, A. M., Edwards, P. A., Rosenfeld, J. M., Alvarez, J. G., Noel, J. P., Nicolaou, K. C., and Evans, R. M. 2003. A chemical, genetic, and structural analysis of the nuclear bile acid receptor FXR. *Mol. Cell* 11:1079–1092.
- Evans, R. M. 2005. The nuclear receptor superfamily: A Rosetta stone for physiology. *Mol. Endocrinol.* 19:1429–1438.
- Xu, W., Cho, H., Kadam, S., Banayo, E. M., Anderson, S., Yates, J. R., 3d, Emerson, B. M., and Evans, R. M. 2004. A methylationmediator complex in hormone signaling. *Genes Dev.* 18:144–156.
- Evans, R. M. 1988. The steroid and thyroid hormone receptor superfamily. Science 240:889–895.
- Yamamoto, K. R. 1985. Steroid receptor regulated transcription of specific genes and gene networks. Annu. Rev. Genet. 19:209–252.
- Tanenbaum, D. M., Wang, Y., Williams, S. P., and Sigler, P. B. 1998. Crystallographic comparison of the estrogen and progesterone receptor's ligand binding domains. *Proc. Natl. Acad. Sci. U.S.A.* 95:5998–6003.
- Schwabe, J. W., Chapman, L., Finch, J. T., and Rhodes, D. 1993. The crystal structure of the estrogen receptor DNA-binding domain bound to DNA: How receptors discriminate between their response elements. *Cell* 75:567–578.
- Shiau, A. K., Barstad, D., Loria, P. M., Cheng, L., Kushner, P. J., Agard, D. A., and Greene, G. L. 1998. The structural basis of estrogen receptor/coactivator recognition and the antagonism of this interaction by tamoxifen. *Cell* 95:927–937.
- Collingwood, T. N., Urnov, F. D., and Wolffe, A. P. 1999. Nuclear receptors: Coactivators, corepressors and chromatin remodeling in the control of transcription. J. Mol. Endocrinol. 23:255–275.

Posttranscriptional Regulation

- Rouault, T. A., Stout, C. D., Kaptain, S., Harford, J. B., and Klausner, R. D. 1991. Structural relationship between an iron-regulated RNA-binding protein (IRE-BP) and aconitase: Functional implications. *Cell* 64:881–883.
- Klausner, R. D., Rouault, T. A., and Harford, J. B. 1993. Regulating the fate of mRNA: The control of cellular iron metabolism. *Cell* 72:19–28.
- Gruer, M. J., Artymiuk, P. J., and Guest, J. R. 1997. The aconitase family: Three structural variations on a common theme. *Trends Biochem. Sci.* 22:3–6.
- Theil, E. C. 1994. Iron regulatory elements (IREs): A family of mRNA non-coding sequences. *Biochem. J.* 304:1–11.

MicroRNAs

- Ruvkun, G. 2008. The perfect storm of tiny RNAs. Nat. Med. 14:1041–1045.
- Sethupathy, P., and Collins, F. S. 2008. MicroRNA target site polymorphisms and human disease. *Trends Genet*. 24:489–497.

- Adams, B. D., Cowee, D. M., and White, B. A. 2009. The role of miR-206 in the epidermal growth factor (EGF) induced repression of estrogen receptor- α (ER α) signaling and a luminal phenotype in MCF-7 breast cancer cells. *Mol. Endocrinol.* 23:1215–1230.
- Jegga, A. G., Chen, J., Gowrisankar, S., Deshmukh, M. A., Gudivada, R., Kong, S., Kaimal, V., and Aronow, B. J. 2007. GenomeTrafac: A whole genome resource for the detection of transcription factor binding site clusters associated with conventional and microRNA encoding genes conserved between mouse and human gene orthologs. *Nucleic Acids Res.* 35:D116–D121.

Chapter 33

Where to Start

- Axel, R. 1995. The molecular logic of smell. Sci. Am. 273(4):154–159.
- Dulac, C. 2000. The physiology of taste, vintage 2000. Cell 100:607-610.
- Yarmolinsky, D. A., Zuker, C. S., and Ryba, N. J. (2009) Common sense about taste: From mammals to insects. *Cell* 139:234–244.
- Stryer, L. 1996. Vision: From photon to perception. Proc. Natl. Acad. Sci. U.S.A. 93:557–559.
- Hudspeth, A. J. 1989. How the ear's works work. Nature 341:397-404.

Olfaction

- Buck, L., and Axel, R.1991. A novel multigene family may encode odorant receptors: A molecular basis for odor recognition. *Cell* 65:175–187.
- Saito, H., Chi, Q., Zhuang, H., Matsunami, H., and Mainland, J. D. 2009. Odor coding by a mammalian receptor repertoire. Sci. Signal. 2:ra9.
- Malnic, B., Hirono, J., Sato, T., and Buck, L. B. 1999. Combinatorial receptor codes for odors. *Cell* 96:713–723.
- Zou, D. J., Chesler, A., and Firestein, S. 2009. How the olfactory bulb got its glomeruli: A just so story? Nat. Rev. Neurosci. 10:611-618.
- De la Cruz, O., Blekhman, R., Zhang, X., Nicolae, D., Firestein, S., and Gilad, Y. 2009. A signature of evolutionary constraint on a subset of ectopically expressed olfactory receptor genes. *Mol. Biol. Evol.* 26:491–494.
- Mombaerts, P., Wang, F., Dulac, C., Chao, S. K., Nemes, A., Mendelsohn, M., Edmondson, J., and Axel, R. 1996. Visualizing an olfactory sensory map. *Cell* 87:675–686.
- Buck, L. 2005. Unraveling the sense of smell (Nobel lecture). Angew. Chem. Int. Ed. Engl. 44:6128–6140.
- Belluscio, L., Gold, G. H., Nemes, A., and Axel, R. 1998. Mice deficient in G_(olf) are anosmic. *Neuron* 20:69–81.
- Vosshall, L. B., Wong, A. M., and Axel, R. 2000. An olfactory sensory map in the fly brain. *Cell* 102:147–159.
- Lewcock, J. W., and Reed, R. R. 2003. A feedback mechanism regulates monoallelic odorant receptor expression. Proc. Natl. Acad. Sci. U.S.A.101:1069–1074.
- Reed, R. R. 2004. After the holy grail: Establishing a molecular mechanism for mammalian olfaction. *Cell* 116:329–336.

Taste

- Chandrashekar, J., Yarmolinsky, D., von Buchholtz, L., Oka, Y., Sly, W., Ryba, N. J., and Zuker, C. S. 2009. The taste of carbonation. *Science* 326:443–445.
- Zhao, G. Q., Zhang, Y., Hoon, M. A., Chandrashekar, J., Erlenbach, I., Ryba, N. J. P., and Zuker, C. S. 2003. The receptors for mammalian sweet and umami taste. *Cell* 115:255–266.
- Herness, M. S., and Gilbertson, T. A. 1999. Cellular mechanisms of taste transduction. Annu. Rev. Physiol. 61:873–900.
- Adler, E., Hoon, M. A., Mueller, K. L., Chandrashekar, J., Ryba, N. J., and Zuker, C. S. 2000. A novel family of mammalian taste receptors. *Cell* 100:693–702.

- Chandrashekar, J., Mueller, K. L., Hoon, M. A., Adler, E., Feng, L., Guo, W., Zuker, C. S., and Ryba, N. J. 2000. T2Rs function as bitter taste receptors. *Cell* 100:703–711.
- Mano, I., and Driscoll, M. 1999. DEG/ENaC channels: A touchy superfamily that watches its salt. *Bioessays* 21:568–578.
- Benos, D. J., and Stanton, B. A. 1999. Functional domains within the degenerin/epithelial sodium channel (Deg/ENaC) superfamily of ion channels. J. Physiol. (Lond.) 520(part 3):631–644.
- McLaughlin, S. K., McKinnon, P. J., and Margolskee, R. F. 1992. Gustducin is a taste-cell-specific G protein closely related to the transducins. *Nature* 357:563–569.
- Nelson, G., Hoon, M. A., Chandrashekar, J., Zhang, Y., Ryba, N. J., and Zuker, C. S. 2001. Mammalian sweet taste receptors. *Cell* 106:381–390.

Vision

- Stryer, L. 1988. Molecular basis of visual excitation. Cold Spring Harbor Symp. Quant. Biol. 53:283–294.
- Jastrzebska, B., Tsybovsky, Y., and Palczewski, K. 2010. Complexes between photoactivated rhodopsin and transducin: Progress and questions. *Biochem. J.* 428:1–10.
- Wald, G. 1968. The molecular basis of visual excitation. *Nature* 219:800–807.
- Ames, J. B., Dizhoor, A. M., Ikura, M., Palczewski, K., and Stryer, L. 1999. Three-dimensional structure of guanylyl cyclase activating protein-2, a calcium-sensitive modulator of photoreceptor guanylyl cyclases. J. Biol. Chem. 274:19329–19337.
- Nathans, J. 1994. In the eye of the beholder: Visual pigments and inherited variation in human vision. *Cell* 78:357–360.
- Nathans, J. 1999. The evolution and physiology of human color vision: Insights from molecular genetic studies of visual pigments. *Neuron* 24:299–312.
- Palczewski, K., Kumasaka, T., Hori, T., Behnke, C. A., Motoshima, H., Fox, B. A., LeTrong, I., Teller, D. C., Okada, T., Stenkamp, R. E., Yamamoto, M., and Miyano, M. 2000. Crystal structure of rhodopsin: A G protein-coupled receptor. *Science* 289:739–745.
- Filipek, S, Teller, D. C., Palczewski, K., and Stemkamp, R. 2003. The crystallographic model of rhodopsin and its use in studies of other G protein-coupled receptors. *Annu. Rev. Biophys. Biomol. Struct.* 32:375–397.

Hearing

- Furness, D. N., Hackney, C. M., and Evans, M. G. 2010. Localisation of the mechanotransducer channels in mammalian cochlear hair cells provides clues to their gating. J. Physiol. 588:765–772.
- Lim, K., and Park, S. 2009. A mechanical model of the gating spring mechanism of stereocilia. J. Biomech. 42:2158–2164.
- Siemens, J., Lillo, C., Dumont, R. A., Reynolds, A., Williams, D. S., Gillespie, P. G., and Muller, U. 2004. Cadherin 23 is a component of the tip link in hair-cell stereocilia. *Nature* 428:950–955.
- Spinelli, K. J., and Gillespie, P. G. 2009. Bottoms up: Transduction channels at tip link bases. Nat. Neurosci. 12:529–530.
- Hudspeth, A. J. 1997. How hearing happens. Neuron 19:947-950.
- Pickles, J. O., and Corey, D. P. 1992. Mechanoelectrical transduction by hair cells. *Trends Neurosci*. 15:254–259.
- Walker, R. G., Willingham, A. T., and Zuker, C. S. 2000. A Drosophila mechanosensory transduction channel. Science 287:2229–2234.
- Hudspeth, A. J., Choe, Y., Mehta, A. D., and Martin, P. 2000. Putting ion channels to work: Mechanoelectrical transduction, adaptation, and amplification by hair cells. *Proc. Natl. Acad. Sci. U.S.A.* 97:11765–11772.

Touch and Pain Reception

Myers, B. R., Bohlen, C. J., and Julius, D. 2008. A yeast genetic screen reveals a critical role for the pore helix domain in TRP channel gating. *Neuron* 58:362–373.

- Lishko, P. V., Procko, E., Jin, X., Phelps, C. B., and Gaudet, R. 2007. The ankyrin repeats of TRPV1 bind multiple ligands and modulate channel sensitivity. *Neuron* 54:905–918.
- Franco-Obregon, A., and Clapham, D. E. 1998. Touch channels sense blood pressure. Neuron 21:1224–1226.
- Caterina, M. J., Schumacher, M. A., Tominaga, M., Rosen, T. A., Levine, J. D., and Julius, D. 1997. The capsaicin receptor: A heatactivated ion channel in the pain pathway. *Nature* 389:816–824.
- Tominaga, M., Caterina, M. J., Malmberg, A. B., Rosen, T. A., Gilbert, H., Skinner, K., Raumann, B. E., Basbaum, A. I., and Julius, D. 1998. The cloned capsaicin receptor integrates multiple painproducing stimuli. *Neuron* 21:531–543.
- Caterina, M. J., and Julius, D. 1999. Sense and specificity: A molecular identity for nociceptors. *Curr. Opin. Neurobiol.* 9:525–530.
- Clapham, D. E. 2003. TRP channels as cellular sensors. *Nature* 426:517–524.

Chapter 34

Where to Start

- Nossal, G. J. V. 1993. Life, death, and the immune system. Sci. Am. 269(3):53-62.
- Tonegawa, S. 1985. The molecules of the immune system. Sci. Am. 253(4):122-131.
- Leder, P. 1982. The genetics of antibody diversity. Sci. Am. 246(5):102-115.
- Bromley, S. K., Burack, W. R., Johnson, K. G., Somersalo, K., Sims, T. N., Sumen, C., Davis, M. M., Shaw, A. S., Allen, P. M., and Dustin, M. L. 2001. The immunological synapse. *Annu. Rev. Immunol.* 19:375–396.

Books

- Kindt, T. J., Goldsby, R. A., and Osborne, B. A. 2007. Kuby Immunology (6th ed.). W. H. Freeman and Company.
- Abbas, A. K., and Lichtman, A. H. 2003. Cellular and Molecular Immunology (5th ed). Saunders.
- Cold Spring Harbor Symposia on Quantitative Biology, 1989. Volume 54. Immunological Recognition.
- Nisinoff, A. 1985. Introduction to Molecular Immunology (2d ed.). Sinauer.
- Weir, D. M. (Ed.). 1996. Handbook of Experimental Immunology (5th ed.). Oxford University Press.
- Janeway, C. A., Travers, P., Walport, M., and Shlomchik, M. 2005. *Immunobiology* (6th ed.). Garland Science.

Innate Immune System

- Janeway, C. A., Jr., and Medzhitov, R. 2002. Innate immune recognition. Annu. Rev. Immunol. 20:197–216.
- Choe, J., Kelker, M. S., and Wilson, I. A. 2005. Crystal structure of human toll-like receptor 3 (TLR3) ectodomain. *Science* 309:581–585.
- Khalturin, K., Panzer, Z., Cooper, M. D., and Bosch, T. C. 2004. Recognition strategies in the innate immune system of ancestral chordates. *Mol. Immunol.* 41:1077–1087.
- Beutler, B., and Rietschel, E. T. 2003. Innate immune sensing and its roots: The story of endotoxin. Nat. Rev. Immunol. 3:169–176.
- Xu, Y., Tao, X., Shen, B., Horng, T., Medzhitov, R., Manley, J. L., and Tong, L. 2000. Structural basis for signal transduction by the Toll/ interleukin-1 receptor domains. *Nature* 408:111–115.

Structure of Antibodies and Antibody-Antigen Complexes

- Davies, D. R., Padlan, E. A., and Sheriff, S. 1990. Antibody-antigen complexes. Annu. Rev. Biochem. 59:439–473.
- Poljak, R. J. 1991. Structure of antibodies and their complexes with antigens. Mol. Immunol. 28:1341–1345.
- Davies, D. R., and Cohen, G. H. 1996. Interactions of protein antigens with antibodies. Proc. Natl. Acad. Sci. U.S.A. 93:7–12.

B 3 8

SELECTED READINGS

- Marquart, M., Deisenhofer, J., Huber, R., and Palm, W. 1980. Crystallographic refinement and atomic models of the intact immunoglobulin molecule Kol and its antigen-binding fragment at 3.0 Å and 1.9 Å resolution. J. Mol. Biol. 141:369–391.
- Silverton, E. W., Navia, M. A., and Davies, D. R. 1977. Threedimensional structure of an intact human immunoglobulin. *Proc. Natl. Acad. Sci. U.S.A.* 74:5140–5144.
- Padlan, E. A., Silverton, E. W., Sheriff, S., Cohen, G. H., Smith, G. S., and Davies, D. R. 1989. Structure of an antibody-antigen complex: Crystal structure of the HyHEL-10 Fab lysozyme complex. *Proc. Natl. Acad. Sci. U.S.A.* 86:5938–5942.
- Rini, J., Schultze-Gahmen, U., and Wilson, I. A. 1992. Structural evidence for induced fit as a mechanism for antibody-antigen recognition. *Science* 255:959–965.
- Fischmann, T. O., Bentley, G. A., Bhat, T. N., Boulot, G., Mariuzza, R. A., Phillips, S. E., Tello, D., and Poljak, R. J. 1991. Crystallographic refinement of the three-dimensional structure of the FabD1.3-lysozyme complex at 2.5-Å resolution. J. Biol. Chem. 266:12915–12920.
- Burton, D. R. 1990. Antibody: The flexible adaptor molecule. Trends Biochem. Sci. 15:64–69.
- Saphire, E. O., Parren P. W., Pantophlet, R., Zwick, M. B., Morris, G. M., Rudd, P. M., Dwek, R. A., Stanfield, R. L., Burton, D. R., and Wilson, I. A. 2001. Crystal structure of a neutralizing human IgG against HIV-1: A template for vaccine design. *Science* 293:1155–1159.
- Calarese, D. A., Scanlan, C. N., Zwick, M. B., Deechongkit, S., Mimura, Y., Kunert R., Zhu, P., Wormald, M. R., Stanfield, R. L., Roux, K. H., Kelly, J. W., Rudd, P. M., Dwek, R. A., Katinger, H., Burton, D. R., and Wilson, I. A. 2003. Antibody domain exchange is an immunological solution to carbohydrate cluster recognition. *Science* 300:2065–2071.

Generation of Diversity

- Tonegawa, S. 1988. Somatic generation of immune diversity. Biosci. Rep. 8:3–26.
- Honjo, T., and Habu, S. 1985. Origin of immune diversity: Genetic variation and selection. Annu. Rev. Biochem. 54:803–830.
- Gellert, M., and McBlane, J. F. 1995. Steps along the pathway of VDJ recombination. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 347:43–47.
- Harris, R. S., Kong, Q., and Maizels, N. 1999. Somatic hypermutation and the three R's: Repair, replication and recombination. *Mutat. Res.* 436:157–178.
- Lewis, S. M., and Wu, G. E. 1997. The origins of V(D)J recombination. *Cell* 88:159–162.
- Ramsden, D. A., van Gent, D. C., and Gellert, M. 1997. Specificity in V(D)J recombination: New lessons from biochemistry and genetics. *Curr. Opin. Immunol.* 9:114–120.
- Roth, D. B., and Craig, N. L. 1998. VDJ recombination: A transposase goes to work. *Cell* 94:411–414.
- Sadofsky, M. J. 2001. The RAG proteins in V(D)J recombination: More than just a nuclease. Nucleic Acids Res. 29:1399–1409.

MHC Proteins and Antigen Processing

- Bjorkman, P. J., and Parham, P. 1990. Structure, function, and diversity of class I major histocompatibility complex molecules. *Annu. Rev. Biochem.* 59:253–288.
- Goldberg, A. L., and Rock, K. L. 1992. Proteolysis, proteasomes, and antigen presentation. *Nature* 357:375–379.
- Madden, D. R., Gorga, J. C., Strominger, J. L., and Wiley, D. C. 1992. The three-dimensional structure of HLA-B27 at 2.1 Å resolution suggests a general mechanism for tight binding to MHC. *Cell* 70:1035–1048.
- Fremont, D. H., Matsumura, M., Stura, E. A., Peterson, P. A., and Wilson, I. A. 1992. Crystal structures of two viral peptides in complex with murine MHC class I H-2Kb. *Science* 257:880–881.

- Matsumura, M., Fremont, D. H., Peterson, P. A., and Wilson, I. A. 1992. Emerging principles for the recognition of peptide antigens by MHC class I. Science 257:927–934.
- Brown, J. H., Jardetzky, T. S., Gorga, J. C., Stern, L. J., Urban, R. G., Strominger, J. L., and Wiley, D. C. 1993. Three-dimensional structure of the human class II histocompatibility antigen HLA-DR1. *Nature* 364:33–39.
- Saper, M. A., Bjorkman, P. J., and Wiley, D. C. 1991. Refined structure of the human histocompatibility antigen HLA-A2 at 2.6 Å resolution. J. Mol. Biol. 219:277–319.
- Madden, D. R., Gorga, J. C., Strominger, J. L., and Wiley, D. C. 1991. The structure of HLA-B27 reveals nonamer self-peptides bound in an extended conformation. *Nature* 353:321–325.
- Cresswell, P., Bangia, N., Dick, T., and Diedrich, G. 1999. The nature of the MHC class I peptide loading complex. *Immunol. Rev.* 172:21–28.
- Madden, D. R., Garboczi, D. N., and Wiley, D. C. 1993. The antigenic identity of peptide-MHC complexes: A comparison of the conformations of five viral peptides presented by HLA-A2. *Cell* 75: 693–708.

T-Cell Receptors and Signaling Complexes

- Hennecke, J., and Wiley, D. C. 2001. T-cell receptor-MHC interactions up close. Cell 104:1–4.
- Ding, Y. H., Smith, K. J., Garboczi, D. N., Utz, U., Biddison, W. E., and Wiley, D. C. 1998. Two human T cell receptors bind in a similar diagonal mode to the HLA-A2/Tax peptide complex using different TCR amino acids. *Immunity* 8:403–411.
- Reinherz, E. L., Tan, K., Tang, L., Kern, P., Liu, J., Xiong, Y., Hussey, R. E., Smolyar, A., Hare, B., Zhang, R., Joachimiak, A., Chang, H. C., Wagner, G., and Wang, J. 1999. The crystal structure of a T-cell receptor in complex with peptide and MHC class II. *Science* 286:1913–1921.
- Davis, M. M., and Bjorkman, P. J. 1988. T-cell antigen receptor genes and T-cell recognition. *Nature* 334:395–402.
- Cochran, J. R., Cameron, T. O., and Stern, L. J. 2000. The relationship of MHC-peptide binding and T cell activation probed using chemically defined MHC class II oligomers. *Immunity* 12:241–250.
- Garcia, K. C., Teyton, L., and Wilson, I. A. 1999. Structural basis of T cell recognition. Annu. Rev. Immunol. 17:369–397.
- Garcia, K. C., Degano, M., Stanfield, R. L., Brunmark, A., Jackson, M. R., Peterson, P. A., Teyton, L. A., and Wilson, I. A. 1996. An αβ T-cell receptor structure at 2.5 Å and its orientation in the TCR-MHC complex. *Science* 274:209–219.
- Garboczi, D. N., Ghosh, P., Utz, U., Fan, Q. R., Biddison, W. E., Wiley, D. C. 1996. Structure of the complex between human T-cell receptor, viral peptide and HLA-A2. *Nature* 384:134–141.
- Gaul, B. S., Harrison, M. L., Geahlen, R. L., Burton, R. A., and Post, C. B. 2000. Substrate recognition by the Lyn protein-tyrosine kinase: NMR structure of the immunoreceptor tyrosine-based activation motif signaling region of the B cell antigen receptor. J. Biol. Chem. 275:16174–16182.
- Kern, P. S., Teng, M. K., Smolyar, A., Liu, J. H., Liu, J., Hussey, R. E., Spoerl, R., Chang, H. C., Reinherz, E. L., and Wang, J. H. 1998. Structural basis of CD8 coreceptor function revealed by crystallographic analysis of a murine CD8 αβ ectodomain fragment in complex with H-2Kb. *Immunity* 9:519–530.
- Konig, R., Fleury, S., and Germain, R. N. 1996. The structural basis of CD4-MHC class II interactions: Coreceptor contributions to T cell receptor antigen recognition and oligomerization-dependent signal transduction. *Curr. Top. Microbiol. Immunol.* 205:19–46.
- Davis, M. M., Boniface, J. J., Reich, Z., Lyons, D., Hampl, J., Arden, B., and Chien, Y. 1998. Ligand recognition by αβ T-cell receptors. *Annu. Rev. Immunol.* 16:523–544.
- Janeway, C. J. 1992. The T cell receptor as a multicomponent signalling machine: CD4/CD8 coreceptors and CD45 in T cell activation. Annu. Rev. Immunol. 10:645–674.

- Podack, E. R., and Kupfer, A. 1991. T-cell effector functions: Mechanisms for delivery of cytotoxicity and help. Annu. Rev. Cell Biol. 7:479–504.
- Davis, M. M. 1990. T cell receptor gene diversity and selection. Annu. Rev. Biochem. 59:475–496.
- Leahy, D. J., Axel, R., and Hendrickson, W. A. 1992. Crystal structure of a soluble form of the human T cell coreceptor CD8 at 2.6 Å resolution. *Cell* 68:1145–1162.
- Bots, M., and Medema, J. P. 2006. Granzymes at a glance. J. Cell. Sci. 119:5011–5014.
- Lowin, B., Hahne, M., Mattmann, C., and Tschopp, J. 1994. Cytolytic T-cell cytotoxicity is mediated through perform and Fas lytic pathways. *Nature* 370:650–652.
- Rudolph, M. G., and Wilson, I. A. 2002. The specificity of TCR/ pMHC interaction. Curr. Opin. Immunol. 14:52–65.

HIV and AIDS

- Fauci, A. S. 1988. The human immunodeficiency virus: Infectivity and mechanisms of pathogenesis. Science 239:617–622.
- Gallo, R. C., and Montagnier, L. 1988. AIDS in 1988. Sci. Am. 259(4):41-48.
- Kwong, P. D., Wyatt, R., Robinson, J., Sweet, R. W., Sodroski, J., and Hendrickson, W. A. 1998. Structure of an HIV gp120 envelope glycoprotein in complex with the CD4 receptor and a neutralizing human antibody. *Nature* 393:648–659.

Vaccines

- Johnston, M. I. and Fauci, A. S. 2007. An HIV vaccine—evolving concepts. N. Engl. J. Med. 356:2073–2081.
- Burton, D. R., Desrosiers, R. C., Doms, R. W., Koff, W. C., Kwong, P. D., Moore, J. P., Nabel, G. J., Sodroski, J., Wilson, I. A., and Wyatt, R. T. 2004. HIV vaccine design and the neutralizing antibody problem. *Nature Immunol.* 5:233–236.
- Ada, G. 2001. Vaccines and vaccination. N. Engl. J. Med. 345:1042-1053.
- Behbehani, A. M. 1983. The smallpox story: Life and death of an old disease. *Microbiol. Rev.* 47:455–509.

Discovery of Major Concepts

- Ada, G. L., and Nossal, G. 1987. The clonal selection theory. Sci. Am. 257(2):62–69.
- Porter, R. R. 1973. Structural studies of immunoglobulins. Science 180:713–716.
- Edelman, G. M. 1973. Antibody structure and molecular immunology. Science 180:830–840.
- Kohler, G. 1986. Derivation and diversification of monoclonal antibodies. Science 233:1281–1286.
- Milstein, C. 1986. From antibody structure to immunological diversification of immune response. Science 231:1261–1268.
- Janeway, C. A., Jr. 1989. Approaching the asymptote? Evolution and revolution in immunology. Cold Spring Harbor Symp. Quant. Biol. 54:1-13.
- Jerne, N. K. 1971. Somatic generation of immune recognition. Eur. J. Immunol. 1:1–9.

Chapter 35

Where to Start

- Gennerich, A., and Vale, R. D. 2009. Walking the walk: How kinesin and dynein coordinate their steps. Curr. Opin. Cell Biol. 21:59–67.
- Vale, R. D. 2003. The molecular motor toolbox for intracellular transport. Cell 112:467–480.
- Vale, R. D., and Milligan, R. A. 2000. The way things move: Looking under the hood of molecular motor proteins. *Science* 288:88–95.
- Vale, R. D. 1996. Switches, latches, and amplifiers: Common themes of G proteins and molecular motors. J. Cell Biol. 135:291–302.

- Mehta, A. D., Rief, M., Spudich, J. A., Smith, D. A., and Simmons, R. M. 1999. Single-molecule biomechanics with optical methods. *Science* 283:1689–1695.
- Schuster, S. C., and Khan, S. 1994. The bacterial flagellar motor. Annu. Rev. Biophys. Biomol. Struct. 23:509–539.

Books

- Howard, J. 2001. Mechanics of Motor Proteins and the Cytosketon. Sinauer.
- Squire, J. M. 1986. Muscle Design, Diversity, and Disease. Benjamin Cummings.
- Pollack, G. H., and Sugi, H. (Eds.). 1984. Contractile Mechanisms in Muscle. Plenum.

Myosin and Actin

- Lorenz, M., and Holmes, K. C. 2010. The actin-myosin interface. Proc. Natl. Acad. Sci. U.S.A. 107:12529–12534.
- Yang, Y., Gourinath, S., Kovacs, M., Nyitray, L., Reutzel, R., Himmel, D. M., O'Neall-Hennessey, E., Reshetnikova, L., Szent-Györgyi, A. G., Brown, J. H., and Cohen, C. 2007. Rigor-like structures from muscle myosins reveal key mechanical elements in the transduction pathways of this allosteric motor. *Structure* 15:553–564.
- Himmel, D. M., Mui, S., O'Neall-Hennessey, E., Szent-Györgyi, A. G., and Cohen, C. 2009. The on-off switch in regulated myosins: Different triggers but related mechanisms. J. Mol. Biol. 394:496-505.
- Houdusse, A., Gaucher, J. F., Krementsova, E., Mui, S., Trybus, K. M., and Cohen, C. 2006. Crystal structure of apo-calmodulin bound to the first two IQ motifs of myosin V reveals essential recognition features. *Proc. Natl. Acad. Sci. U.S.A.* 103:19326–19331.
- Li, X. E., Holmes, K. C., Lehman, W., Jung, H., and Fischer, S. 2010. The shape and flexibility of tropomyosin coiled coils: Implications for actin filament assembly and regulation. J. Mol. Biol. 395:327–339.
- Fischer, S., Windshugel, B., Horak, D., Holmes, K. C., and Smith, J. C. 2005. Structural mechanism of the recovery stroke in the myosin molecular motor. *Proc. Natl. Acad. Sci. U.S.A.* 102:6873–6878.
- Holmes, K. C., Angert, I., Kull, F. J., Jahn, W., and Schroder, R. R. 2003. Electron cryo-microscopy shows how strong binding of myosin to actin releases nucleotide. *Nature* 425:423–427.
- Holmes, K. C., Schroder, R. R., Sweeney, H. L., and Houdusse, A. 2004. The structure of the rigor complex and its implications for the power stroke. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 359:1819–1828.
- Purcell, T. J., Morris, C., Spudich, J. A., and Sweeney, H. L. 2002. Role of the lever arm in the processive stepping of myosin V. Proc. Natl. Acad. Sci. U.S.A. 99:14159–14164.
- Purcell, T. J., Sweeney, H. L., and Spudich, J. A. 2005. A forcedependent state controls the coordination of processive myosin V. Proc. Natl. Acad. Sci. U.S.A. 102:13873–13878.
- Holmes, K. C. 1997. The swinging lever-arm hypothesis of muscle contraction. Curr. Biol. 7:R112–R118.
- Berg, J. S., Powell, B. C., and Cheney, R. E. 2001. A millennial myosin census. *Mol. Biol. Cell* 12:780–794.
- Houdusse, A., Kalabokis, V. N., Himmel, D., Szent-Györgyi, A. G., and Cohen, C. 1999. Atomic structure of scallop myosin subfragment S1 complexed with MgADP: A novel conformation of the myosin head. *Cell* 97:459–470.
- Houdusse, A., Szent-Györgyi, A. G., and Cohen, C. 2000. Three conformational states of scallop myosin S1. Proc. Natl. Acad. Sci. U.S.A. 97:11238–11243.
- Uyeda, T. Q., Abramson, P. D., and Spudich, J. A. 1996. The neck region of the myosin motor domain acts as a lever arm to generate movement. *Proc. Natl. Acad. Sci. U.S.A.* 93:4459–4464.
- Mehta, A. D., Rock, R. S., Rief, M., Spudich, J. A., Mooseker, M. S., and Cheney, R. E. 1999. Myosin-V is a processive actin-based motor. *Nature* 400:590–593.
- Otterbein, L. R., Graceffa, P., and Dominguez, R. 2001. The crystal structure of uncomplexed actin in the ADP state. *Science* 293:708–711.

SELECTED READINGS

- Holmes, K. C., Popp, D., Gebhard, W., and Kabsch, W. 1990. Atomic model of the actin filament. *Nature* 347:44–49.
- Schutt, C. E., Myslik, J. C., Rozycki, M. D., Goonesekere, N. C., and Lindberg, U. 1993. The structure of crystalline profilin-β-actin. *Nature* 365:810–816.
- van den Ent, F., Amos, L. A., and Lowe, J. 2001. Prokaryotic origin of the actin cytoskeleton. *Nature* 413:39–44.
- Schutt, C. E., and Lindberg, U. 1998. Muscle contraction as a Markov process I: Energetics of the process. Acta Physiol. Scand. 163:307–323.
- Rief, M., Rock, R. S., Mehta, A. D., Mooseker, M. S., Cheney, R. E., and Spudich, J. A. 2000. Myosin-V stepping kinetics: A molecular model for processivity. *Proc. Natl. Acad. Sci. U.S.A.* 97:9482–9486.
- Friedman, T. B., Sellers, J. R., and Avraham, K. B. 1999. Unconventional myosins and the genetics of hearing loss. Am. J. Med. Genet. 89:147–157.

Kinesin, Dynein, and Microtubules

- Yildiz, A., Tomishige, M., Gennerich, A., and Vale, R. D. 2008. Intramolecular strain coordinates kinesin stepping behavior along microtubules. *Cell* 134:1030–1041.
- Yildiz, A., Tomishige, M., Vale, R. D., and Selvin, P. R. 2004. Kinesin walks hand-over-hand. Science 303:676–678.
- Rogers, G. C., Rogers, S. L., Schwimmer, T. A., Ems-McClung, S. C., Walczak, C. E., Vale, R. D., Scholey, J. M., and Sharp, D. J. 2004. Two mitotic kinesins cooperate to drive sister chromatid separation during anaphase. *Nature* 427:364–370.
- Vale, R. D., and Fletterick, R. J. 1997. The design plan of kinesin motors. Annu. Rev. Cell. Dev. Biol. 13:745–777.
- Kull, F. J., Sablin, E. P., Lau, R., Fletterick, R. J., and Vale, R. D. 1996. Crystal structure of the kinesin motor domain reveals a structural similarity to myosin. *Nature* 380:550–555.
- Kikkawa, M., Sablin, E. P., Okada, Y., Yajima, H., Fletterick, R. J., and Hirokawa, N. 2001. Switch-based mechanism of kinesin motors. *Nature* 411:439–445.
- Wade, R. H., and Kozielski, F. 2000. Structural links to kinesin directionality and movement. Nat. Struct. Biol. 7:456–460.
- Yun, M., Zhang, X., Park, C. G., Park, H. W., and Endow, S. A. 2001. A structural pathway for activation of the kinesin motor ATPase. *EMBO J.* 20:2611–2618.
- Kozielski, F., De Bonis, S., Burmeister, W. P., Cohen-Addad, C., and Wade, R. H. 1999. The crystal structure of the minus-end-directed microtubule motor protein ncd reveals variable dimer conformations. Struct. Fold. Des. 7:1407–1416.
- Lowe, J., Li, H., Downing, K. H., and Nogales, E. 2001. Refined structure of αβ-tubulin at 3.5 Å resolution. *J. Mol. Biol.* 313:1045–1057.
- Nogales, E., Downing, K. H., Amos, L. A., and Lowe, J. 1998. Tubulin and FtsZ form a distinct family of GTPases. *Nat. Struct. Biol.* 5:451–458.
- Zhao, C., Takita, J., Tanaka, Y., Setou, M., Nakagawa, T., Takeda, S., Yang, H. W., Terada, S., Nakata, T., Takei, Y., Saito, M., Tsuji, S., Hayashi, Y., and Hirokawa, N. 2001. Charcot-Marie-Tooth disease type 2A caused by mutation in a microtubule motor KIF1Bβ. *Cell* 105:587–597.
- Asai, D. J., and Koonce, M. P. 2001. The dynein heavy chain: Structure, mechanics and evolution. *Trends Cell Biol.* 11:196–202.
- Mocz, G., and Gibbons, I. R. 2001. Model for the motor component of dynein heavy chain based on homology to the AAA family of oligomeric ATPases. *Structure* 9:93–103.

Bacterial Motion and Chemotaxis

- Baker, M. D., Wolanin, P. M., and Stock, J. B. 2006. Systems biology of bacterial chemotaxis. Curr. Opin. Microbiol. 9:187–192.
- Wolanin, P. M., Baker, M. D., Francis, N. R., Thomas, D. R., DeRosier, D. J., and Stock, J. B. 2006. Self-assembly of receptor/ signaling complexes in bacterial chemotaxis. *Proc. Natl. Acad. Sci.* U.S.A. 103:14313–14318.

- Sowa, Y., Rowe, A. D., Leake, M. C., Yakushi, T., Homma, M., Ishijima, A., and Berry, R. M. 2005. Direct observation of steps in rotation of the bacterial flagellar motor. *Nature* 437:916–919.
- Berg, H. C. 2000. Constraints on models for the flagellar rotary motor. Philos. Trans. R. Soc. Lond. B Biol. Sci. 355:491–501.
- DeRosier, D. J. 1998. The turn of the screw: The bacterial flagellar motor. *Cell* 93:17–20.
- Ryu, W. S., Berry, R. M., and Berg, H. C. 2000. Torque-generating units of the flagellar motor of *Escherichia coli* have a high duty ratio. *Nature* 403:444–447.
- Lloyd, S. A., Whitby, F. G., Blair, D. F., and Hill, C. P. 1999. Structure of the C-terminal domain of FliG, a component of the rotor in the bacterial flagellar motor. *Nature* 400:472–475.
- Purcell, E. M. 1977. Life at low Reynolds number. Am. J. Physiol. 45:3-11.
- Macnab, R. M., and Parkinson, J. S. 1991. Genetic analysis of the bacterial flagellum. *Trends Genet.* 7:196–200.

Historical Aspects

- Huxley, H. E. 1965. The mechanism of muscular contraction. Sci. Am. 213(6):18–27.
- Summers, K. E., and Gibbons, I. R. 1971. ATP-induced sliding of tubules in trypsin-treated flagella of sea-urchin sperm. Proc. Natl. Acad. Sci. U.S.A. 68:3092–3096.
- Macnab, R. M., and Koshland, D. E., Jr. 1972. The gradient-sensing mechanism in bacterial chemotaxis. Proc. Natl. Acad. Sci. U.S.A. 69:2509–2512.
- Taylor, E. W. 2001. 1999 E. B. Wilson lecture: The cell as molecular machine. *Mol. Biol. Cell* 12:251–254.

Chapter 36

Books

- Kenakin, T. P. 2006. A Pharmacology Primer: Theory, Applications, and Methods (2d ed.). Academic Press.
- Brunton, L., Lazo, J., and Parker, K. 2005. Goodman and Gilman's The Pharmacological Basis of Therapeutics (11th ed.). McGraw-Hill Professional.
- Walsh, C. T., and Schwartz-Bloom, R. D. 2004. Levine's Pharmacology: Drug Actions and Reactions (7th ed.). Taylor and Francis Group.
- Silverman, R. B. 2004. Organic Chemistry of Drug Design and Drug Action. Academic Press.
- Walsh, C. 2003. Antibiotics: Actions, Origins, Resistance. ASM Press.

ADME and Toxicity

- Caldwell, J., Gardner, I., and Swales, N. 1995. An introduction to drug disposition: The basic principles of absorption, distribution, metabolism, and excretion. *Toxicol. Pathol.* 23:102–114.
- Lee, W., and Kim, R. B. 2004. Transporters and renal drug elimination. Annu. Rev. Pharmacol. Toxicol. 44:137–166.
- Lin, J., Sahakian, D. C., de Morais, S. M., Xu, J. J., Polzer, R. J., and Winter, S. M. 2003. The role of absorption, distribution, metabolism, excretion and toxicity in drug discovery. *Curr. Top. Med. Chem.* 3:1125–1154.
- Poggesi, I. 2004. Predicting human pharmacokinetics from preclinical data. Curr. Opin. Drug Discov. Devel. 7:100–111.

Case Histories

- Flower, R. J. 2003. The development of COX2 inhibitors. Nat. Rev. Drug Discov. 2:179–191.
- Tobert, J. A. 2003. Lovastatin and beyond: The history of the HMG-CoA reductase inhibitors. Nat. Rev. Drug Discov. 2:517–526.
- Vacca, J. P., Dorsey, B. D., Schleif, W. A., Levin, R. B., McDaniel, S. L., Darke, P. L., Zugay, J., Quintero, J. C., Blahy, O. M., Roth, E., et al. 1994. L-735,524: An orally bioavailable human immunodeficiency

B41

virus type 1 protease inhibitor. Proc. Natl. Acad. Sci. U.S.A. 91:4096-4100.

Wong, S., and Witte, O. N. 2004. The BCR-ABL story: Bench to bedside and back. Annu. Rev. Immunol. 22:247–306.

Structure-Based Drug Design

- Kuntz, I. D. 1992. Structure-based strategies for drug design and discovery. Science 257:1078–1082.
- Dorsey, B. D., Levin, R. B., McDaniel, S. L., Vacca, J. P., Guare, J. P., Darke, P. L., Zugay, J. A., Emini, E. A., Schleif, W. A., Quintero, J. C., et al. 1994. L-735,524: The design of a potent and orally bioavailable HIV protease inhibitor. J. Med. Chem. 37:3443–3451.
- Chen, Z., Li, Y., Chen, E., Hall, D. L., Darke, P. L., Culberson, C., Shafer, J. A., and Kuo, L. C. 1994. Crystal structure at 1.9-Å resolution of human immunodeficiency virus (HIV) II protease complexed with L-735,524, an orally bioavailable inhibitor of the HIV proteases. J. Biol. Chem. 269:26344–26348.

Combinatorial Chemistry

Baldwin, J. J. 1996. Design, synthesis and use of binary encoded synthetic chemical libraries. *Mol. Divers.* 2:81–88.

- Burke, M. D., Berger, E. M., and Schreiber, S. L. 2003. Generating diverse skeletons of small molecules combinatorially. *Science* 302:613–618.
- Edwards, P. J., and Morrell, A. I. 2002. Solid-phase compound library synthesis in drug design and development. *Curr. Opin. Drug Discov. Devel.* 5:594–605.

Genomics

- Zambrowicz, B. P., and Sands, A. T. 2003. Knockouts model the 100 best-selling drugs: Will they model the next 100? Nat. Rev. Drug Discov. 2:38–51.
- Salemme, F. R. 2003. Chemical genomics as an emerging paradigm for postgenomic drug discovery. *Pharmacogenomics* 4:257–267.
- Michelson, S., and Joho, K. 2000. Drug discovery, drug development and the emerging world of pharmacogenomics: Prospecting for information in a data-rich landscape. *Curr. Opin. Mol. Ther.* 2:651–654.
- Weinshilboum, R., and Wang, L. 2004. Pharmacogenomics: Bench to bedside. Nat. Rev. Drug Discov. 3:739–748.

This page intentionally left blank

Note: Page numbers followed by f, t, and b refer to figures, tables, and boxed material, respectively. **Boldface** page numbers indicate structural formulas and ribbon diagrams.

A band, 1016, 1016f A site, ribosomal, 900, 900f, 902-904 AAA ATPases in amino acid degradation, 677 DnaA as, 834 in dynein, 1010, 1011f Ab initio prediction, of protein structure, 54 ABC transporters, 378-380, 378f, 379f ABO blood groups, glycosyltransferases in. 335 Absorption, drug, 1031–1033, 1032f Abzymes, production of, 244 Acceptor control, 556 Acceptor stem, 891 Accessory pigments, in photosynthesis, 581-584, 582f, 583f Acetaminophen, 1037 hepatic toxicity of, 1036, 1037 Acetate, molecular models of, 23f Acetoacetate, 653, 653-654. See also Ketone bodies in amino acid degradation, 694 from phenylalanine, 696, 696f from tryptophan, 696, 696f Acetoacetyl ACP, 658-659, 658t, 659 Acetoacetyl CoA, 653 Acetyl ACP, 658-659, 658t, 659 Acetyl CoA (acetyl coenzyme A), 440, 440, 497 in amino acid degradation, 694 carboxylation of, 657 in cholesterol synthesis, 767 in citric acid cycle, 497, 500-501, 501, 503-505, 514-515, 653, 656 in ethanol metabolism, 811 in fatty acid metabolism, 646-647, 656 formation of, 468 in fatty acid oxidation, 646-647 from ketone bodies, 633f, 653-655 in glyoxylate cycle, 518 oxidation of, ATP synthesis from, 437-438 transfer from mitochondria to cytoplasm, 662,662f Acetyl CoA carboxylase, 657, 658t, 666-667, 666f Acetyl CoA synthetase, in glyoxylate cycle, 518 Acetylation, 297-298, 298t in gene regulation, 949-951 histone, 298t, 949-950, 950f protein, 57 Acetylcholine, 389 Acetylcholine receptor generation of, 392, 392f as ligand-gated ion channel, 389-390, 391f structure of, 389-390, 390f

Acetylcholinesterase, inhibitors of, 241, 241f N-Acetylglutamate, 686, 688 *N*-Acetyl-L-phenylalanine *p*-nitrophenyl ester, 256, 257f Acetyllysine-binding domain, 949-950, 950f Acid(s). See also Hydrogen ion(s) fatty. See Fatty acid(s) protonation/deprotonation of, 15-16, 15f, 16f sour taste of, 962. 965-966 tactile response to, 974 Acid-base catalysis, 254 Acid-base reactions, 13-17 buffers in, 15-17, 15f, 16f double helix disruption in, 14-15 Aconitase, 952, 953 in citric acid cycle, 506 iron-response element-binding protein as, 952, **953** Acquired immunodeficiency syndrome. See Human immunodeficiency virus infection Actin, decorated, 1014 Actin/actin filaments, 90, 90f, 1012-1018 definition of. 1012 F, 1013, 1013f formation of, 1013-1014, 1013f G. 1013 myosin and, 1016-1017. See also Myosin polymerization of, 1013-1014 structure of, 183, 183-184, 1012-1013, 1013f in thin filaments, 1016 Actin-myosin complex, 1014, 1014f Actinomycin D, 861, 862, 862 Action potentials, 382, 382f equilibrium, 391-392, 391f generation of, 392, 392f ion channels and, 382, 382f, 391-392, 391f Activated carriers, in metabolism, 438–442, 438f, 439f Activated methyl cycle, 717-718, 717f Activating enzymes, in translation, 893-897. See also Aminoacyl-tRNA synthetases Activation energy, 222, 226 Active sites, 185–186, 227–228, 227f, 228f convergent evolution of, 185-186 definition of, 185-186 Active transport, 371, 373. See also Membrane transport Acute intermittent porphyria, 731 Acyl adenylate in asparagine synthesis, 713 in fatty acid metabolism, 644-645, 645 in translation, 893-895

Acyl carnitine, 645, 645-646, 646f Acyl carnitine translocase, 646f Acyl carrier proteins (ACPs), in fatty acid metabolism, 657-658, 658 Acvl CoA, 440, 440, 645 in fatty acid metabolism, 645, 646f Acvl CoA dehydrogenase, 646, 647 Acyl CoA synthetase, 644-645 Adaptive control, 667 Adaptive immunity, 977. See also under Immune Adaptor proteins, in signal transduction, 414, 417 Adenine, 4, 4, 111, 111 deamination of, 838, 838f methylation of, 277-278, 277f, 278f Adenine nucleotide translocase, 552-553, 553f. 554f inhibition of, 558 Adenine phosphoribosyltransferase, 744 Adenosine, 111 degradation of. 752-753, 753f Adenosine 5'-triphosphate. See ATP (adenosine triphosphate) Adenosine diphosphate. See ADP (adenosine diphosphate) Adenosine monophosphate. See AMP (adenosine monophosphate) cyclic. See cAMP S-Adenosylhomocysteine in activated methyl cycle, 717, 717f in amino acid synthesis, 717, 717 S-Adenosylmethionine (SAM), 693, 693 in amino acid synthesis, 716-718 S-Adenosylmethionine, in phospholipid synthesis, 763 Adenvlate cyclase, 474 in signal transduction, 406, 406, 406f Adenvlate kinase P-loop NTPase domains in, 284, 284f structure of, 284 Adenylate, synthesis of, 743-744, 743f Adenylation, in amino acid synthesis, 725-726, 725f Adenylsuccinate synthase, 744 Adenylyl transferase, 725, 726f Adipokines, 796 Adiponectin, 796 Adipose cells (adipocytes), 640f, 641 in appetite regulation, 795–796 metabolism in, in starvation, 809-810 Adipose tissue brown, 556-557, 557f white, 556 Adipose triacylglyceride lipase (ATGL), 643.643f Adler, Julius, 1022

C2

INDEX

ADME properties, of drugs, 1031–1036, 1032f A-DNA helix, 115-116, 116t ADP (adenosine diphosphate) in activated carriers, 447, 447f ATP synthesis from, 430–431, 434, 435, 435f in citric acid cycle, 508, 555-556, 556f energy charge and, 446-447, 446f in kinesin movement, 1021, 1021f in membrane transport, 380 in oxidative phosphorylation, 552, 555-556 in starch synthesis, 597 structure of, 431 ADP ribosylation, 298t ADP-ATP cycle, energy exchange in, 430-431, 435, 435f ADP-VO4³⁻ complex, 1011, 1012f Adrenaline. See Epinephrine Adrenodoxin, 782 Advance glycosylation end products, 326 Aerobic glycolysis, 478-479 Affinity chromatography, 70-71, 70f, 75, 75t, 78 Affinity maturation, 989 Affinity tags, 78, 241–242 Aflatoxin B₁, 838, 838 Aggrecan, 332 Agonists, 948 Agouti-related peptide, 796 Agre, Peter, 394 Agrobacterium tumefaciens, tumor-inducing plasmids in, 166-167, 167f Akt, in signal transduction, 414, 799 Alanine in gluconeogenesis, 490 in glucose-alanine cycle, 684-685, 685f in muscle contraction, 490 in nitrogen transport, 684-685, 685f pyruvate formation from, 691, 691f structure of, 28, 29f synthesis of, 712-713 transamination of, 691, 691f Alanine aminotransferase, 680 Alanyl-tRNA sequence, 889, 890f Albinism, 698t Albumin, in drug distribution, 1033, 1033f Alcaptonuria, 697 Alcohol(s) fermentation of, 453, 466-467, 467f formation from pyruvate, 466-467 formation of from pyruvate, 467f metabolism of, 232, 810-813 monosaccharides and, 326 in phospholipid synthesis, 763 toxicity of, 811-813 Alcohol dehydrogenase, 466-467, 467f, 810-811 in ethanol metabolism, 232 NAD⁺ binding sites in, 469, 469f Alcoholic fermentation, 453 Alcoholic hepatitis, 811-812

Aldehyde(s). See also Monosaccharides oxidation of, in glycolysis, 461, 462f Aldehyde dehydrogenase, 811 in ethanol metabolism, 232 Aldimine, 682, 682 in amino acid degradation, 682, 682f in amino acid synthesis, 712-713, 712f Aldolase in Calvin cycle, 595, 595f in glycolysis, 456f, 458 Aldose(s), 320, 320f in Calvin cycle, 595 in transaldolase reaction, 605, 606f Aldosterone, 780 synthesis of, 780f, 783, 784 Alkali cations, properties of, 386t Allolactose, 926, 926 Allopurinol, 754 Allosteric effector, 204-205 Allosteric enzymes, 237, 237f, 289, 290-296. See also Aspartate transcarbamoylase (ATCase) definition of, 291 kinetics of, 291 heterotropic effects on, 296 homotropic effects on, 294 kinetics of, 291f, 293-295 substrate binding by concerted mechanism in, 294-296 sequential model of, 294 subunits of catalytic. 291-292 interaction of, 292-295, 294f-296f regulatory, 291-292 T-to-R state equilibrium in, 294–296, 294f, 295f, 296f T-to-R state transition in, 294–296, 294f, 295f Allosteric inhibitors, 291, 446 Allosteric interactions, 291, 446 Allosteric sites, 291 α chains, 982, 983f of class I MHC proteins, 992-993, 993f of hemoglobin, 199, 199f, 211 α helix, 38-40, **39, 40,** 40f amino acid residues in, 50–51, 51t destabilization of. 40 of α-keratin, **43**, 43–44, 44f of membrane proteins, 356, 358, 359-360, 360f. 360t of myosin, 1009-1010, 1010f α-hemoglobin, 199, 199f, 211, **211**. See also Hemoglobin α -hemoglobin stabilizing protein, 211, 211 α_1 -antitrypsin, 307 abnormalities of, 312 $\alpha\beta$ dimers, of hemoglobin, 199, 199f, 202, 202f α-tubulin, 1018–1019, 1018f, 1019f Alternative splicing, 133, 133f, 878-879, 878f. See also Splicing Altman, Sidney, 852, 863

Alcohol-related disorders, 811-813

Altrose, 321f Alu sequences, 160 Alzheimer disease, amyloid in, 55-56, 57 α-Amanitin, **865**, 865–866 Ames test, 843, 843f Amethopterin, 750 Amidation, in amino acid synthesis, 713 Amide bonds. See Peptide bonds Amiloride, 965 salty taste and, 965 Amiloride-sensitive sodium ion channels, 965,965f Amines, monosaccharides and, 326 Amino acid(s), 18–19. See also Protein(s) abbreviations for, 32-33, 32t acetyl groups of, 57 alpha, 27 aromatic. 28 degradation of, 695-696 synthesis of, 719-722, 720f, 721f as biomolecular precursors, 726-731 branched-chain, degradation of, 693-695 chirality of, 27, 29f, 713 classification of, 28 D isomer of, 27, 27f definition of, 18 dipolar form of, 27-28, 27f elution volume of, 80, 80f essential, 674, 674t, 711, 711t, 712f synthesis of, 711-712, 719-723. See also Amino acid synthesis evolutionary favorability of, 33 glucogenic, 690 hydrophilic, 30 protein folding and, 46, 47 hydrophobic, 28, 29f protein folding and, 45-46, 46f, 47 hydroxyl groups of, 30, 30f, 57, 57-58 identification of, 80-82, 80f ionization state of, 27-28, 27f ketogenic, 690 L isomer of, 27, 27f metabolic fates of, 673 metabolism of, 437f, 673-698. See also Amino acid degradation; Amino acid synthesis modifications of, 57-59, 57f negatively charged, 31-33 nonessential, 711, 711t, 712f synthesis of, 712-713 peptide bonds of, 33, 33f polar, 30 positively charged, 30-31 properties of, 33 reactive, 30, 33f in secondary structures, 38-45, 50-52, 51t sequences of. See Amino acid sequences side chains of. See Amino acid side chains translational activation of, 893-895 zwitterionic form of, 27-28, 27f Amino acid composition, determination of, 80-82,80f

C3 Index

Amino acid degradation, 437f, 673-700 acetoacetate in, 694 acetyl CoA in, 694 S-adenosylmethionine in, 693, 693 aminotransferases in, 680-682 of aromatic amino acids, 695-696 of branched-chain amino acids, 694-695 carbon skeletons from, metabolic fates of, 690-696, 691f deamination in, 680-684 dehydration in, 684 digestive enzymes in, 255-260, 303-307, 303t, 304f, 674, 674f disorders of, 697-698, 698t glucose-alanine cycle in, 684-685, 685f glutamate dehvdrogenase in, 680-681 glutamate oxidative deamination in, 680-681 homocysteine in, 693, 693 intermediates in, 690-696 isoleucine in, 694-695 α -ketobutyrate in, 693, 693 α -ketoglutarate in, 692 leucine in, 693-694 liver in, 680 in muscle, 685 one-carbon units in, 716 overview of, 673-674 oxaloacetate in, 692 oxygenation in, 695-696 propionyl CoA in, 693, 693, 694 proteasomes in, 677, 677f, 678f in protein turnover, 675 pyruvate in, 684-685, 685f, 691, 691f rate of, 675, 676t regulatory functions of, 678-679, 679t Schiff-base intermediates in, 681-682 serine dehydratase in, 684, 691 in starvation, 809 succinyl coenzyme A in, 693 tetrahydrofolate in, 716 threonine dehydratase in, 684 transamination in, 680-685 ubiguitination in, 298t, 675-678 urea cycle in, 685–690 valine in, 694-695 Amino acid residues, 34 definition of. 34 molecular weight of, 34 in secondary structures, 38-45, 50-52, 51t Amino acid sequences, 18-19, 19f, 25, 27-38 alignment of, 175-181, 176f-182f alternative conformations of, 51-52, 51f of collagen, 44, 44f databases of, in sequence homology analysis, 181, 182f determination of, 79-84 DNA sequencing in, 79-84 Edman degradation in, 81-83, 81f genetic, 35 protein cleavage for, 82-83, 82f, 83f uses of, 79-80

directionality of, 34, 34f in disease, 35-36 DNA-derived. 84 evolution of, 36, 79, 174-181 in genetic code, 129-130, 129t. See also Genetic code glycosylation sites in, 330 homologous, 175-181. See also Sequence homologies identification of, 65-66 internal repeats in, 79 protein structure and, 18-19, 19f, 25, 35-36, 49-59 shuffled, 177-178, 177f substitutions in, 178-181, 179f-181f Amino acid side chains, 27, 28–32 acidic, 31-33, 32f aliphatic, 28, 29f aromatic, 28 basic, 30-31, 31f carboxamide-containing, 30, 30f charge of, 30-33, 31f, 32f definition of, 27 hydrophilic, 30f, 46, 46f, 47, 47f hydrophobic, 28, 45-46, 46f, 47, 47f hydroxyl-containing, 30, 30f ionizable, 32, 32t sulfhydryl-containing, 30, 30f thiol-containing, 30, 30f Amino acid synthesis, 705–732 activated methyl cycle in, 717-718, 717f adenylation in, 725-726, 725f ammonia in, 706-710 branched pathways in, 723 chorismate in, 719-720, 720f committed step in, 723 enzymatic cascades in, 725-726, 725f, 726f enzyme multiplicity in, 724, 724f of essential amino acids, 711-712, 719-723 feedback inhibition/activation in, 723-724, 726f hvdroxyethyl-TPP in, 723, 723f metabolic precursors in, 711, 711f methylation in, 716-718, 717f nitrogen fixation in, 706-710 of nonessential amino acids, 712-719 one-carbon units in, 715-716, 715t, 716f overview of, 705-706 phosphorylation in, 724 rate of, 723 regulation of, 723-726 regulatory domains in, 724, 724f reversible covalent modification in, 725-726, 725f S-adenosylmethionine in, 716-718 shikimate in, 719, 720f in starvation, 809 substrate channeling in, 722-723, 722f tetrahydrofolate in, 715, 715-716, 716f threonine deaminase in, 723-724, 723f vitamin B₁₂ in, 650–652, 650f, 651f, 717

Amino acid-binding regulatory domains, 724, 724f Amino sugars, 330 2-Amino-3-carboxymuconate-6semialdehyde, 696, 696 Aminoacrylate, Schiff base of, 722, 722, 722f Aminoacyl adenylate, in translation, 893, 893-895, 894f Aminoacyl-AMP, 893-895, 894f Aminoacyl-tRNA, 128, 128f, 893, 893-894 Aminoacyl-tRNA synthetases, 128, 888, 893-897 activation sites of, 894, 894-895 classes of, 897, 897t editing sites of, 895, 895 proofreading by, 895 specificity of, 894-895 tRNA recognition by, 896-897 Aminoglycosides, 909, 909-910, 909t 5-Aminoimidazole ribonucleotide, 742, 742f 5-Aminoimidazole-4-carboxamide ribonucleotide, 741t, 742f 5-Aminoimidazole-4-(Nsuccinvlcarboxamide) ribonucleotide, 742, 742f δ -Aminolevulinate synthase, in porphyrin synthesis, 728, 729f Aminopterin, 750 Aminotransferases in amino acid degradation, 680-682 evolution of, 713 pyridoxal phosphate-dependent, 712-713 Ammonia in amino acid degeneration, 680-681, 685-690 in amino acid synthesis, 706-710 conversion to urea, 685-690 formation of, 680-681, 684, 713 in glucose-alanine cycle, 684-685, 685f in glutamate synthesis, 709 in glutamine synthesis, 709–710 neurotoxicity of, 689 in purine synthesis, 741 in pyrimidine synthesis, 737, 738 transport of, 684-685 in urea cycle defects, 689 AMP (adenosine monophosphate) in adenylation, 725, 725f degradation of, 752-753, 753f energy charge and, 446-447, 446f in fatty acid metabolism, 644-645, 646f in glycogen metabolism, 626 in reversible covalent modification, 725 structure of, 431 synthesis of, 743-744, 743f, 751, 751f AMP-dependent protein kinase, 666-667 Amphibolic pathways, 429 Amphipathic molecules, 351 α-Amylase, 328, 454 in glycogen branching, 629 Amyloid fibers, in neurodegenerative diseases, 55-56, 56f, 57

C4

INDEX

Amyloid plaques, in Alzheimer's disease, 55-56, 57 Amyloid precursor protein (APP), 57 Amyloidoses, 55-57 Amylopectin, 328 Amylose, 328 Amyotrophic lateral sclerosis, 147-148, 157 animal models of, 164 mutations in, 157, 163 Amytal, 558 Anabolic steroids, 785, 948 Anabolism, 428-429 definition of, 706b electron carriers in, 439 Anaerobes, obligate, 468-469 pathogenic, 468, 469t Anaplerotic reaction, 516-517 Anchor residues, 993, 993f Ancient DNA, amplification and sequencing of, 147, 188-189, 189f Andersen disease, 634t Androgen(s). See also Sex hormones; Steroid hormones functions of, 784-785 reduction of, 785 synthesis of, 780, 780f, 784, 784 Androstenedione, 784, 784, 948 Anemia Coolev. 210 erythropoietin for, 331 hemolytic, G6PD deficiency and, 609-610.611 sickle-cell, 209-210, 209f, 210f Anfinesen, Christian, 49-50 Angiogenin, sequence comparison for, 173, 174 Angstrom (Å), 7 Animal testing, in drug development, 1037 Anion exchange, 70 Ankyrin repeats, 972-973, 972f Annealing, of DNA strands, 120-121 Anomers, 321f, 323 Anosmia, 958 ANT (adenine nucleotide translocase). See ATP-ADP translocase Antagonists, 948 Anthranilate, 720, 721 Anthrax. 831 Antibiotics. See also Drug(s) and specific antibiotics protein synthesis inhibition by, 909-910 transcription termination by, 862-863 translocation inhibition by, 909-910, 909t Antibodies, 979–980. See also Immunoglobulin(s) antigen binding by, 984-986, 985f, 986f antigen interaction with, 83-86, 85f antigen-binding units of, 981-983 binding sites of, 984-985, 984f, 985f catalytic, production of, 243-244, 243f class switching in, 990-991, 991 constant domains of, 984, 984f definition of, 84-85, 979 diversity of, 980-981

formation of, 988–989, 990–991 gene recombination in, 987-991, 988f class switching and, 990-991, 991f hypervariable loops of, 984, 984f monoclonal for cancer treatment, 420-421 preparation of, 86-88, 87f number of, 980 oligomerization of, 989-990 polyclonal, 86, 86f production of, 86-88, 87f secretion of, 950, 989-990 structure of. 85f. 981-982 variable domains of, 983, 984, 984f, 985f Antibody diversity, 987–991 Anticipation, in genetic disease, 842 Anticoagulants, 310, 312 Anticodon loop, 890f, 891 Anticodons, 128, 889, 890f, 891-893, 891f, 892f, 896. See also Codons Antidiuretic hormone, 96 Antielastase, 307 Antigen(s), 85–86 ABO blood group, 335-336, 335f antibody binding of, 85, 85f, 981-986, 985f. 986f cross-linking of, 982, 982f definition of. 979 self. 1001 Antigen-antibody interactions, 83-86, 85f, 981-986, 985f, 986f Antigenic determinants, 85, 85f, 980 Antigen-presenting cells, 996-998, 996f Antihemophilic factor, 311, 311f Antimalarial drugs, hemolytic anemia and, 609-610 Antimycin A, 558 Antioxidants vitamin C as, 813 vitamin E as, 542 Antiporter, 380, 380f Antisense strand, 854 Antiserum, 86 Antithrombin III, 312 Antitransporters, 380-381, 380f, 381f α_1 -Antitrypsin, 304–307 abnormalities of. 312 Antituberculosis drugs, 679 AP endonuclease, 840 Apoenzymes, 221 Apolipoprotein B-48, 642, 773, 773t, 774, 872-873 Apolipoprotein B-100, 773t, 774, 774f, 775, 872-873 Apolipoprotein E, 773t, 774 Apolipoproteins, 642, 773, 773t, 774, 774f, 775. See also Lipoprotein(s) Apoproteins, 773 Apoptosis mitochondria in, 559 T cell, 1000 zymogens in, 303

effector units of, 981–983

Apoptosome, 559 Appetite regulation, 794–797, 795f–797f. 795t. See also Caloric homeostasis Approximation, catalysis by, 254 Aquaporins, 394, 395f Aqueous solutions. See also Water hydrogen ions in, 13-17 Arabidopsis, 783 Arachidonate, 664-665, 665f Arachidonic acid, 358 Arber, Werner, 141 Archaea, 3, 3f membranes of, 350-351 proteasomes of, 677-678, 678f transcription in, 869 Arginase, 687 Arginine, 687 nitric oxide from. 727–728 structure of, 30-31, 31f, 687 synthesis of, 712, 714, 714f in urea cycle, 687 Argininosuccinase, 687 deficiency of, 689, 689f Argininosuccinate, 686, 686, 689, 689 Argininosuccinate synthetase, 686 Argonaute-miRNA, 953, 953f Aromatase, 785 Aromatic amino acids, 28. See also Amino acid(s) degradation of, 695-696 synthesis of, 719-722, 720f, 721f Arrestin, 408, 969 Arsenite poisoning, 517, 518f Arteriosclerosis. See Atherosclerosis Arthritis, degenerative, 332 Artificial chromosomes, 151 Ascorbic acid (vitamin C), 442, 442t, 812-813, 813 deficiency of, alcohol-related, 812-813 forms of, 813, 813 Asparaginase in amino acid degradation, 692 in glycoproteins, 330, 331 structure of, 30, 30f synthesis of, 713 Asparagine in amino acid degradation, 692 carbohydrate units of, 57 synthesis of, 713, 713f Aspartate asparagine formation from, 713, 713f in catalytic triads, 258-260. See also Catalytic triads conversion to fumurate, 687, 692 conversion to oxaloacetate, 692 in double-displacement reactions, 237 in purine synthesis, 743 structure of, 31, 32f, 237, 290 synthesis of, 712-713 taste of, 962, 965 Aspartate aminotransferase, 683 in amino acid degradation, 680, 682

Index

C5

Aspartate transcarbamovlase (ATCase), 290-296 active sites of, 293-294 allosteric interactions in, 292-295 cytidine triphosphate inhibition of, 291, 291f, 295-296, 296f kinetics of, 291, 291f, 294-296, 294f-296f in pyrimidine synthesis, 290-295, 290f, 738, 751 structure of, 290, 291–292, 292 subunits of catalytic, 291-292 interaction of, 292-295, 294f-296f regulatory, 291-292 T-to-R state transition in, 294–296, 294f, 295f ultracentrifugation studies of, 291-292, 292f Aspartokinases, 724, 724f Aspartyl proteases, in peptide bond cleavage, 263, 264, 264f Aspirin discovery and development of, 1038-1040 as prostaglandin inhibitor, 358, 665, 1040 Assays definition of, 67 of enzyme activity, 67 in protein purification, 67 ATCase (aspartate transcarbamoylase), 289 Atenolol, 1045, 1045 Atheroma, 1040 Atherosclerosis, 759, 774-775, 776-779,777f drug therapy for, 1040 HMG-CoA reductase inhibitors for, 779, 779, 1040, 1040–1041, 1041 homocysteine in, 719 Atkinson, Daniel, 446 Atorvastatin, 1041, 1041 ATP (adenosine triphosphate), 112, 112, 279, 279 as activated carrier of phosphoryl groups, 438 ADP units in, 447, 447f as allosteric effector of ATCase, 296, 296f binding of conformational changes and, 280-282, 282f, 284f by RNA, 190-191, 190f body requirements for, 525, 554-560 in citric acid cycle, 498, 512, 514-515, 514f electrostatic repulsion and, 433 energy charge and, 446-447, 446f during exercise, 805-806, 805t in fatty acid metabolism, 644-645, 646f formation of, 430-431 from acetyl CoA oxidation, 437-438 from ADP, 430-431, 434, 435, 435f

binding-change mechanism in, 547, 547f from carbon oxidation, 434, 435-438, 435f. 437f chemiosmotic hypothesis for, 544-545, 544f, 545f creatine phosphate in, 434-435, 805 during exercise, 434-435, 435f, 805-806, 805t in fatty acid metabolism, 648 from glycolysis, 463-466 ion gradients in, 437-438, 437f in mitochondria, 526 NADH in, in ATP synthesis, 439 from oxidative phosphorylation, 436-437, 437f, 525, 543-550. See also Oxidative phosphorylation in photosynthesis, 565, 566, 578-581 proton-motive force in, 525-526, 544-545, 544f, 545f rotational catalysis in, 547-550, 547f-549f yield in, 554-555, 555t in gluconeogenesis, 484 in glycolysis as phosphoryl-group donor, 455, 458 as product, 463–466 in GMP synthesis, 743-744, 743f, 751 high-energy bonds of, 433-434 hydration stabilization in, 433 hydrolysis of in coupled reactions, 431-433 in DNA replication, 824-825 in DNA supercoiling, 830-831, 830f free energy from, 429-435 heat generation in, 489 myosin and, 279-282, 281f-283f in kinesin movement, 1021–1022, 1021fin muscle contraction, 1009–1012, 1011f, 1012f, 1015, 1016f rotational motion from, 547-550, 548f, 549f in splicing, 876-877 in transcription, 876-877 in membrane transport, 374-381. See also Ion pumps in mitochondria, 526 in muscle contraction, 1009-1012, 1011f, 1012f, 1015, 1016f in myosin movement, 1009-1012, 1011f, 1012f, 1015, 1016f in nitrogen fixation, 707, 708, 709-710 in oxidative phosphorylation, 525-526 in pentose phosphate pathway, 607f, 608-609 in phosphorylation, 298-301, 432-434, 437 - 438phosphoryl-group-transfer and, 434-435, 434t. See also Phosphorylation phosphoryl-transfer and, 434f in purine synthesis, 742f as reaction energy source, 429-435 resonance structures of, 433

in signal transduction, 405 structure of, 430, 431 in substrate cycles, 489 triphosphate moiety of, 430 in ubiquitination, 675-676, 675f utilization rates for, 435 ATP synthase (Complex V), 578-579, 579f a subunit of, 546, 548, 549 α subunit of, 545, 545 ATP-driven rotation in, 547-550, 547f-549f b subunit of, 546 β subunit of, 545, 545-547, 547f binding-change mechanism and, 547, 547f c ring of, 548–549, 549f catalytic action of, 546-548 in chloroplasts, 578-579, 583, 583f F₀ subunit of, **545**, 546 F₁ subunit of, **545**, 545–546 G proteins and, 550 γ subunit of, **545**, 545–546, 547 rotation of, 547-550, 547f-549f, 1023 inhibition of, 558 in mitochondria, 544-550, 558 in oxidative phosphorylation, 544-550, 558 in photosynthesis, 578-579, 583, 583f P-loop NTPase domains in, 284, 284f proton flow through, 546-547 rotational, 547-550, 547f-549f respiratory chain linkage to, 544-545, 544f, 545f structure of, 545, 545-546 in thylakoid membrane, 583 ATP synthasome, 554 ATP-ADP cycle, energy exchange in, 430-431, 435, 435f ATP-ADP translocase, 552-553, 553f, 554f inhibition of, 558 ATPase(s) AAA in amino acid degradation, 677 DnaA as, 834 in dynein, 1010, 1011f mitochondrial. See ATP synthase in motor proteins, 1008-1010, 1011f P-type, 374-378 evolution of, 374, 378 functions of, 378 in membrane transport, 374–378. See also Ion pumps ATPase domains, of myosin, 279-280, 280f ATP-binding cassettes, 378–380, 379f ATP-citrate lyase, 662 ATP-driven pumps, 371 ATP-grasp domain, 482, 482f ATP-grasp fold, 737 ATP-magnesium complex, 280 ATP-myosin complex, 279-280, 280f Atrazine, 584, 584 Attenuation, 931-932, 932f

C6

INDEX

Auditory processing, 971-973 Autoimmune diseases, 1001 Autoinducers, 930, 930f Autonomously replicating sequence, 151, 151f Autoradiography, 73, 143 Autotrophs, 565, 590 Axel, Richard, 959 Axonemes, 1019 Azathioprine, 1048, 1048 B cell, 989–990 memory, 1002 B cell receptor, 989, 989f B vitamins. See Vitamin(s) Backbone models, 61-62, 62f Backbones, in nucleic acids, 110, 110f, 111 Bacteria, 3, 3f. See also Escherichia coli chemotaxis in, 1024-1025, 1024f DNA replication in, 831-834. See also DNA replication flagellar movement of, 1022-1025 glyoxylate cycle in, 518-519, 519f nitrogen fixation in, 706-710 photosynthetic, 584, 584t reaction center of, 569-572, 570f proteasomes of, 677-678, 678f Bacterial artificial chromosomes, 151 Bacterial photosynthetic reaction center, 569-572, 570f Bacteriochlorophyll, 570, 570 Bacteriochlorophyll b, 570–571 Bacteriopheophytin b, 570, 570, 570–571 Bacteriorhodopsin, 356, 356f Baculovirus vectors, 163–164 BAL (British anti-lewisite), 517, 518f Ball-and-chain model, of ion channel inactivation, 388-389, 389f Ball-and-stick models, 22, 23f, 61, 61f Band centrifugation, 77–78, 77f Basal transcription complex, 941 Base-excision repair, 840 Base-pair substitutions, tests for, 843 Base-paired hairpin. See Hairpin turns Bases abbreviations for, 112, 112f in acid-base reactions, 13-17. See also Acid-base reactions nucleic acid. See Bases/base pairs sequence of, notation for, 112, 112f Bases/base pairs, 110-113, 111, 114t. See also Double helix adenine, 4, 111, 111 adenine-thymine, 5, 5 complementarity of, 822, 822f, 823f cross-linkage of, 838, 839f cytosine, 4, 111, 111 damage to, 838-839 deamination of, 680-684, 838, 841 errors in base pairing and, 117-118, 837-838 repair of, 839-843. See also DNA repair

formation of, 891-893, 901. See also Translation codon-anticodon interactions in. 891-893 in genetic code, 128-131, 129t. See also Genetic code guanine, 4, 111, 111 guanine-cytosine, 5, 5 hydrogen bonds in, 5, 5f, 10. See also Hydrogen bonds methylated, in tRNA, 889-890 mismatched, 117-118, 118f, 837-838 nomenclature of, 736t nonstandard, 117-118, 118f, 837-838 non-Watson-Crick, 837-838 in nucleosides, 111, 111. See also Nucleoside(s) in nucleotides, 111-112, 112. See also Nucleotide(s) oxidation of, 838 purine, 111, 111 synthesis of, 740-745, 740f pyrimidine, 111, 111 synthesis of, 736-740, 736f recognition sites and, 141, 141f, 272, 275-277 in recombinant DNA technology, 139. See also Recombinant DNA technology RNA, homologous, 186-187, 187f rules for, 114 stacking of, 10, 10f, 114-115, 119f hypochromism and, 120, 120f thymine, 4, 111, 111 in translation, 891–893, 901 uracil, 111, **111** Watson-Crick, 5, 113-115, 114f, 186, 187f wobble in, 891-893 Basic-leucine zipper, 942 Bathorhodopsin, 967-968 Bcr-Abl, in chronic myelogenous leukemia, 421, 421f B-DNA helix, 115-116, 116t Bennett, Claude, 987 Benzaldehvde, 958, 958 Benzoate, 689 Berg, Howard, 548, 1023 Berg, Paul, 148, 644 Beriberi, 517 β cells, in diabetes, 802 β chains, of hemoglobin, 199, 199f, 211 β sheet, 40–42, 40f–42f amino acid residues in, 50-52, 51t β strands, 41–42, 41f amino acid residues in, 50-51, 51t of membrane proteins, 357, 357f β turns, 42, 42f amino acid residues in, 50–51, 51t β_2 subunit, of *E. coli*, in DNA replication, 831-832, 831f β-adrenergic receptor, 404-405, 404f β-adrenergic receptor kinase, 408 β-arrestin, 408

β-globin gene. See also Hemoglobin transcription and processing of, 131-132, 132f β-hemoglobin, 199, 199f, 211. See also Hemoglobin β-lactam ring, of penicillin, 244, 244f β₂-microglobulin, 992–993, 993f β-oxidation pathway, in fatty acid metabolism, 646-647 β-thalassemia, 210-211, 211 β-tubulin, 1018–1019, 1018f, 1019f Bicarbonate dehvdration of, carbonic anhydrase in. 266-271 in pyrimidine synthesis, 737 Bicarbonate ion, in oxygen transport, 207-208.208f Bifunctional enzyme, 487 Bile, in drug metabolism, 1035-1036 Bile salts, 642, 642f synthesis of, 779-780, 780f Bilirubin, 730, 730 Biliverdin, 730, 730 Biliverdin reductase, 730, 730 Binding energy, 229 definition of, 254 enzyme specificity and, 275-277 Binding-change mechanism, 547, 547f Biochemistry definition of. 1 unifying concepts in, 1-4, 2f, 3f Bioenergetics. See also Energy of coupled reactions, 429-430, 431-433 enzymes and, 222-225 of metabolism, 429-435 thermodynamic principles and, 11-13 Biofilms, 930-931 Bioinformatics, 173-192. See also Sequence homologies Biological diversity, biochemical unity underlying, 1-4, 2f Biological macromolecules, 2 Biopterin, 695 Biotin, 441t in fatty acid synthesis, 657 in gluconeogenesis, 482-483, 483f 1,3-Bisphosphoglycerate, 434 in glycolysis, 464, 484 oxidation of, 436-437 phosphoryl-transfer potential of, 434, 434t 2,3-Bisphosphoglycerate in glycolysis, 464 oxygen affinity of hemoglobin and, 204-205 Bisubstrate reactions, 235–236 Bitter taste, 962, 963-964 Bjorkman, Pamela, 992 Blackburn, Elizabeth, 837 BLAST search, 181, 182f Bleach, urine as, 687b Bleeding disorders, 311-312, 312 The Blind Watchmaker (Dawkins), 53 Blood clotting. See Clotting

Blood groups, glycosyltransferases in, 335-336 Blood-brain barrier, 1033 Blow, David, 257 Blue photoreceptors, 969-970, 970f Boat form, 324, 325f Body mass index (BMI), 792, 793f Body weight, regulation of, 791-798. See also Caloric homeostasis Bohr, Christian, 206 Bohr effect, 206-208, 207f Bombardment-mediated transformation, 167 Bonds covalent, 7 cleavage of, 13 disulfide, 35, 35f cleavage of, 49 reduction of, 83 electrostatic interactions, 7-8 in antigen-antibody binding, 984, 985, 985f ATP and, 433-434 in enzyme-substrate complex, 228, 229f glycosidic in disaccharides, 327 in monosaccharide alcohol/amine complexes, 326, 326f high-energy, 433-434 hydrogen. See Hydrogen bonds hydrophobic effect and, 9-10, 9f hydrophobic interactions and, 10 isopeptide, 675, 675f noncovalent, 7-11. See also Noncovalent bonds peptide. See Peptide bonds phosphodiester, 110, 110, 111 hydrolysis of, 272-274 in resonance structures, 7 surface complementarity and, 11 units of measure for. 7 van der Waals interactions. See Van der Waals interactions Bonitus, Jacob, 517b Bordetella pertussis, 422 Bortezomib (Velcade), 678, 679 Bovine spongiform encephalopathy, 56-57, 56f Boyer, Herbert, 148 Brain glucose metabolism in, in starvation, 809-810.810t sensory processing in. See Sensory systems vanishing white matter in, 908-909, 909f Branched-chain amino acids. See also Amino acid(s) degradation of, 693-695 Branched-chain ketoaciduria, 697 Branching enzyme, in glycogen metabolism, 629 Brenner, Sydney, 128 Briggs, George, 231 British anti-lewisite (BAL), 517

3-Bromoacetol phosphate, 242, 242 Bromodomains, 949-950, 950f Brown fat. 556-557 Brown, Michael, 767b, 775, 776 Bubble boy disease, 753 Buchner, Eduard, 454 Buchner, Hans, 454 Buck, Linda, 959 Buffers, 15-17, 15f, 16f in carbon dioxide hydration, 270, 270f Burkitt lymphoma, 869 t-Butyloxycarbonyl amino acid, 97, 97 Butyryl ACP, 658t, 659, 659 bZip, 942 C genes, 987 in antibody switching, 991 C₃ plants, 600 C₄ pathway, in photosynthesis, 599-600, 600f Ca²⁺ pump, 374–378, 375f, 376f CAAT box, 127, 867, 867f CAD, 688 Caenorhabditis elegans genome of, 159 RNA interference in, 166 Caffeine, metabolism of, 783 Calciferol (vitamin D), 442, 442t deficiency of, 786 sources of, 786 synthesis of, 785, 785f Calcineurin, 990, 990 Calcitriol, 785, 785 Calcium ATPase, in phosphoinositide cascade, 409 Calcium ATPase, 375-377 Calcium ion(s) in calmodulin activation, 410-411 in insulin secretion, 802 in phosphorylase kinase activation, 624, 624f prothrombin binding of, 310-311 in pyruvate dehydrogenase complex activation, 514 in signal transduction, 409-411, 409f-411f, 802 structure of, 403 in vision, 969 Calcium ion channels, 367, 384f. See also Ion channels sequence homologies of, 367, 384, 384f, 386 Calcium ion pump, 374-378, 375f, 376f evolution of, 378 Calmodulin, 79, 79, 624 in glycogen metabolism, 624 in signal transduction, 410-411 Calmodulin-dependent protein kinases, 411 Calnexin, 339 Caloric homeostasis, 791-798 definition of, 791 in diabetes, 799-802 dieting and, 797-798

evolution and, 792-793 exercise in, 803-806 fasting and, 796 food intake and. 793–794 insulin in, 796 melanocyte-stimulating hormone in, 796 obesity and, 792-803. See also Obesity satiation signals in, 795-797, 796f signaling in, 794-797, 794t, 795f-797f suppressors of cytokine signaling in, 797, 797f Calorie (cal), 223b Calreticulin, 339 Calvin cycle, 590-601 C₄ pathway and, 599-600 energy expenditure in, 597 evolution of, 600 hexose sugar formation in, 594-597, 594f oxygenase reaction in, 593, 593f pentose phosphate pathway and, 609 3-phosphoglycerate formation in, 591-592, 592f reactions in, 590-597, 590f, 596f regulation of, 598-600 ribulose 1,5-bisphosphate regeneration in, 594-597, 596f salvage pathway in, 593-594, 593f stages of, 590-591, 590f, 596f thioredoxin in, 598-599, 598t, 599 CaM kinase, 411 cAMP in glycogen metabolism, 625-626, 625f. 626 in protein kinase A activation, 301, 301f as second messenger, 402-403 in signal transduction, 405, 406-407 structure of, 301, 403 cAMP receptor protein (CRP), 927-928, 927f Camptothecin, 831 Cancer aerobic glycolysis in, 478-479 breast, taxol for, 1019-1020 carcinogen testing and, 843-844 citric acid cycle defects in, 515 colorectal, 842-843 cytochrome P450 and, 783 defective DNA repair in, 842-843 drug resistance in, 1050-1051 enhancers in, 869 fatty acid synthase in, 663 glycolysis in, 478-479, 479f hypoxia inducible factor 1 in, 515 immune response in, 1001 leukemia, 421 in Li-Fraumeni syndrome, 843 metabolic derangements in, 515 mutagens and, 838 myc gene in, 869 oncogenes and, 420 protein kinase inhibitors for, 421 proto-oncogenes and, 420 signal transduction defects in, 420

INDEX

Cancer—(continued) skin, in xeroderma pigmentosum, 842 telomerase inhibitors for, 837 thymidylate inhibitors for, 749-750 tumor hypoxia in, 478-479 tumor-suppressor genes and, 420 Warburg effect in, 478 CAP (catabolite activator protein), 927–928, 927f CAP-cAMP complex, 927-928 Capsaicin, 973-974, 973f Captopril, 264 Carbamate, 208, 208 in Calvin cycle, 592-593 Carbamic acid, 737, 737, 738, 738f Carbamoyl phosphate, 290, 686 in pyrimidine synthesis. 737 in urea cycle, 685-687, 685f Carbamoyl phosphate synthetase, 689f active sites on, 737-738, 738f deficiency of, 689, 689f isozymes of, 688 in pyrimidine synthesis, 737, 751 structure of, 737, 737-738, 738 synthesis of, 737 in urea cycle, 685–686 Carbamoylaspartate, 290, 738, 738. See also Aspartate transcarbamoylase (ATCase) biosynthesis of, 290-296 Carbohydrate-asparagine adduct, 57 Carbohydrates, 319-340 abbreviations for, 330b disaccharides, 327-329. See also Disaccharides in glycoproteins, 57, 329-337 lectins and, 337-339, 339f metabolism of, 454 monosaccharide, 320-327. See also Monosaccharides N-linked, 330, 330f, 334, 336 oligosaccharides, 327. See also Oligosaccharides O-linked, 330, 330f, 334 overview of. 319-320 peptide bond cleavage in, 337 polysaccharides, 328, 328f, 329f. See also Polysaccharides in red blood cells, 335 sequencing of, 336-337 starch, 328-329, 329f in viral infections, 339-340 Carbo-loading, 806 Carbon dioxide activated, biotin carrier for, 483, 483f in Calvin cycle, 591-597 from carbon oxidation, 435 hydration of, 206 carbonic anhydrase in. 266-271 in oxygen release by hemoglobin, 207-208, 207f, 208f transport of, C₄ pathway for, 599-600.600f

Carbon fuels body reserves of, 809 definition of, 498 for exercise, 805-806, 805t for glucose metabolism, 808, 809 oxidation of. See also Catabolism in citric acid cycle, 498. See also Citric acid cycle free energy of, 434, 435-438, 435f Carbon monoxide, in oxidative phosphorylation inhibition, 558 Carbon monoxide poisoning, 213 Carbon skeletons, of amino acids, metabolic fates of, 690-696, 691f Carbonic acid, biosynthesis of, 266-271 Carbonic anhydrases, 207, 207f, 266-271 alpha-type, 271 beta-type, 271 catalytic activity of, 266-271 evolution of, 271 gamma-type, 271, 271 structure of, 267, 267-268, 271, 271 zinc site in, 267-269, 267f-269f Carboxamides, 30, 30f 2-Carboxy-3-keto-D-arabinitol 1,5bisphosphate, 592f Carboxyaminoimidazole ribonucleotide, 742, 742f Carboxybiotin, 483, 483 Carboxyglutamate, 57, 58, 310, 310 biosynthesis of, 310-311 in prothrombin, 310-311 in vitamin K deficiency, 57, 310 Carboxyhemoglobin, 213-214 γ -Carboxylation, 298t Carboxyl-terminal domain (CTD), 865, 868, 877, 877f 1-Carboxyphenylamino-1-deoxyribulose 5-phosphate, 721, 722 Carcinoembryonic antigen, 1001 Carcinogens, tests for, 843-844 Cardiac muscle, H and M isozymes in, 297, 297f Cardiolipin, 349, 349 functions of, 762 synthesis of, 762–763 Cardiotonic steroids, 377, 377f Cardiovascular disease, 744-745, 759, 776-779 homocysteine in, 719 Cargo receptors, 913-914 Carnitine, 645, 645 deficiency of, 646 Carnitine acyltransferase I, 645, 666 Carnitine acyltransferase II, 646 Carotenoids, 582-583 Carriers, 371 Cartilage, 332, 332f Carvone, 958, 958 Caspases, 303, 559 Cassette mutagenesis, 157, 157f Catabolism, 428 definition of. 706b

electron carriers in, 438-439, 438f, 439f energy from, 435-438 free energy of, 434, 435-438, 435f, 437f stages of, 437-438, 437f Catabolite activator protein (CAP), 927-928, 927f Catabolite repression, 927-928 Catalase, 542 Catalysis by approximation, 254 in carbon dioxide hydration, 266-271 Circe effect in, 235b covalent, 254, 255-260. See also Covalent catalysis in DNA cleavage, 272 in double-displacement (ping-pong) reactions, 237 efficiency of, 235, 235t electrophilic, 683 enzyme activity in assays for, 67. See also Protein purification specific, 67 enzyme-substrate complex formation in, 226-227 general acid-base, 254 in metabolic regulation, 446 metal ion, 221, 221t, 254-255 in carbon dioxide hydrolysis, 267–269, 267f-269f in DNA cleavage, 274, 277 Michaelis constant (K_M) for, 231-235 in peptide bond cleavage, 258-260, 259f rate of, 231-235. See also Enzyme kinetics; Reaction rates reaction equilibrium and, 224-225 reaction rate acceleration and, 206, 220t, 225-229, 226f by RNA, 879-881 in selective binding of transition state, 225-226, 225f, 227-228, 243-244 in sequential reactions, 236-237 site-detected mutagenesis and, 262-263, 262f Catalytic antibodies, production of, 243-244, 243f Catalytic groups, 227 Catalytic RNA, 879-881, 900 Catalytic strategies, 254-286 of carbonic anhydrases, 253, 266-271 covalent, in chymotrypsin, 255-260 of myosins, 253-254 overview of, 253-254 of restriction endonucleases, 253, 271 - 279of serine proteases, 253, 255-266 Catalytic triads, 257-260 in chymotrypsin, 257-260, 261f in elastase. 261. 261f site-directed mutagenesis and, 262-263, 262f in subtilisin, 261, 261f in trypsin, 260-261, 261f

C9 Index

Catalytically perfect enzymes, 234, 235, 460 Cataracts, 472 Cation exchange, 70 CCA terminal region, 891 CCA terminus, of tRNA, 890f, 891 CD3, 995, 995f CD4, 997 CD8, 994-995, 995f cDNA (complementary DNA), 154-155, 154f, 162f in quantitative polymerase chain reaction, 162, 162f cDNA library, 154 CDP, hydrolysis of, 431 CDP-diacylglycerol, 761, 761 CDP-ethanolamine, synthesis of, 763 CDRs (complementarity-determining regions), 984, 984f, 985-986 Cech, Thomas, 852, 879 Celecoxib, 1044, 1044 Celecoxib (Celebrex), 1044 Cell cycle, 836, 836f Cell types, in eukaryotes, 938, 939t Cell-mediated immunity, 991 Cell-to-cell ion channels, 372, 393-394, 393f, 394f Cellular energy. See Energy Cellular immune response, 980 Cellular respiration, 498, 499f, 593f, 594. See also Respiratory chain definition of, 526b in photosynthesis, 593f, 594 regulation of, 554-560 Cellulase, 313 Cellulose, 328-329 Centrifugation band, 77-78, 77f density-gradient equilibrium, 119-120, 119f differential, 67-68, 68f gradient, 77-78, 77f homogenate in, 67, 68f sedimentation coefficients in, 76, 76t, 77f supernatant in, 67, 68f zonal, 77-78, 77f Ceramide, 764 sphingolipids from, 763-764 synthesis of, 764, 764f Cerebroside, 350, 350 synthesis of, 764 Cetuximab, 421 CF₁–CF₀ complex, 578–579. See also ATP synthase (Complex V) cGMP structure of, 403 in vision, 968f, 969 cGMP phosphodiesterase, in vision, 968 cGMP-gated Ca²⁺ channel, in vision, 968 Chain, Ernest, 1037 Chain-terminated fragments, in DNA sequencing, 143 Chair form, 324, 325f Changeux, Jean-Pierre, 202

Channels ion. See Ion channels water, 394 Charcot-Marie-Tooth disease, 1018 Chargaff, Erwin, 114 Charge separation, photoinduced, 569, 569f Charged tRNA, 891f, 893 Chemical modification reaction, 256 Chemical protons, 540f Chemical reactions. See Reaction(s) Chemical shifts, in NMR spectroscopy, 101, 101f Chemiosmotic hypothesis, 544–545, 544f, 545f Chemistry, combinatorial in drug development, 1041. See also Drug development in evolution studies, 188-189 Chemoattractants, 1024 Chemorepellants, 1024 Chemotaxis, 1024-1025, 1025f Chemotherapy, cancer, resistance in, 1050-1051 Chemotrophs, 428 Cheng–Prusoff equation, 1031 CheY, 1024 ChIP (chromatin immunoprecipitation), 945 Chirality, 27 of amino acids, 28, 29f, 713 Chitin, 333, 333f Chloramphenicol, 909t, 910 Chlorobium thiosulfatophilum, 584, 584t Chlorophyll a, 568, 568–572, 568f, 569f in photosystem II, 573, 573f Chlorophyll b, 582, 582-583, 582f Chloroplasts, 567-568, 567f ATP synthesis in, 578–581, 578f evolution of, 568 genome of, 568 starch in, 597 structure of, 567, 567f Chlorpromazine, 1038 discovery of. 1038 mechanism of action of. 1038.1039f Cholecalciferol, 785, 785 Cholecystokinin, in caloric homeostasis, 794-795, 795f Cholera, 421-422, 765 Cholesterol. 350. 350 "bad," 744 bile salts from, 779-780 elevated levels of, 744-745, 759, 776-779, 777f "good," 744–745, 776 labeling of, 767f in lipid membrane, 363 metabolic fates of. 775-776 metabolism of, 775-779 properties of, 767, 767b receptor-mediated endocytosis and, 385f steroid hormones from, 780-782

transport of, 773-774, 774-775 reverse, 778-779 Cholesterol synthesis, 767-779 condensation mechanism in, 769, 769f hepatic, 770, 774f isopentenyl pyrophosphate in, 768 mevalonate in, 767-768 rate of, 770-773 regulation of, 770-779 site of, 773, 774f squalene in, 768-770 stages of, 767 sterol regulatory element binding protein in, 770-771, 771f Choline, in phospholipids, 349 Chondroitin 6-sulfate, 331, 332 Chorismate, in amino acid synthesis, 719-720, 720f Chromatin, 937-941, 938-941 definition of, 938-939 structure of, 938f DNA packing in, 941 in gene regulation, 944-951 remodeling of, 944-951, 949-951, 950f structure of, 939 transcription factors and, 946-947 Chromatin immunoprecipitation (ChIP), 945 Chromatin-remodeling engine, 950, 950f Chromatography affinity, 70-71, 70f, 75, 78 gel-filtration, 69, 69f, 75 high-pressure liquid, 71, 71f ion-exchange, 69-70, 69f, 75, 75t in amino acid identification, 80-81, 80f Chromogenic substrate, 256 Chromophores, 967 Chromosomes bacterial artificial, 151 veast, 937, 938f yeast artificial, 151, 151f Chronic myelogenous leukemia, treatment of, 421 Chylomicron remnants, 773 Chylomicrons, 642, 642f, 773, 773t Chymotrypsin, 186, 255-260 active sites of, 185-186, 186f, 257-258 catalytic triad in, 257-260 covalent catalysis in, 255-260 acyl-enzyme intermediate in, 257, 259f tetrahedral intermediate in, 258-259, 259f as two-stage process, 256-257, 257f, 259f homologs of, 260-261, 261f inhibition of, 53f, 241-242, 241f in peptide bond cleavage, 259f serine residue of, 255-256, 256f specificity of, 256f, 260, 260f structure of, 256, 257-258, 258, 260, 261f substrate preferences of, 234-235, 235t trypsin and, 260-261, 261f Chymotrypsin inhibitor, structure of, 53f Chymotrypsinogen, 257-258, 303-305

INDEX

Cilia cochlear, 971-973, 971f, 972f microtubules in, 1019 Ciprofloxacin, 831, 831 Circadian rhythms, 974 Circe effect, 235, 235b Circular DNA, 117, 117f Cirrhosis, 811-812 Cis configuration, of peptide bonds, 36–37, 36f, 37f Citrate in fatty acid metabolism, 662, 662f, 666, 667f isomerization of, in citric acid cycle, 506, 506f Citrate synthase, in citric acid cycle, 504, 505f Citric acid cycle, 497-520 acetyl coenzyme A in, 497, 500-501, 501, 503-505, 514-515, 653, 656 aconitase in, 506, 506 ATP in, 498, 512, 514-515, 514f, 554-555, 555t in cancer, 515 in cellular respiration, 499f citrate isomerization in, 506, 506f citrate synthase in, 503-505, 505f citryl coenzyme A in, 505 definition of, 497 electron-transport chain in, 498, 512 enol intermediate in, 504 enzyme complexes in, 512 evolution of. 518 function of, 498 glycolysis and, 499-503, 499f intermediates in, from amino acid degradation, 690-696 isocitrate dehydrogenase in, 506, 514, 514f isocitrate in, 506-507, 507f α -ketoglutarate dehydrogenase complex in, 499, 507, 514-515, 514f α -ketoglutarate in, 506–507, 507f, 692 ketone bodies in, 810, 810f malate in. 510 in mitochondria, 498, 498f net reaction of, 510 nucleoside diphosphokinase in, 508 overview of, 497-499 oxaloacetate in, 498, 498f, 504, 505f, 509-510, 515, 653 oxalosuccinate in, 506, 507f pyruvate carboxylase in, 516 pyruvate dehydrogenase complex in, 499-503, 499f, 499t, 502f, 503f pyruvate dehydrogenase in, 500 rate of, 556f reaction rate in, 556 reactions of, 510-512, 511f, 511t regulation of, 512-516, 555-556, 556f as source of biosynthetic precursors, 516-518 stoichiometry of, 510-512, 511f, 511t substrate channeling in, 512

succinate dehydrogenase in, 509-510 succinyl coenzyme A in, 507-508, 507f. 514 succinyl coenzyme A synthetase in, 507-508 Citrulline, 686, 686, 689 Citrullinemia, 698t Citryl coenzyme A, in citric acid cycle, 505 Clamp loaders, 832 Class I MHC proteins, 991-996, 993f, 998-999. See also Major histocompatibility complex proteins Class II MHC proteins, 996-999, 996f Class switching, 990-991, 991f Clathrin, 775, 776f Cleavage DNA. See DNA, cleavage of protein, 82-83, 82f Cleland notation, 236, 237 Cleland, W. Wallace, 236 Clinical trials, 1048-1049, 1049f Clones, DNA, 154-155 Cloning, 87-88 expression, 154–155, 155f plasmid vectors in, 149-151, 149f, 150f Cloning vectors, 148, 149 Clotting, 307-313 extrinsic pathway of, 308, 308f impaired, 310-313, 311-313 in hemophilia, 311 intrinsic pathway of, 308, 308f regulation of, 311-312 zymogen activation in, 307-312 Clotting factors, 312 CMP (cytidine monophosphate), 762 Coactivators, 947-948, 948f, 949-951 Coagulation. See Clotting Coat proteins, 914 Coated pits, 775, 776f Cobalamin (vitamin B₁₂), 441t, 649 in amino acid synthesis, 650-652, 650f, 651f, 717 as coenzyme, 650-652, 650f, 651f, 717 in fatty acid metabolism, 649-652 structure of, 650, 650-651 Cobratoxin, 390 Cochlear hair cells, 971-973, 971f, 972f Coding strand, 854 Codons, 19, 109-110, 128. See also Anticodons definition of, 128 in genetic code, 129-130, 129t initiation, 900-901 in translation, 889, 891–893, 892f Coenzyme(s), 221, 221t vitamin, 441t Coenzyme A, 440, 440, 441t as acyl group carrier, 440 ADP units in. 447, 447f in fatty acid metabolism, 644 Coenzyme B₁₂, 650–652, 650f, 651f Coenzyme Q (ubiquinone), 532-533, 533f in fatty acid metabolism, 646

Cofactors, enzyme. See Coenzyme(s) Cognate DNA, 272 cleavage of, 275-277 Cohen, Stanley, 148 Cohesive-end method, 148-149, 149f Coiled-coil proteins, 43, 44f Collagen, 303 amino acid sequences of, 44, 44f ascorbate and, 813 in cartilage, 332 helix of, 44-45, 44f Collagenase, 303 Color blindness, 970-971 Color vision, 966, 969-971. See also Vision in animals, 970, 970f defective, 970-971 evolution of, 970, 970f Colorectal cancer, 842-843 Combinatorial association, antibody diversity and, 988-989 Combinatorial chemistry, in evolution studies, 188-189 in drug development, 1041. See also Drug development Combinatorial control, 943 Committed step, 474, 723 Compactin, 1040, 1040 Comparative genomics, 160-161, 160f Compartments, drug target, 1033, 1033f Competitive inhibition, 238-240, 238f, 239f, 240f Complement cascade, 981 Complementarity-determining regions (CDRs), 984, 984f, 985-986 Complementary DNA (cDNA), 154-155, 154f in quantitative polymerase chain reaction, 162, 162f Complementary single-stranded ends, 148 Computer databases, for amino acids, 65-66, 181, 182f Concentration gradient, 373-374 in ATP synthesis, 437-438, 437f Concerted mechanism, in substrate binding, 294-296 Concerted (MWC) model of allosteric enzyme kinetics, 296, 296f of hemoglobin oxygen binding, 202-203, 203f, 214-216, 216f Cones, 966, 969-970, 969f Congenital erythropoietic porphyria, 730-731 Congestive heart failure, digitalis for, 377 Conjugation, in drug metabolism, 1034-1035 Connexin, 394 Connexon, 394, 394f Consensus sequences, 126b, 126f, 299 in phosphorylation, 299 in promoters, 857, 857f in splicing, 132f, 133, 873-874, 873f Conservative substitutions, 178 Constant (C) genes, 987 in class switching, 991

C11 Index

Constant regions, 984, 984f CRP (cAMP receptor protein), Constitutional isomers, 321, 321f Continuous genes, 131, 132 Controlled termination of replication, Convergent evolution, 185-186 Cooperative binding, 200-201, 201f, Cooperative transition, in protein folding, 52 Coproporphyrinogen III, 729, 729 Cyclin, 836 Coronary artery disease. See Atherosclerosis Corrin ring, 650-651, 650f Corticosteroids. See also Steroid hormones Cortisol, synthesis of, 780, 780f, 783, 784 Corynebacterium diphtheriae, 910 Covalent bonds, 7. See also Bonds Covalent catalysis, 254, 255-260 acyl-enzyme intermediate in, 257, 259f chymotrypsin and, 255-260 Cysteine tetrahedral intermediate in, 258-259, 259f as two-stage process, 256-257, 257f, 259f Covalent modification, of proteins, 57-58, mechanisms of, 298t. See also

development of, 1043-1045 CpG islands, 946 CpG, methylation of, 945-946 Crassulacean acid metabolism, 600-601, 600f Cre recombinase, 845-846 Creatine kinase, 236-237 in sequential reactions, 236-237, 236f Creatine phosphate, 434 during exercise, 434-435, 435f, 805, 805t phosphoryl-transfer potential of, 434, 434t Creutzfeldt-Jakob disease, 56-57, 56f Crick, Francis, 5, 114, 127, 128, 891 Cristae, 526, 527f Critical concentration, 1014 Crixivan (indinivar), development of, 1043.1043f Cro, λ repressor and, 929 Cross-links, 838, 839f Crotonyl ACP, 658t, 659, 659

Crown gall, 166-167, 166f

143-144, 143f

Cooley anemia, 210-211

203f, 294

Cooperativity, 289

Cordycepin, 872

Core enzyme, 856

Corepressors, 927

Corev, Robert, 38

Cori disease, 634t

Cori, Gerty, 454, 634

synthesis of, 783

Cotransporters, 380, 380f

57f, 297-302, 298t

Phosphorylation

COX2 inhibitors, 1044

Corticosterone, 784

Coulomb's law, 7-8

cleavage of, 13

Cori cycle, 490, 490f, 685

Cori, Carl, 454

Core promoter, 857

927-928, 927f Crystallography, X-ray, 98-100, 98f-100f of enzyme-substrate complexes, 227, 227f c-Src, in cancer, 420 CTD (carboxyl-terminal domain), 865, 868, 877, 877f CTP. See Cytidine triphosphate (CTP) Cumulative feedback inhibition, 725 Cupric ion, 325 Cyanide, 558 Cvanobacteria, 568, 568f Cyanogen bromide, in protein cleavage, 82, 82f Cyclic adenosine monophosphate. See cAMP Cyclic hemiacetals, 322 Cyclic photophosphorylation, 580-581, 580f ubiquitination of, 298, 298t Cyclin B, degradation of, 673f Cyclin destruction boxes, 676 Cyclin-dependent protein kinases, 836 Cycloheximide, 909t, 910 in taste, 963-964 Cyclooxygenase 2, 1044 Cyclooxygenase inhibitors, development of, 1044-1045 Cyclophosphamide-glutathione conjugate, 1034, 1034 Cyclosporin, 990, 990 Cystathionine, 718-719, 719 molecular models of, 23f pyruvate formation from, 691, 691f structure of, 30, 30f synthesis of, 714-715, 718-719 Cysteine proteases, in peptide bond cleavage, 263, 263–264, 264f Cystine, 35, 35f Cytidine, 111 synthesis of, 739-740 Cytidine diphosphodiacylglycerol (CDPdiacylglycerol), 761, 761 Cytidine monophosphate (CMP), 762 Cytidine triphosphate (CTP), 761 ATCase inhibition by, 291, 291f, 295–296, 296f hydrolysis of, 431 in pyridine synthesis, 291, 291f, 295-296, 296f structure of, 290, 291 synthesis of, 739-740 Cytidine triphosphate synthetase, 739 Cytochrome definition of, 535 in photosynthetic reaction center, 572. 572f Cytochrome *b*₅, 664, 664f Cytochrome bf complex, 575, 575f location of, 583, 583f

Cytochrome c, 533, 535 in apoptosis, 559 evolution of, 543, 543f in oxidative phosphorylation, 531, 532f, 532t Cytochrome c oxidase (Complex IV), 531, 532f, 532t in oxidative phosphorylation, 531 in respiratory chain, 537-540, 538f-540f Cytochrome c oxidoreductase, 535–536 Cytochrome P450, 782-783, 782f in drug metabolism, 1034 Cytochrome P450 monooxygenases, 781 Cytochrome reductase, 531, 532f, 532t, 535-536, 536f Cytoglobin, 212 Cvtokines, 998, 998f Cytoplasm fatty acid synthesis in, 657 glycolysis in, 498 Cytosine, 4, 4, 111, 111 deamination of, 838, 841 Cytoskeleton, 1007, 1008f Cytotoxic T cells, 980, 994–996, 997–998, 997f. See also T cell(s) D amino acids, 27, 27f D genes, 988 in antibody switching, 991 D stereoisomers monosaccharide, 320 of monosaccharides, 321f, 322f DAG. See Diacylglycerol (DAG) Dalton, 34 Dark reactions, of photosynthesis, 566, 589, 590-601. See also Calvin cycle; Photosynthesis Darst, Seth, 853 Databases, of amino acid sequences, 65-66, 181, 182f Dawkins, Richard, 53 DCC (dicyclohexylcarbodiimide), 97, 97 Deamination, 838, 838f, 841 in amino acid degradation, 680-684 Decarboxylation, 484 in citric acid cycle, 500, 507 in fatty acid synthesis, 659 in gluconeogenesis, 484 in pentose phosphate pathway, 601 Decorated actin, 1014 Degenerative arthritis, 332 Degron, 676 Dehydration, in amino acid degradation, 684 Dehydroascorbic acid, 813, 813 7-Dehydrocholesterol, 785, 785 Dehydrogenases, NAD⁺ binding sites in, 469,469f 3-Dehydroquinate, in amino acid synthesis, 719, 720 3-Dehydroshikimate, 720 Deletions, production of, 156

C12

INDEX

Denatured proteins, 49-50, 49f, 50f Density-gradient equilibrium sedimentation, 119-120, 119f Deoxyadenosine, 111, 872 5'-Deoxyadenosyl radical, 651, 651f 5'-Deoxyadenosylcobalamin, 650 Deoxyadenylate, 111 3-Deoxyarabinoheptulosanate 7-phosphatase, in amino acid synthesis, 719, 720 Deoxycycline, 240 Deoxycytidine, 111 Deoxycytidylate, 111 Deoxyguanosine, 111 Deoxyguanosine 3-monophosphate (3'-dGMP), 112, 112 Deoxyguanylate, 111 Deoxymyoglobin, 196, 197. See also Myoglobin Deoxyribonucleic acid. See DNA Deoxyribonucleoside 3'-phosphoramidites, in DNA synthesis, 144, 144, 144f Deoxyribonucleoside triphosphate, in replication, 822, 823f Deoxyribonucleotide synthesis, 745-750 deoxyuridylate in, 748 dihydrofolate reductase in, 749 regulation of, 752, 752f ribonucleotide reductase in, 745, 745-750, 745f, 746, 752, 752f thymidylate in, 748, 749f Deoxyribose, 110, **110,** 322f Dephosphorylation, 300 (-)Deprenyl, 242, **242** Dermatan sulfate, 331, 332 1-Desamino-8-D-arginine vasopressin, 96, 96 Desaturase, 664, 664f Designer genes, 157 Desmolase, 783 α-Dextrinase, 454 DHAP. See Dihydroxyacetone phosphate (DHAP) DHU loop, 891 of tRNA, 890f Diabetes insipidus, 96 Diabetes mellitus, 325-326, 798-803 as autoimmune disease, 1001 derivation of name, 798b glucose homeostasis in, 798-800 incidence of, 798 insulin in, 798-800 insulin resistance and, 799-802 ketosis in, 633f, 653, 655, 803 obesity and, 798-803 pancreatic failure in, 801-802 type 1, 798, 802-803 type 2, 798-802 diabetes mellitus, type 2 pathogenesis of, 800-802 treatment of. 802 Diabetic ketoacidosis, 803 Diacylglycerol (DAG), 408, 642 in phospholipid synthesis, 761, 761f, 762,762f

in signal transduction, 408-409, 409f synthesis of, 761 Diacylglycerol 3-phosphate, 348, 348 in membrane lipid synthesis, 760 Diacylglycerol kinase, 760 Dialysis, in protein purification, 69, 69f Dianabol, 948 Diastereoisomers, 321, 321f Diazotrophic microorganisms, 707 5,5-Dibromo-4,4'-dichloro-indigo, 924 Dicarboxylate carrier, 554, 554f Dicoumarol, 310, 310 Dictvostelium discoideum, myosin-ATP complex in. 280, 280f Dicyclohexylcarbodiimide (DCC), 97, 97 Dideoxy method, 143-144, 143f 2,4-Dienoyl CoA reductase, 649, 649 Diet, 20, 20f. See also specific nutrients high-protein low-carbohydrate, 798 ketogenic, 656 low-phenylalanine, 674 spicy food in, capsaicin in, 973-974, 973f starved-fed cycle and, 807-808. See also Starvation Dietary fiber, 329 Dieting, 797-798 Differential centrifugation, 67-68, 68f Diffusion active, 371, 373 facilitated, 371, 373 lipid lateral, 361-364, 362f transverse, 362 simple, 372 Digestion, 437, 674, 674f. See also Amino acid degradation chymotrypsin in, 255-260. See also Chymotrypsin enzymes in, 255-260, 303-307, 303t, 304f, 674, 674f starved-fed cycle and, 807-808 Digitalis, Na^+-K^+ pump inhibition by, 377 Digitoxigenin, 377, 377 Diglyceride acyltransferase, 761 Dihydrobiopterin, 695, 695 Dihydroceramide, 764 1,25-Dihydrocholecalciferol, 785, 785 Dihydrofolate, in deoxyribonucleotide synthesis, 748, 749 Dihydrofolate reductase, 695, 695f in deoxyribonucleotide synthesis, 749 Dihydrolipoyl dehydrogenase, in citric acid cycle, 499t, 501, 502f Dihydrolipoyl transacetylase, in citric acid cycle, 499t, 501 Dihydroorotate, 738, 738 Dihydropteridine reductase, 695, 695f Dihydrosphingosine, 764 in sphingolipid synthesis, 737 Dihydrotestosterone, 785 Dihydrouridine (UH₂), 890

Dihydroxyacetone, 320, 320, 321f structure of, 321f in transaldolase reaction, 605-606, 606f Dihydroxyacetone phosphate (DHAP), 326, 326f, 458, 479 in Calvin cycle, 594f, 595, 595 in fructose metabolism, 469-470, 469f in gluconeogenesis, 480f in glycolysis, 456f, 458-459, 458f, 460f, 469-470, 480f isomerization to glyceraldehyde 3-phosphate, 224, 224, 458-459, 460f Dihydroxycholesterol, 783, 783 Diisopropylphosphofluoridate (DIPF), enzyme inhibition by, 241, 241f, 241 2,2-Dimercaptopropanol (BAL), 517, 518f Dimerization arm, 416 Dimers, 48 Dimethylallyl pyrophosphate, 742f, 768, 768.768-769 Dimethylbenzimidazole, 651 N, N-Dimethylpropargylamine, 242, 242 2,4-Dinitrophenol (DNP), 558, 558 Dioxygenases, 696 Dipalmitoyl phosphatidylcholine deficiency, 765-766 DIPF (diisoprophylphosphofluoridate), enzyme inhibition by, 241, 241, 241f Diphosphatidylglycerol, 349, 349 functions of, 762 synthesis of, 762 Diphthamide, 910-911 Diphtheria, 910–911 Dipolar ions, 27-28, 27f Direct repair, 840 Disaccharides, 327-329. See also Carbohydrates abbreviations for, 330b structure of, 327, 327f Discontinuous genes, 131 Discontinuous (split) genes, 131-132, 132f. 133f evolutionary advantages of, 133 Diseases and disorders albinism, 698t alcaptonuria, 697 alcohol-related, 811-813 Alzheimer disease, 55-56, 57 amino acid sequences and, 35-36 amyloidoses, 55-57 amyotrophic lateral sclerosis, 147-148 Andersen disease, 634t anemia, 330-331 anthrax, 831 anticipation in, 842 argininosuccinase deficiency, 689, 689f arsenite poisoning, 517, 518f atherosclerosis. See Atherosclerosis autoimmune, 1001 beriberi, 517 bleeding disorders, 310-313, 311-313

C13

Index

bovine spongiform encephalopathy, 56 cancer. See Cancer carbamovl phosphate synthetase deficiency, 689, 689f carbon monoxide poisoning, 213 cardiovascular disease, 719, 759, 774-775, 776-779. See also Atherosclerosis carnitine deficiency, 646 cataracts, 472 Charcot-Marie-Tooth disease, 1018 cholera, 421-422, 765 citrullinemia, 698t congenital disorders of glycosylation, 336 congestive heart failure, 377 Coolev anemia, 210-211 coronary artery disease, 759, 774-775, 776-779, 777f. See also Atherosclerosis Creutzfeldt-Jakob disease, 56 diabetes insipidus, 96 diabetes mellitus. See Diabetes mellitus diagnosis of, polymerase chain reaction in, 146–148 diphtheria, 910-911 drug-resistant, 1050-1051 emphysema, 307 environmental factors in, 19-20 epilepsy, 656 familial hypercholesterolemia, 759, 776-779 galactosemia, 472 gargovlism, 332, 332f gene therapy for, 167-168 genetic variations and, 19-20 glucose 6-phosphate dehydrogenase deficiency, 609-610, 611 glycogen storage diseases, 634-635, 634t gout, 753-754 heart disease, 719 hemolytic anemia, 609-610, 611 hemophilia, 311 Hers disease, 634t HIV infection. See Human immunodeficiency virus infection homocystinuria, 698t Huntington disease, 55-56, 842 Hurler disease, 332, 332f hyperlysinemia, 698t I-cell disease, 336 inborn errors of metabolism, 697-698, 698t infant respiratory distress syndrome, 765-766 lactose intolerance, 471 Leber hereditary optic neuropathy, 559 Lesch-Nyhan syndrome, 754 Li-Fraumeni syndrome, 843 long QT syndrome, 392-393 mad cow disease, 56 malaria. 610 maple syrup urine disease, 697 mercury poisoning, 517 misfolded proteins in, 56-57 mitochondrial, 558-559

mucopolysaccharidoses, 332 multidrug resistance in, 378-379 multiple myeloma, 87-88 mushroom poisoning, 865, 865-866 mutations causing, 147-148, 157 neurological, protein misfolding in, 56-57, 56f ornithine transcarbamoylase deficiency, 689 osteoarthritis, 332 osteogenesis imperfecta, 45 osteomalacia, 786 Parkinson disease, 55-56, 57, 242, 676 phenylketonuria, 674, 697-698 phosphatase deficiency, 514 porphyrias, 730-731 predisposition to, 19-20 prion, 56-57, 56f protein aggregates in, 56-57 recombinant DNA technology and, 146-148 retinitis pigmentosa, 877-878 ricin poisoning, 911 rickets, 786 scurvy, 57, 812-813 seizures, 656 severe combined immunodeficiency, 167 - 168sickle-cell anemia, 209-210, 209f, 210f smallpox. 1003 spina bifida, 755 splicing defects and, 877-878, 879t steatorrhea. 642 Tay-Sachs disease, 765-766 thalassemia, 210-211 transmissible spongiform encephalopathies, 55-56 tuberculosis, 679 tyrosinemia, 698t urea cycle defects, 668-669 vanishing white matter, 908-909, 909f vitamin D deficiency, 786 Wernicke-Korsakoff syndrome, 812 whooping cough, 422 Zellwegger syndrome, 652 Dismutation, 541b, 542f D isomers, 321f, 322f monosaccharide, 320 Displacement loop, 845, 845f Dissociation constant (K_d), 1030–1031 for ligand binding, 1030-1031, 1030f Distal histidine, 198, 198 Distributive enzymes, 832b Disulfide bonds, 35, 35f cleavage of, 49 Diuron, 584, 584 Divergent evolution, 185 Diversity (D) genes, 988 in class switching, 991 D-loop, 845, 845f DNA, 110 A-form, 115–116, 116, 116t ancient, amplification and sequencing of, 147, 188-189, 189f

annealing of, 120-121 antisense strand of, 854 backbone of, 110, 110f, 111 bases in, 4, 4, 4f, 5, 5-6, 110-111, 111. See also Bases/base pairs B-form, 115-116, 116, 116t in chromatin, 938f, 939-941 circular, 117, 117f cleavage of in cognate vs. noncognate DNA, 275-277, 276f in-line displacement of, 272-274 magnesium in, 274, 275f, 277 mechanisms of, 272-274 methylation in, 277-278 phosphodiester bridge hydrolysis in, 272-274, 272f phosphorothioates in, 273-274, 274f restriction enzymes in, 141 42, 271-279 restriction-modification systems in, 272, 272f sites of, 141, 141f, 271-272, 272, 275-277. See also Recognition sites stereochemistry of, 273-274, 274f cloned, 154-155 coding strand of, 845 cognate, 272 cleavage of, 275-277 complementarity with mRNA, 126, 126f complementary, 154-155, 154f, 162f condensed, 828 cross-links in, 838, 839f damage to causes of, 837-839 repair of, 839-843. See also DNA repair denaturation of, in acid-base reactions, 14-15, 14f directionality of, 112 double helix of, 5–15, 6f, 26f, 109, 113-115. See also Double helix evolution of, molecular studies of, 189-191, 190f functions of, 2, 18, 19 hybridization of, 121 hypersensitive sites in, 944 junk, 19 lagging strand of, 823f, 824 leading strand of, 823f, 824 length of, 113 linker, 148-149, 149f, 939 linking number of, supercoiling and, 826, 827f methylation of, 718, 718f, 945-946 in amino acid synthesis, 716-718, 718f in cleavage, 277-278, 278f minor groove in, 822, 822f mitochondrial, 527-528 genetic code of, 131, 131t sequencing of, 158 noncoding, 159-160 in nucleosome, 939-941 operator, 922f overview of, 4-6, 819-820

INDEX

DNA—(continued) packing of, 941 palindromic, 141 phosphodiester bridges of, 110, 110, 111 hydrolysis of, 272-274 polarity of, 112 primer strand of, 122, 145, 821 promoter sites in, 126-127, 126f, 852 in bacteria, 852, 856-858, 857f properties of, 4-6 in protein encoding, 18-19 recombinant. See Recombinant DNA relaxed, 117, 117f, 826, 828f renaturation of. 120-121 rewinding of, in transcription, 859, 859f sense strand of, 845, 854 size of. 113 stem-loop motif in, 118f sticky ends of, 148, 149f structure of, 4-6, 4f-6f, 110, 110-113, 112 sugar-phosphate units of, 4, 4f sugars in, 110-111, 110f supercoiled, 117, 117f. See also Supercoiled DNA synthesis of, 5-6, 6f, 113, 119-120, 121-122, 121f, 122f. See also Replication recombinant methods of. 139-157. See also Recombinant DNA technology telomeric, 836-837, 836f, 837f template strand of, 121-122, 122f in replication, 121-122, 122f, 820-821 in transcription, 124, 125f, 126, 126t, 854,855f topoisomerases and, 828, 828-831, 829f, 830f topoisomers of, 826, 828f unwinding of, 826-831, 858, 858f in transcription, 858, 858f Watson-Crick model of, 5, 113-115, 114f. 819-820. See also Double helix X-ray diffraction patterns of, 110, 113f Z-form, 116, 116f, 116t DNA amplification, polymerase chain reaction in, 145–146, 145–147 DNA blots, 142, 142f DNA fingerprint, 141 DNA fragments amplification of, 145-146 joining of, 148-149, 149f production of, 141-142 separation and visualization of, 141-142 DNA gyrase, 831 DNA ligase, 148-149, 823f, 824, 841 DNA linker, 148-149, 149f DNA microarrays, 162-163, 162f DNA mismatch repair, 840, 840f. See also DNA repair DNA photolyase, 840

DNA polymerase(s), 121-122, 121f, 821-823 bacterial, 835t classification of, 821 deoxyribonucleoside triphosphate binding by, 822, 823f error-prone, 835t, 837-838 eukaryotic, 835-836, 835t Klenow fragment of, 821, 821f in leading/lagging strand synthesis, 832-833, 832f, 833f metal ions of, 821-822, 821f primer for, 821, 821b, 823, 823f in prokarvotes, 831–834 in proofreading, 839–840, 839f reaction mechanism of, 821-822, 821f specificity of, 221, 822 structure of. 821. 821 types of, 835t DNA polymerase α, 835, 835t DNA polymerase β , 835, 835t DNA polymerase δ , 835, 835t DNA polymerase I, 834 DNA polymerase II, 835t DNA polymerase III, 831, 831–832, 832–833 sliding clamp unit of, 831-832, 831f, 834 DNA polymerase III holoenzyme, 832-833, 833f, 834 DNA polymerase switching, 835 DNA probes, 80, 142, 144-145 generation of, 144-145, 151-153 solid-phase approach in, 144–145, 144f for genomic library, 151–153, 152f DNA recombination, 844-846 definition of. 844 functions of, 844 Holliday junctions in, 845-846, 845f initiation of, 844-845 mechanisms of, 844-846, 845f *RecA* in, 845 recombinases in, 845–846 DNA recombination synapse, 846 DNA repair, 122, 820, 837-844 base-excision, 840 defective, in cancer, 842-843 direct, 840 double-strand, 841 enzyme complexes in, 840, 840f glycolases in, 840, 841 ligase in, 824, 841 mismatch, 840, 840f nonhomologous end joining in, 841 nucleotide-excision, 840-841, 841f proofreading in, 839-840 single-strand, 839-841 tumor-suppressor genes in, 842 uracil DNA glycolase in, 841, 841f DNA replication, 5-6, 6f, 119-121, 819, 820-837 in bacteria. 831-834 base complementarity in, 822, 822f, 823f cell cycle and, 836, 836f clamp loaders in, 832 controlled termination, 143-144, 143f

coordinated processes in, 831-837 cross-linkage in, 838, 839f definition of, 113 directionality of, 112, 122, 823, 832 DNA polymerase III holoenzyme in, 832-833, 833f, 834 DNA polymerases in, 121-122, 121f, 821-823, 835-836, 835t. See also DNA polymerase(s) DNA probes in, 145 DnaA in, 834, 834f in E. coli, 831-834 errors in, 837-838 repair of, 837-844. See also DNA repair in eukarvotes, 835-836 helicases in, 120, 824, 824-825, 824f, 825f, 832 initiation of. 823, 834-836 in bacteria, 834, 834f in eukaryotes, 835-836 sites of, 834-836 lagging strand in, 823f, 824 synthesis of, 823f, 824, 832-833, 833f leading strand in, 823f, 824 synthesis of, 823f, 824, 832-833, 833f licensing factors in, 835 ligase in, 823f, 824 Okazaki fragments in, 823-824, 823f, 833 origin of, 834, 834f origin of replication complexes in, 835 prepriming complex in, 834, 834f primer in, 122, 145, 821, 821b, 823, 823f processivity in, 831-832, 831f proofreading in, 839-840, 839f rate of, 831-832 recombinant DNA technology in, 145. See also Recombinant DNA technology replication fork in, 823, 823f, 832-833, 833f RNA polymerase in, 823, 823f semiconservative, 118-120, 120f sites of in bacteria. 834. 834f in eukarvotes. 835-836 sliding DNA clamp in, 831-832, 831f specificity of, 822, 822f, 823f strand joining in, 823f, 824 strand separation in, 120, 824-825, 825f telomeres in, 836-837, 836f, 837f template in, 121-122, 122f, 820-821 trombone model of, 833, 833f DNA sequencing, 143-144, 143f in amino acid sequencing, 84 chain-termination method in, 143, 143f fluorescence detection in, 143-144, 143f in forensics, 147, 147f for Neanderthals, 188-189, 189f Sanger dideoxy method in, 143-144, 143f DNA transfer. See also Recombinant DNA technology by electroporation, 167, 167f gene guns for, 167

C15 Index

by microinjection, 163, 163f vectors for, 149-151, 149f, 150f, 154-155 DNA vectors, 149-151, 149f-151f DnaA, assembly of, 834, 834f DNA-binding domains, 941-942, 943 in eukarvotes, 943 in prokaryotes, 942. See also DNAbinding proteins DNA-binding proteins. See also Transcription factors basic-leucine zipper in, 942 homeodomains in, 942 match with regulatory site, 922-923 in prokaryotes, 922-928, 922f, 923-928, 923f transcription inhibition by, 924-927 DNA-binding sites chromatin and, 941-942, 943 evolution of, 927 hypersensitive, 944 Dolichol phosphate, 334, 334 Domains DNA-binding, 941-942, 943 homeodomains, 943 immunoglobulin, 981-982 of living organisms, 3, 3f protein, 47, 47f exon encoding of, 133, 133f Dopamine, 1038 Double helix, 5, 5f, 26f, 109, 113-121 A, 115–116, 116t B, 115–116, 116t base pairing in, 5, 5f, 10-11, 10f, 113–115, **114**, 114t. See also Bases/base pairs discovery of, 5, 113-114 disruption of, in acid-base reactions, 14-15, 14f formation of, 6, 6f electrostatic interactions in, 10 heat released in, 12-13, 13f hydrogen bonds in, 10 hydrophobic interactions in, 10 van der Waals forces in, 10 hydrogen bonds in, 5, 5f, 10, 114, 118f left-handed, in Z-DNA, 116, 116t melting of, 120-121, 120f, 824-825, 825f in replication, 118-121 unwinding of, 826, 828-829 supercoiling and, 826 topoisomerases in, 828-831, 829f, 830f in transcription, 858, 858f Z, 116, 116t Double-displacement reactions, 237 Double-reciprocal plots, 233, 233f, 240, 240f Dreyer, William, 987 Drosophila melanogaster alternative splicing in, 879 sensory bristles in, 972 Toll receptor in, 978–979 Drug(s). See also specific drugs absorption of, 1031-1033, 1032f ADME properties of, 1031-1036, 1032f

agonist, 948 antagonist, 948 concentration of, 1031 distribution of, 1033, 1033f excretion of, 1035-1036 immune-modulating, 990 metabolism of, 1034-1036 cytochrome P450 in, 783 receptors for, 948, 949f resistance to, 1050-1051 response to, genetic variations in, 1047-1048 routes of administration for, 1032 side effects of, 1031, 1036 genetic variations in, 1047-1048 therapeutic index of, 1036 Drug development, 1001–1052 animal testing in, 1037 candidate drugs in absorption of, 1031-1033, 1032f ADME properties of, 1031–1036, 1032f distribution of, 1003f, 1033 effective concentrations of, 1031, 1031f essential characteristics of, 1030-1037 ligand binding and, 1030-1031, 1030f metabolism and excretion of, 1034-1036 number of, 1041-1042 oral bioavailability of, 1032-1033, 1032f potency of, 1030-1031 routes of administration for, 1032 side effects of, 1031, 1036-1037 target compartments of, 1003f, 1033 therapeutic index of, 1036 clinical trials in, 1048-1049, 1049f combinatorial chemistry in, 1041 drug resistance and, 1050-1051 dual pathways for, 1029-1030, 1030f genetic variations and, 1047-1048 genomics in, 1045-1048 high-throughput screening in, 1041 phases of, 1048-1049 screening libraries in, 1039-1042, 1042f serendipitous observation in, 1037-1038 7TM receptors in. 404. 1045 split-pool synthesis in, 1041, 1042f structure-based, 1042-1045, 1043f, 1044f Dynamic instability, 1019 Dynein, 1008 ATP binding to, 1010, 1011f structure of, 1010, 1011f E. coli. See Escherichia coli E site, ribosomal, 900, 900f, 904 E'_0 (oxidation-reduction potential), 528–531 Ear, hair cells of, 971-973, 971f, 972f

Early humans, DNA sequencing for,

EcoRV endonuclease, 272, 273-277,

recognition site of, 275-277, 275f

188-189, 189f

binding affinity of, 275

EcoRI endonuclease, 278

274f-277f

Editing, RNA, 852

Edman degradation, 81-83, 81f, 82f, 93 protein cleavage for, 82-83, 82f Edman, Pehr, 81 EF hand, 410-411, 411f Effector functions, 981 EGF. See Epidermal growth factor (EGF) Eicosanoids, 664-665, 665, 665f Elastase, catalytic triad in, 261, 261f Elastase inhibitor, 307 Electrochemical potential. See Membrane potential Electrocyclic reactions, 720 Electromagnetic spectrum, 966f Electron carriers. See also Transporters in metabolism, 438-439, 438f, 439f in oxidative phosphorylation, 438-439, 438f, 439f, 531-543, 532t, 551f, 552-553, 552f, 554f. See also Respiratory chain in photosynthesis, 568-572, 569f, 571f, 572f Electron sink, 605, 683 Electron transfer. See also Electrontransport chain in oxidative phosphorylation, 528-531, 542-543. See also Respiratory chain rate of, 542f in photosynthesis, 568-572, 569f, 571f, 572-577, 572f Electron-density maps, 99-100, 100f Electronic nose, 961-962, 962f Electron-transfer potential, 528 in oxidative phosphorylation, 528-530 Electron-transferring flavoprotein (ETF), 646 Electron-transport chain, 498, 512, 528 electron flow through, 540f inhibition of, 558, 582f rate of, 555-556 in oxidative phosphorylation, 438–439, 438f, 439f, 531-543, 532t, 540f. See also Respiratory chain in photosynthesis, 568-572 Electrophilic catalyst, 683 Electrophoresis gel, 71-73, 72f, 73f, 141-142, 142f in restriction fragment separation, 141-142, 142f SDS, 73, 78 SDS-polyacrylamide, of membrane proteins, 355, 355f two-dimensional, 74, 74f Electroporation, 167, 167f Electrospray ionization (ESI), 91-92 Electrostatic interactions, 7-8. See also Bonds in antigen-antibody binding, 984, 985, 985f ATP and, 433 ELISA (enzyme-linked immunosorbent assay), 88-89, 89f Elongation factor(s), 902-903, 903f G, 904-905, 904-906, 904f Ts, 903 Tu, 902-903, 903f

C16

INDEX

Embden, Gustav, 454 Embden-Meyerhof pathway, 454. See also Glycolysis Emphysema, 307 Enantiomers, 321, 321f Endergonic reactions, 222 Endocytosis, receptor-mediated, 365f, 366, 775-776, 776f Endonucleases. See Restriction enzymes (endonucleases) Endoplasmic reticulum, 365, 979f in diabetes, 802 fatty acid synthesis in. 664 protein glycosylation in, 333-334, 333f protein synthesis in, 911–914 protein targeting from, 911-914 ribosome binding to, 911-914 rough, 911 smooth, 911 Endoplasmic reticulum (ER) stress, 802 Endosymbiosis, 365, 527 Endotoxin, 978 End-product inhibition. See Feedback inhibition Enediol intermediate, in glycolysis, 459, 460f Energy. See also Bioenergetics; Thermodynamics activation, 222, 226 from ATP hydrolysis, 429-435 binding, 229 definition of, 254 enzyme specificity and, 275-277 body requirements for, 549b from carbon oxidation, 435-437, 435-438, 435f, 437f from catabolism, 434, 435-438, 435f, 437f enzymatic transformation of, 221-222 free. See Free energy glycogen storage of, 328, 615, 629-630 kinetic. 11 laws of thermodynamics and, 11-13, 222 lipid storage of, 641-642 from oxidative phosphorylation, 437-438 potential, 11 of reactions, 222-225 units of, 7, 223b Energy charge, 446-447, 446f Energy homeostasis. See Caloric homeostasis Enhancers, 127, 868-869, 943-944, 943f in cancer, 869 Enol phosphate, 465 Enolase, in glycolysis, 465 5-Enolpyruvylshikimate 3-phosphate, 720 Enoyl CoA, 646-647, 647, 648, 649 Enoyl CoA hydratase, 646-647 Ensemble studies, 246, 246f Enterohepatic cycling, 1036, 1036f Enteropeptidase, in trypsin activation, 305, 306f Enthalpy, 11–12 Entropy, 11–12

Envelope, nuclear, 365 Enzymatic cascades, 307 in amino acid synthesis, 725–726, 725f, 726f in blood clotting, 307-308, 308f Enzymatic cleavage, of proteins, 82-83, 82f Enzyme(s), 219-248 active sites of, 185–186, 227–228, 227f, 228f convergent evolution of, 185-186 definition of, 185-186 mapping of, 241-242 affinity tags for, 241–242 allosteric, 237, 237f, 289, 290-296. See also Allosteric enzymes amounts of, regulation of, 446 apoenzymes, 221 bifunctional, 487 catalytic activity of, 205-208. See also Catalysis assays for, 67. See also Protein purification specific, 67 catalytic efficiency of, 235, 235t catalytically perfect, 234, 235, 460 classification of, 248-249 coenzymes. See Coenzyme(s) cofactors for, 221, 221t core. 856 covalent modification of, 57-58, 57f, 297-302, 298t definition of. 26, 219 denatured, 49-50, 49f, 50f digestive, 255-260, 303-307, 303t, 304f, 674, 674f distributive, 832b energy-transducing, 221–222 holoenzymes, 221 inhibition of, 238–245 affinity tags in, 241-242, 241f competitive, 238-240, 238f, 239f, 240f feedback, 291, 446. See also Feedback inhibition group-specific agents in, 241f group-specific reagents in, 241 irreversible, 238, 238f, 241-242 noncompetitive, 230-240, 238, 238f, 239-240. 239f. 240f reversible, 238, 238f suicide (mechanism-based), 242, 750.754 transition-state analogs in, 243, 243f uncompetitive, 238, 238f, 239, 239f, 240, 240f isozymes, 289–290, 296–297, 297f kinetic properties of, 229-237. See also Enzyme kinetics kinetically perfect, 234, 235, 460 in linked pathways, 744. See also Metabolic pathways in multienzyme complexes, 660–661 nomenclature for, 248-249 overview of, 220-222

processive definition of, 832b in DNA replication, 831-832 prosthetic groups of, 221 proteolytic, 220-221, 302-313. See also Proteolytic enzymes reaction equilibrium and, 224-225 reaction rate acceleration by, 220, 220t, 225-229, 226f restriction, 141-142, 141f, 253, 271-279. See also Restriction enzymes specificity of, 220-221, 221f, 275-277 binding energy and, 275-277 substrate binding by. See also Enzymesubstrate complex concerted mechanism in, 294-296 cooperative, 200-201, 201f, 294 energy for, 229, 254, 275-277. See also Binding energy sequential model of, 294 specificity of, 220-221, 221f, 275-277 substrates of, 220. See also Enzymesubstrate complex in transition state formation, 225–226, 225f, 227-228 turnover number of, 234, 234t Enzyme cofactors. See Coenzyme(s) Enzyme kinetics, 229-237 definition of, 229 in first-order reactions, 229-230 fraction of active sites filled ($f_{\rm FS}$) in, 234 initial velocity and, 230, 230f k_{cat}/K_M, 234–235 kinetic perfection and, 234, 235, 460 Lineweaver-Burk plot of, 233, 233f Michaelis constant (K_M) in, 231-235, 233t Michaelis-Menten, 229-237, 231f in pseudo-first-order reactions, 230 in second-order reactions, 230 sigmoidal, 200, 291, 291f, 294, 295f Enzyme multiplicity, 724, 724f Enzyme regulation, 289-314 allosteric, 289, 290-296 dephosphorylation in, 300 isozymes in, 289–290, 296–297, 297f phosphorylation in, 298-301 protein kinases in, 298-301 proteolytic activation in, 290, 302-313 reversible covalent modification in, 290, 297-392 zymogens in, 303-313 Enzymes ensemble studies of, 246, 246f single molecule studies of, 246, 246f Enzyme-substrate complex, 226-227, 227, 227f attractive forces in, 228, 228f cooperative binding and, 200-201, 201f, 203f. 294 dissociation constant for, 233-234 in double-displacement (ping-pong) reactions, 237

C17 Index

induced-fit model of, 228, 228f, 254, 457, 457f lock-and-key model of, 228, 228f in sequential reactions, 236-237 Epidermal growth factor (EGF), in signal transduction, 402f, 415-418, 416f-418f Epidermal growth factor receptor, 415-418, 416f-418f in cancer, 421 Epigenetic factors, 20 Epigenome, 938 Epilepsy, ketogenic diet for, 656 Epimers, 321f, 322 Epinephrine, 403 in fatty acid metabolism, 643, 666-667 in glycogen metabolism, 624-626 in signal transduction, 402f, 403-410, 406f. See also Signal transduction synthesis of, 726, 726f Epitopes, 85, 85f, 980 EPO (erythropoietin), 331, 331f ε chains, 982, 983f Equilibrium constant for dissociation of water, 14 free-energy change and, 222-225, 224t Equilibrium potential, 391-392, 391f Erbitux, 421 Error-prone DNA polymerase, 835t, 838 Erythrocytes. See Red blood cells Erythromycin, 909t, 910 Erythropoietin (EPO), 331, 331f Erythrose 4-phosphate, 603 in amino acid synthesis, 719, 720f in Calvin cycle, 595 in pentose phosphate pathway, 602f, 603, 604f Escherichia coli. See also Bacteria DNA cloning and expression in, 154-155 DNA repair in, 839-840 DNA replication in, 831-834. See also DNA replication flagella of, 1022-1025 gene expression in, 922-928, 923-928. See also Gene expression/regulation, in prokaryotes genome of, 113, 113f, 182f, 928, 937 lac operon of, 922, 925-928, 925f, 926f lactose permease symporter in, 381, 381f pyruvate dehydrogenase complex in, 499f, 499t replication in, 831-834 restriction enzymes of, 272-279, 274f-277f ESI (electrospray ionization), 91–92 Essential amino acids, 674, 674t, 711, 711t, 712f synthesis of, 719-723. See also Amino acid synthesis Essential fatty acids, 664 Estradiol, 784 synthesis of, 785

Estrogen(s). See also Sex hormones; Steroid hormones regulation of, 946 synthesis of, 780, 780f, 784, 785 Estrogen receptors, 946 drug binding to, 948, 949f Estrogen response elements (EREs), 946 Estrone, synthesis of, 780, 780f, 784, 785 ETF (electron-transferring flavoprotein), 646 ETF : ubiquinone reductase, 646 Ethanol. See also Alcohol(s) fermentation of, 466-467, 467f formation of from pyruvate, 466-467, 467f in glycolysis, 453 metabolism of, 232, 783, 810-813 toxicity of, 811-813 Ethanolamine, 349, 763, 763f Ethylene, synthesis of, 718, 718f Eukarva, 3 Eukaryotes, 3, 3f cell types in, 938, 939t Eversion, 376 Evolution, 3, 3f of amino acid sequences, 36, 79, 174-181 of blood types, 335-336 caloric homeostasis and, 792-793 of Calvin cycle, 600 of carbonic anhydrases, 271 of chloroplasts, 568 of citric acid cycle, 518 convergent, 185-186, 185f of cytochrome c, 543, 543, 543f of cytochrome P450, 783 divergent, 185 of DNA binding sites, 927 endosymbiotic events in, 527, 568 essential elements of, 189 experimental studies of ancient DNA amplification and sequencing in, 188–189 combinatorial chemistry in, 188-189 molecular, 188-191 of feedback inhibition, 724 of glucose metabolism, 490-491 of glycogen metabolism, 627 horizontal gene transfer in, 278-279 of immune system, 978-981, 984 of immunoglobulin fold, 984 of introns and exons, 133 of ion pumps, 374, 378-379 of metabolic pathways, 447 of microRNA, 954 of mitochondria, 527 obesity and, 792-793 of olfaction, 959-960, 959f of photoreceptors, 970, 970f of photosynthesis, 568, 584, 600 of proofreading, 895 of proteasomes, 677-678, 678f, 679 of protein degradation pathways, 677-678 of proteins, 133 of P-type ATPases, 374, 378-379

of restriction enzymes, 278-279 of ribosomes, 900 RNA catalysis and, 881 of rubisco, 600 sequence homology and, 174-181, 176f-182f. See also Sequence homologies signal transduction pathways, 419 split genes in, 133 of succinvl coenzyme A, 509 tertiary structure and, 183-184 time line for, 3f of transaminases, 713 of transcription, 869 of urea cycle, 688 of vision, 970, 970f Evolutionary trees, 187-188, 187f Exercise. See also Muscle contraction ATP in, 434-435, 435f, 805-806, 805t caloric homeostasis and, 803-806 carbo-loading for, 806 citric acid cycle during, 514 creatine phosphate in, 434-435, 435f, 805, 805t fatty acid metabolism in, 804, 804f, 806 fuel sources for, 805-806, 805t gene expression and, 804 gluconeogenesis during, 489-490, 490f glycolysis during, 473-474, 475f, 478-479, 489-490 lactate in, 489-490, 490f, 805 mitochondrial biogenesis and, 804, 804f oxidative phosphorylation in, 805-806 superoxide dismutase and, 542 Exergonic reactions, 222 Exon(s), 110, 132, 133 evolution of, 133 Exon shuffling, 133, 133f Exonuclease(s), 821 in DNA polymerase III holoenzymes, 832, 833f in proofreading, 839-840, 839f Expression cloning, 154-155, 155f Expression vectors, 149, 154–155 External aldimine, 682, 682 in amino acid degradation, 682, 682f in amino acid synthesis, 712f, 713 Extrinsic pathway, coagulation, 308, 308f F₀F₁₋ATPase. See ATP synthase Fab fragments, 981-982, 982f in antigen-antibody binding, 985f, 986, 986f Facilitated diffusion, 371, 373 F-actin, 1013, 1013f Factor VIII, 311, 311f Factor XIII_a, 309 FAD (flavin adenine dinucleotide) ADP units in, 447, 447f in catabolism, 438-439, 439, 439f in citric acid cycle, 498, 498f, 501-502, 509-510, 512 in fatty acid oxidation, 646, 647f

FADH₂ (flavin adenine dinucleotide reduced) in catabolism, 438-439, 439 in citric acid cycle, 498, 503, 510, 512 in fatty acid oxidation, 646-647 in oxidative phosphorylation, 505, 535 electron-transfer potential of, 530-531 reduction potential of, 528-530 Familial hypercholesterolemia, 759, 776-779 Farnesyl pyrophosphate, 769, 769 Farnesvlation, 298t 5-Farnesylcysteine methyl ester, 359, 359 Fasting, 796 Fat body, energy storage in, 641 brown, 556-557, 557f fecal, 642 neutral. See Triacylglycerol(s) white, 556 Fat cells, 641 energy storage in, 641 Fatty acid(s), 346-348, 347f, 347t carbon atoms in, 347-348, 347f chain length in, 437-438 covalent attachment of, 639 essential, 664 examples of, 347t functions of, 639 in lipid membrane, 362-363, 363f in muscle during exercise, 804, 804f, 806 in obesity, 800-801 odd-chain, 649-650, 650f polyunsaturated, eicosanoids from, 664-665, 665, 665f saturation of, 348 storage form of, 648-649 structure of, 346-348, 347, 362-363, 363f synthesis of, 643-644, 643f, 644f unsaturated, 648-649, 664-665 Fatty acid metabolism, 437f, 639–669 acetyl CoA carboxylase in, 657, 658t, 666-667,666f acetyl CoA in, 646-647, 657 activation in. 644-645 acyl carnitine in, 645-646, 646f acyl carrier proteins in, 657-658, 658, 658t, 659f in animals vs. plants, 656 β-oxidation pathway in, 646–647 chylomicrons in, 642, 642f citrate in, 662, 662f, 666, 667f coenzyme A in, 644-645 coenzyme B₁₂ in, 651–652 cytoplasmic, 657 degradation in, 640f, 646-657, 666-667 reactions in, 647f, 647t vs. synthesis, 657, 658t diet and, 667 in endoplasmic reticulum, 664 energy yield from, 641, 647-648 enoyl CoA hydration in, 646-647, 647f, 647t epinephrine in, 643, 666-667

ethanol and, 811 during exercise, 804, 804f, 806 fatty acid synthase in, 658–659, 660f. 661f. 663 fatty acid unsaturation in, 664–665 glucagon in, 643, 666-667 in glucose homeostasis, 808 hormones in, 643, 666-667 ketone bodies in, 653-656 lipases in, 641-642, 642f, 643 lipid mobilization and transport in, 642-644, 642f, 643f lipolysis in. 643-644, 644f malonyl CoA in. 657, 658t, 659, 666 in muscle, 804, 804f oxidation in, 642-656, 647f, 647t palmitoyl CoA in, 647-648, 666 pancreatic lipases in, 641-642, 642f peroxisomal, 652, 653f reactions of, 640-641, 640f regulation of, 666-667 in starvation, 809, 809f, 810 synthesis in, 640-641, 640f, 643-644, 643f, 644f, 656-657 in animals. 659-661 citrate carriers in, 662, 662f condensation in, 658-659, 658t, 659f coordinated metabolic pathways in, 654f. 663. 666f dehydration in, 658t, 659, 659f elongation in, 658, 658t, 659f, 664-665 insulin in. 667 kev features of, 657 NADPH sources for, 662-663 reactions in, 658-659, 658t, 659f reduction in, 658t, 659, 659f of unsaturated fatty acids, 664-665, 665f vs. degradation, 657, 658t thiolysis in, 647-648, 647f, 647t triacylglycerol hydrolysis in, 643-644, 643f of unsaturated fatty acids, 648-649, 648f, 664-665 Fatty acid synthase, 656-663, 660f in animals, 659-661, 660f, 661f in cancer. 663 Fatty acid synthase inhibitors, 663 Fatty acid thiokinase, 644-645 Fatty acyl CoA dehydrogenase, 646 in oxidative phosphorylation, 535 F_c fragments, 981 FDA (Food and Drug Administration), 1049 Fe protein, in nitrogen fixation, 707, 707 Feedback inhibition, 291, 446 in amino acid synthesis, 723–726 in cholesterol synthesis, 770 cumulative, 725 enzyme multiplicity and, 724, 724f evolution of. 724 in nucleotide synthesis, 751 Feedforward stimulation, 476 Fehling's solution, 325 FeMo cofactor, 708, 708-709

Fermentation, 466-468 to alcohol, 453, 466-467, 467f as anaerobic process, 468-469 in glycolysis, 453 to lactic acid, 453, 467-468 start/end points for, 469t Ferredoxin, 575-577, 576 in nitrogen fixation, 707 Ferredoxin-NADP⁺ reductase, 576, 576-577, 577f Ferredoxin-thioredoxin reductase, 599 Ferritin, 730, 951, 951–952 Ferrochelatase, 730 Fetal alcohol syndrome, 812 Fetal hemoglobin. See also Hemoglobin oxygen affinity of, 205, 205f Fiber, dietary, 329 Fibrin clot formation of, 308-309, 308f lysis of, 312 Fibrin, structure of, 309, 309f Fibrinogen conversion to thrombin, 308-309 structure of, 308, 308f sulfation of, 298t Fibrinopeptides, 308 Fibrous proteins, 43-45 50S subunit, of ribosome, 897-898 Fight or flight response, 301 Filters, selectivity, 385-386 Fingerprint, DNA, 141 First Law of Thermodynamics, 11. See also Thermodynamics First-order reactions, rate of, 229-230 First-pass metabolism, 1035 Fischer, Emil, 228, 1042 Fischer projections, 21, 22f, 23f 5' cap, 870-871, 871f 5' splice sites, 870-871, 871f, 873-874, 873f Flagella microtubules in, 1019 rotary movement of, 1022-1025 structure of, 1022, 1022f, 1023f Flagellin, 1022, 1022f Flavin adenine dinucleotide. See FAD Flavin mononucleotide, 533-534, 534f, 860f Flavoproteins, 502 electron-transferring, 646 Fleming, Alexander, 1037 FliG, 1023, 1023f FliM, 1023 FliN, 1023 Flip-flop, 362, 362f Flippases, 378 Florey, Howard, 1037 Fluconazole, 1033 distribution of, 1003f, 1033f Fluid mosaic model, 362 Fluorescamine, in Edman degradation, 81.81 Fluorescence detection, in DNA sequencing, 143-144, 143f Fluorescence microscopy, 90-91, 90f

C19 Index

Fluorescence recovery after photobleaching (FRAP), 361, 361f Fluorouracil, 749-750 fMET. See Formylmethionine (fMet) fMRI. See Functional magnetic resonance imaging Folding funnel, 54f Folding, protein. See Protein(s), folding of Folic acid, 441t deficiency of, spina bifida and, 755 Food and Drug Administration (FDA), 1049 Food intake. See also Diet; Starvation starved-fed cycle and, 807-808 Food Pyramid, 20f Food, spicy, capsaicin in, 973-974, 973f 5-Foraminoimidazole-4-carboxamide ribonucleotide, 742f Forensics, polymerase chain reaction in, 147.147f Formamide, molecular models of, 23f N⁵-Formiminotetrahydrofolate, 715, 716 Formylglycinamide ribonucleotide, 741, 742, 742, 742f Formylkynurenine, 696, 696 Formylmethionine (fMET), 130, 130, 901-902 Formylmethionyl-tRNA, 901-902, 902, 902f N^{10} -Formyltetrahydrofolate, 715–716, 742f Fourier transform, 99-100 Frameshift mutations, tests for, 843 Franklin, Rosalind, 113 FRAP (fluorescence recovery after photobleaching), 361, 361f Free energy, 12, 222-225, 226. See also Energy of activation, 222, 226 ATP and, 430-431 binding, 229, 254 from carbon oxidation, 435-437, 435-438, 435f in concentration gradients, 373-374, 437 equilibrium constant and, 222-225, 224t Gibbs, 12, 226 in membrane transport, 373-374, 373f in metabolism, 429-430 from proton gradients, 559-560, 560f in redox reactions, 530 reduction potential and, 528-530 of translation, 893-894 Free radicals. See Reactive oxygen species (ROS) Fructofuranose, 323, 323f, 324f Fructokinase, 469 Fructopyranose, 324f Fructose, 322, 322f in glycolysis, 469-470, 469f, 470f metabolism of, 469-470, 470f open-chain form of, 323f ring form of, 323, 324f structure of, 322f Fructose 1,6-bisphosphatase, in

gluconeogenesis, 482

in Calvin cycle, 594, 594f in gluconeogenesis, 482, 484 in glycolysis, 456f, 458, 460 Fructose 1-phosphate, in glycolysis, 469-470, 469f, 487 Fructose 2,6-bisphosphate, 487, 488f Fructose 6-phosphate, 457 in Calvin cycle, 594, 594f, 595f, 596f in gluconeogenesis, 482, 484-485, 487 in glycolysis, 455f, 456f, 458, 458f, 459, 470, 487 in pentose phosphate pathway, 602f, 603, 604f in sucrose synthesis, 597, 598 Fructose bisphosphatase 2, 487 FtsZ, 1019 Fuels. See Carbon fuels Fumarase, 510 Fumarate, 510, 687, 687f in citric acid cycle, 510 from phenylalanine, 696, 696f in purine synthesis, 742f, 743, 743f in urea cycle, 687 4-Fumarylacetoacetate, 696, 696 Functional groups, of proteins, 26 Functional magnetic resonance imaging, 197-198, 198f Furanose, 323-325, 323f envelope forms of, 325, 325f Futile cycles, 489 G_(olf), 959, 960 G elongation factor, 904–905,

Fructose 1,6-bisphosphate, 458

904-906, 904f G protein(s), 405-406 activation of, 406 ATP synthase and, 550 in cholera, 421-422 definition of, 405 in glycogen metabolism, 624–626 heterotrimeric, 405-406 in olfaction, 959 P-loop NTPases in, 405 resetting of, 407-408 in signal transduction, 405-411. See also Signal transduction small, 418, 418t structure of, 405, 405 subunits of, 405 in taste, 963–965 of bitterness, 962, 963-964 of sweetness, 965–966 in translation, 902-903 in vision, 968, 968f in whooping cough, 422 G6PD. See Glucose 6-phosphate dehydrogenase (G6PD) G-actin. 1013 GAL4 binding sites, 945, 945f Galactitol, 472 Galactokinase, 470 Galactolipids, in thylakoid membrane, 567

Galactose, 321-322, 322f in blood groups, 335 disorders of, 471-472 in glycolysis, 469f, 470-471 Galactose 1-phosphate, 470, 470 Galactose 1-phosphate uridyl transferase, 470 Galactose 6-phosphate, 470 Galactose-glucose interconversion pathway, 470-471 Galactosemia, 472 β-Galactosidase, 328, 924, 926 in lactose metabolism, 924-925 Galactoside permease, 924 Galactouronic acid, 329, 329f v chain(s), 982, 983f γ chain(s), of fetal hemoglobin, 205 Gangliosides, 350 disorders of. 765-766 structure of, 738, 765f synthesis of, 764-765 GAP. See Glyceraldehyde 3-phosphate Gap junctions, 372, 393-394, 393f, 394f GAPs (GTPase activating proteins), 418 Gargoylism, 332, 332f Garrod, Archibald, 697 Gastric enzymes, 674, 674f Gastric H⁺-K⁺ ATPase, 374 Gastric zymogens, 303t GC box, 867, 867f GDP (guanosine diphosphate) in citric acid cycle, 508 hydrolysis of, 431 in olfaction, 959, 960 in signal transduction, 405, 418, 418f in translation, 902-903, 908 in tubulin, 1019 in vision, 968, 968f GEF (guanine-nucleotide-exchange factor), 417 Gel electrophoresis, 71–73, 72f, 73f, 141–142, 142f in restriction fragment separation, 141-142, 142f SDS, 73, 78 SDS-polyacrylamide, of membrane proteins, 355, 355f Gel-filtration chromatography, 69, 69f, 75, 75t Gellert, Martin, 828 Gene(s) comparative analysis of, 160-161, 160f continuous, 131, 132 definition of, 19 designer, 157 discontinuous, 131 discontinuous (split), 131–132, 132f, 133f horizontal transfer of, 278-279 identification of, in genome sequencing, 17 - 18number in genome, 159 protein-encoding, 19 synthetic, 157 tumor suppressor, 420

C20

INDEX

Gene chips (microarrays), 162–163, 162f Gene disruption, 164–165, 165f Gene expression studies, 161-167 cDNA in, 154-155 DNA microinjection in, 163, 163f expression vectors in, 154-155 gene disruption, 164-165 gene-expression analysis in, 161-162, 162f recombinant DNA technology in. See Recombinant DNA technology RNA interference in, 165–166, 167f transgenic animals in, 164, 164f viral vectors in, 163-164 Gene expression/regulation, 109–110, 123 - 128basic-leucine zipper in, 942 constitutive, 921 definition of. 921 in eukaryotes, 937-955 chromatin remodeling in, 944-951, 949-951 coactivators in, 947-948, 948f, 949-951 DNA methylation and, 945-946 enhancers in, 127, 868-869, 943-944 histones in, 939-941, 940f, 949-951,950f in liver, 939t methylation in. 945-946 nuclear hormone receptors in, 946 nucleosomes in, 939-941, 940f overview of, 937-938 in pancreas, 939t tissue-specific, 939t transcription factors in, 946-947. See also Transcription factors vs. in prokaryotes, 937–938 in prokaryotes, 921-933 attenuation in, 931-932, 932f biofilms and, 930-931 catabolite activator protein in, 927-928, 927f catabolite repression in, 927-928 corepressors in. 927 DNA-binding proteins in, 922–923, 922-928 match with regulatory site, 922-923 in prokaryotes, 922-928 genetic switching and, 928-931 helix-turn-helix motif in, 923, 923f lac operon in, 925-928, 925f, 926f. See also under lac levels of, 161-162 ligand binding in, 926 operon model of, 924-928 posttranscriptional, 931-933 pur repressor in, 927 quorum sensing in, 930-931, 930f regulatory sites in, 922-928, 922f RNA in, 123-128 signaling in, 929-930 social interactions in, 929-930 symmetry matching in, 922

transcriptional, 921-928. See also Transcription translational, 951-953 vs. in eukaryotes, 937-938 regulated, 921 transcriptional in eukaryotes, 941-944 in prokaryotes, 921-928 translational, 951–953. See also Translation Gene guns, 167 Gene knockout, 164-165, 165f Gene therapy, 167–168 Gene-expression analysis, 161–162, 162f. 163-164. See also Gene expression studies General acid-base catalysis, 254 Genetic code, 19, 128-131 definition of. 128 degeneracy of, 129, 130, 892 major features of, 129-130 of mitochondrial DNA, 131, 131t universality of, 131 variations of, 131 Genetic engineering. See Recombinant DNA technology Genetic imprinting, 20 Genetic information, flow of, 109-110 Genetic mutations. See Mutations Genetic switch, 928-931 Genetic variation, 19-20 Genome, 157-161, 159f-161f analysis of, 159-161 comparative, 160-161, 160f of Caenorhabditis elegans, 159 of chloroplasts, 568 definition of, 66 of Escherichia coli, 113, 113f, 928, 937 of Haemophilus influenzae, 158, 158f, 181 of Indian muntjak, 113, 113f mitochondrial, 527-528, 527f, 528f noncoding DNA in, 159–160 number of genes in, 159 proteome and, 66 of puffer fish, 161, 161f of Reclinomonas americana, 528 of Rickettsia prowazekii, 527-528 of Saccharomyces cerevisiae, 158, 937, 938f sequencing of, 17-18, 157-161, 158f next-generation methods for, 160 size of. 113 Genomic libraries, 151-153, 152f, 153f Genomics comparative, 160-161, 160f in drug development, 1045–1048 George III (King of England), porphyria in, 731 Geraniol, 958 Geranyl pyrophosphate, 769, 769 Geranyl transferase, 769, 769 Gerhart, John, 291 Ghrelin, 796 Gibbs free energy, 12, 226

Gibbs, Josiah Willard, 12 Gigaseal, 383 Gla domain, in prothrombin, 310, 310f Gleevec, 421 Globin(s). See also Hemoglobin; Myoglobin evolution of, 187-188, 187f sequence alignment in, 175-181, 176t-182t structure of, 48, 48f, 183, 183-184 types of, 211–212 Globin fold, 199 Globin genes, 211–212 Globins, 175 Glomeruli. See also Kidney drug metabolism in, 1035 Glucagon, 488 in fatty acid metabolism, 643, 666-667, 808 in glucose homeostasis, 807-808, 808 in glycogen metabolism, 624-626, 807-808.808 Glucagon-like peptide 1 (GLP1), in caloric homeostasis, 795, 795f Glucocorticoids. See also Steroid hormones synthesis of, 780, 780f, 783 Glucogenic amino acids, 690. See also Amino acid(s) Glucokinase, 807 Gluconeogenesis, 479-491 carboxybiotin in, 483 definition of, 453, 479 in diabetes, 803, 803f evolution of, 490-491 fructose 1,6-bisphosphate in, 480f, 482.484 fructose 6-phosphate in, 480f, 482, 484-485 glucose 6-phosphatase in, 480f, 484, 485f glucose 6-phosphate in, 480f, 484-485, 485f, 807 in glucose homeostasis, 807-810 glycolysis and, 481-482, 486-491 historical perspective on, 454 in kidney, 481 in liver. 481 orthophosphate in, 484 oxaloacetate in, 480f, 481, 482, 483-484 phosphoenolpyruvate in, 480f, 481, 482, 483-484 phosphorylation in, 485 reactions in, 479-485, 480f, 481-485 coupled, 485 regulation of, 486-491, 803, 803f sites of, 481 in starvation, 809 in starved-fed cycle, 807-810 stoichiometry of, 485 urea cycle and, 687 Glucopyranose, 323, 323f boat form of, 324, 325f chair form of, 324, 325f Glucose body stores/requirements for, 479 as cellular fuel, 455

C21 Index

in Cori cycle, 490, 490f in glycogen, 328, 328f, 615, 629-630 metabolism of, 437f. See also Gluconeogenesis; Glycolysis carbon fuels for, 808, 809 in diabetes mellitus, 798-800 evolution of, 490-491 historical perspective on, 454 homeostatic mechanisms in, 798-800. See also Glucose homeostasis in liver, 806 in muscle, 805-806 reactions in, 428f, 454f in starvation, 809 starved-fed cycle in, 807-808 open-chain form of, 323, 323f overview of, 453-454 properties of, 455 regulated release of, 616-617. See also Glycogen metabolism ring form of, 323, 323f storage forms of, 328, 328f, 329f, 615, 629 - 630structure of, 321-322, 321f-323f synthesis of from dietary carbohydrates, 454 from non carbohydrate precursors, 453-454, 454f, 479-491. See also Gluconeogenesis taste of, 962, 964-965 Glucose homeostasis, 807-810. See also Glucose, metabolism of in diabetes, 798-803, 803f in starvation, 809-810 starved-fed cycle in, 807-808 Glucose 1-phosphate, 617 in glycogen metabolism, 617–618 Glucose 6-phosphate, 326, 326f, 457 deficiency of, 634-635 in gluconeogenesis, 484-485, 485f, 807 in glycolysis, 456f, 457-458, 607-609, 807 metabolic fates of, 616f in pentose phosphate pathway, 601, 607-609 reactive oxygen species and, 609 Glucose 6-phosphate dehydrogenase (G6PD) antioxidant properties of, 609 deficiency of, 609-610, 611 in pentose phosphate pathway, 601, 603f Glucose 6-phosphate transporter, 635 Glucose 6-phosphatase, 455 in gluconeogenesis, 484, 485f in glycogen metabolism, 621 in glycolysis, 455-457, 456f Glucose transporters (GLUTs), 372, 477, 477t, 485, 799 in cancer, 479 insulin secretion and. 801-802 Glucose-alanine cycle, 684-685, 685f Glucose-stimulated insulin secretion, 801-802 α-Glucosidase, 454

Glucuronic acid, conjugation of, 1034-1035, 1035 Glutamate in amino acid degeneration, 680-681 arginine synthesis from, 714, 714f chirality of, 709, 710f conversion to ammonia, 680-681 glutamine synthesis from, 709-710, 710f in nitrogen fixation, 709 in nitrogen transport, 685, 685f oxidative deamination of, 680-681 proline synthesis from, 714, 714f structure of. 31, 32f. 237.680 synthesis of, 685, 709, 709f, 712-713 taste of, 962, 962f, 965 Glutamate dehydrogenase, 710 in amino acid degradation, 680-681 in glutamate synthesis, 709 in nitrogen fixation, 709 Glutamate synthase, 710 Glutamate synthetase, in nitrogen fixation, 709-710 Glutamic γ -semialdehyde, 714 in arginine synthesis, 714 in proline synthesis, 714 Glutamine in ammonia formation, 714 in nitrogen fixation, 709 in nitrogen transport, 685 in pyrimidine synthesis, 737 structure of. 30, 30f synthesis of, 709-710, 710f, 725-726 Glutamine phosphoribosyl aminotransferase, 741, 751 Glutamine synthetase, 709-710 regulation of, 725-726, 725f structure of, 725 Glutathione, 727, 727 antioxidant properties of, 609 conjugation of, 1030, 1035 Glutathione peroxidase, 542, 610, 727, 727 Glutathione reductase, 610, 727 Glutathione-drug conjugates, 1034, 1034-1035 GLUTs. See Glucose transporters (GLUTs) Glycan-binding proteins, 337-338 Glyceraldehyde, 320, 320, 321f in glycolysis, 461, 462f, 470 Glyceraldehyde 3-phosphate (GAP), 224, 326, 326f, 603 in Calvin cycle, 594 dihydroacetone phosphate isomerization to, 224, 224, 458-459, 458f, 460f in glycolysis, 224, 456f, 458-459, 458f, 460, 460f, 470 in lipolysis, 644 oxidation of, energy from, 436 in pentose phosphate pathway, 602, 602f, 603f, 604, 604f in sucrose synthesis, 597, 598

α-1,6-Glucosidase, 620, 620f

Glyceraldehyde 3-phosphate dehydrogenase, 481 in Calvin cycle, 594 in glycolysis, 461, 462f, 463f, 464 NAD⁺ binding sites in, 469, 469f Glycerol, 644 in gluconeogenesis, 481 metabolism of, 437f, 643-644, 644 in phospholipids, 348 Glycerol 3-phosphate, in membrane lipid synthesis, 760 Glycerol 3-phosphate shuttle, 551, 551f Glycerol phosphate acyltransferase, 760 Glycerol phosphate dehydrogenase, 551 in oxidative phosphorylation, 535 Glycinamide ribonucleotide, 742, 742f Glycine in Calvin cycle, 594 in collagen, 44-45, 44f heme labeling with, 728, 728b porphyrins from, 728-730, 729f serine formation from, 691 structure of, 28, 29f synthesis of, 714-715, 716 Glycine cleavage enzyme, 716 Glycine synthase, 716 Glycoaldehyde, in pentose phosphate pathway, 605, 605f Glycobiology, 320 Glycocholate, 642, 642 synthesis of, 780, 780f Glycoforms, 330 Glycogen, 328, 328f, 617 branching in, 615, 616f, 629 glucose storage in, 328, 328f, 615, 629-630 as homopolymer, 328 storage sites for, 616, 616f structure of, 328, 328f, 615, 616f, 629f Glycogen metabolism, 615–636 branching enzyme in, 629 carbonium ion intermediate in, 619 degradation in, 616-627 cAMP in, 626 initiation of, 624-626 regulatory cascade in, 624-626, 625f termination of, 626-627 degradative, 630-631 disorders of, 634-635, 634t epinephrine in, 624–626 evolution of, 627 during exercise, 805-806 G proteins in, 624–626 glucagon in, 624-626 α -1,6-glucosidase in, 620, 620f glycogen phosphorylase in, 617–624, 618, 620f, 633-634, 633f. See also Glycogen phosphorylase glycogen synthase in, 627–629, 632-633.633f glycogen synthase kinase in, 629, 630f, 631, 632-633, 632f glycogenin in, 628

INDEX

Glycogen metabolism—(continued) hormones in. 624-626, 632-633, 632f insulin in, 632-633, 632f overview of, 615-617 phosphoglucomutase in, 620, 643 phosphoglycerate mutase in, 620 phosphorolysis in, 617-621 phosphorylase in, 617-624. See also Glycogen phosphorylase phosphorylase kinase in, 623-624, 624f, 629,630f protein kinase A in, 629, 630f protein phosphatase 1 in, 626-627, 631-634, 631f pyridoxal phosphate in, 618-619, 619f pyrophosphate hydrolysis in, 627-628 as regulated release of glucose, 616-617 regulation of, 629-634 in liver, 621, 633-634, 633f in muscle, 631-633 signaling in, 624-626 synthesis in, 616-617, 627-635 transferase in, 620, 620f uridine diphosphate glucose in, 627-630 Glycogen phosphorylase, 617-624, 620f, 633-634, 633f a, 622-623, 644, 644f amino-terminal domain of, 618-619, 618f b. 622-623. 644. 644f carboxyl-terminal domain of, 618-619.618f catalytic mechanism of, 618-619 evolution of, 627 glycogen binding site of, 618-619, 618f regulation of, 621-624 structure of, 618, 618-619 T and R states of, 621–623 Glycogen storage diseases, 634-635, 634t Glycogen synthase, 627-629, 632-633, 633f Glycogen synthase kinase, 629, 630f, 631, 632-633, 632f, 799 Glycogen transferase, 620, 620f Glycogenin, 628 Glycolases, in DNA repair, 840, 841 Glycolate, in Calvin cycle, 593-594, 593f Glycolate oxidase, 594 Glycolipids, 350, 350 Glycolysis, 444, 453-479 aerobic, 478-479 aldehyde oxidation in, 461, 462f ATP formation in, 463–466 yield in, 554-555, 555t 1,3-bisphosphoglycerate in, 484 in cancer, 478-479 citric acid cycle and, 499-503, 499f committed step in, 474 definition of, 453 in diabetes, 803, 803f dihydroxyacetone phosphate in, 456f, 458-459, 458f, 460f, 469-470 enolase in, 465 evolution of, 490-491 during exercise, 478-479, 805-806

feedforward stimulation in, 476 fermentation in, 466-468 fructose 1,6-bisphosphate in, 456f, 457-458, 458f, 460 fructose 6-phosphate in, 456f, 458, 458f. 459 fructose in, 469-470, 469f galactose in, 469f, 470 gluconeogenesis and, 481-482, 486-491 glucose 6-phosphate in, 456f, 457-458, 459.807 in glucose homeostasis, 807-810 glucose transporters in, 477, 477t glucose trapping in, 455–457, 456f glyceraldehyde 3-phosphate dehydrogenase in, 460, 461, 462f, 463f glyceraldehyde 3-phosphate in, 224. 456f, 458-459, 458f, 460, 460f, 461.462f hexokinase in, 455-457, 456f in liver, 476 in muscle, 473-474 historical perspective on, 454 hypoxia-inducible transcription factor in, 479. 479f. 479t isomerization of three-carbon phosphorylated sugars in, 456f, 458-460, 458f in liver. 475–477. 487 in muscle, 473–474, 475f NADH in, 466-468 pentose phosphate pathway and, 601-609 phosphoenolpyruvate in, 464f, 465 phosphofructokinase in in liver, 474-477, 475f, 476f, 487, 488 in muscle, 473-474, 473f phosphoglycerate kinase in, 463-464 phosphoglycerate mutase in, 464, 464f phosphoryl group transfer in, 444, 444f pyruvate formation in, 464–466, 464f-466f pyruvate kinase in, 465 in liver, 476-477, 476f in muscle, 474 reactions of, 445, 456f, 466t-467t regulation of, 472–479, 486–491, 802-803.803f reversal of, stoichiometry of, 485 stages of, 456f in starvation, 809, 809f in starved-fed cycle, 807-810 thioester intermediate in, 461-463, 462f transaldolase in, 601-604, 605, 606f transketolase in, 601-605, 605f triose phosphate isomerase in, 456f, 458-460, 459f Glycomics, 320 Glycopeptide transpeptidase, penicillin inhibition of, 244, 245f Glycoproteins, 329-337 sequencing of, 336-337 Glycosaminoglycans, 329-330, 331

Glycosides, 326 Glycosidic bonds in disaccharides. 327 in monosaccharide-alcohol/amine complexes, 326, 326f Glycosyl phosphatidylinositol, 359, 359 Glycosylated hemoglobin, 325-326 Glycosylation, 329-337 congenital disorders of, 336 proteases in, 337 sites of, 330, 333-334, 337 Glycosyltransferases, 335, 335 in blood groups, 335 Glyoxylate cycle, 518-519, 519f Glyoxylate, in Calvin cycle, 593-594 Glyoxysomes, 519 Glyphosate, 719-720 GMP (guanosine monophosphate), synthesis of, 743-744, 743f, 744, 751, 751f GMP synthetase, 744 Goldstein, Joseph, 767b, 775, 776 Golgi complex protein glycosylation in, 333-334, 333f, 334f protein modification in, 913-914, 914f Gout, 751, 753-754 G-protein receptor kinase 2 (GRK2), 408 G-protein-coupled (7TM) receptors, 404-406 Gradient centrifugation, 77-78, 77f Graft rejection, 998 Granum, 567 Granzymes, 996 Grb-2, 417 Green fluorescent protein, 58, 58f, 90 Green photoreceptors, 969-970, 969f, 970f Greider, Carol, 837 GRK2 (G-protein receptor kinase 2), 408 Group-specific reagents, 241 Group-transfer reactions, 444. See also Phosphoryl group transfer activated carriers in, 438-442 active carriers in. 438f. 439f GTP (guanosine triphosphate) in AMP synthesis, 743f, 744, 751 hydrolysis of, 431 in translation, 902-903, 905f in olfaction, 959, 960 in protein targeting, 913 in signal transduction, 405-406, 407-408, 407f, 418, 418f in translation, 902-903, 908, 913 in tubulin, 1019 in vision, 968, 968f GTPase(s) in protein targeting, 912 in signal transduction, 407, 407f GTPase activating proteins (GAPs), 418 GTPases, in signal transduction, 298, 298t Guanidinium. 31. 31 Guanidinium chloride, in bond cleavage, 49. **49**
C23 Index

Guanine, 4, 4, 111, 111 deamination of, 838 degradation of, 753f Guanine-nucleotide exchange factor (GEF), 417 Guanosine, 111 Guanosine diphosphate. See GDP (guanosine diphosphate) Guanosine monophosphate. See GMP (guanosine monophosphate) Guanosine triphosphate. See GTP (guanosine triphosphate) Guanylate. See GMP (guanosine monophosphate) Guanylate cyclase, in vision, 969 Guanylate kinase, structure of, 284 Gustation, 962-966. See also Taste Gustducin, 963, 963f Gyrase inhibitors, 831 H chains. See Heavy (H) chains H isozyme, 297, 297f H zone, 1016, 1016f Haber, Fritz, 707 Haemophilus influenzae, genome of, 158, 158f, 181 Hair cells, 971-973, 971f, 972f Hairpin turns, 42, 43f amino acid residues in, 50-51, 51t in mRNA, 42, 43f, 127, 127f in transcription termination, 859-860.859f Haldane, John, 231 Half-life of drugs, 1036 of proteins, 675, 676t Haptenic determinant, 990 Haptens, 990 HAT (histone acetyltransferase), 949.950 Hatch, Marshall Davidson, 599 Haworth projections, 323 Hayaishi, Omar, 696 HDLs (high-density lipoproteins), 773, 773t, 774-775, 776, 778-779. See also Lipoprotein(s) Hearing, 971-973 Heart disease, homocysteine in, 719 Heart failure, digitalis for, 377 Heart, H and M isozymes in, 297, 297f Heat, from oxidative phosphorylation, 556-557 Heat shock protein 70, 183, 183-184 Heat-shock promoter, 858, 858f Heat-shock response element, 868 Heat-shock transcription factor, 868 Heavy (H) chains, 981-982, 981f, 987-989, 988f in class switching, 991 formation of, 991 Heavy meromyosin, 1009, 1009f Heimiacetals, 322 Heinz bodies, 610, 610f

Helicases, 120, 824-825, 824f, 825f in DNA replication, 120, 824-825, 825f. 832 in splicing, 876-877 structure of, 824, 824–825, 824f, 825f Helix alpha, 38-40, 39, 40, 40f amino acid residues in, 50-51, 51t of α-keratin, **43**, 43–44, 44f of kinesin, 1012 of membrane proteins, 356, 358, 359-360, 360f, 360t of myosin, 1009-1010, 1010f, 1012f of collagen, 44-45, 44f double. See Double helix recognition, 923 relay, 1012, 1012f screw sense of, 39, 40f Helix-loop-helix, in EF hand, 410-411, 411f Helix-turn-helix motif, 47, 47f, 923. 923f Helper T cells, 980, 996-998, 998f. See also T cell(s) in HIV infection, 999-1000 Hemagglutinin, 339, 340f Heme, 45, 196, 196–198 in cytochrome c oxidase, 538-539 degradation of, 730, 730f labeling of, 728, 728b, 728f in Q-cytochrome c oxidoreductase, 535-536, 536f synthesis of, 728-730, 729f Hemiacetals, 322 cyclic, 323 Hemichannels, 394, 394f Hemiketals, 322-323 Hemoglobin, 195-216 α chains of, 199, 199f, 211, 211 αβ dimers of, 199, 199f, 202, 202f β chains of, 199, 199f, 211 β-globin gene of, transcription and processing of, 131-132, 132f evolution of. 175-181. 187-188. 187f fetal, oxygen affinity of, 205, 205f glycosylated, 325-326 Hill plot for, 213-214, 214f leghemoglobin and, 180, 180f, 181f, 183, 183f. See also Leghemoglobin overview of, 195-196 oxygen affinity of, 204-205, 204f pH and, 206-207, 206f oxygen binding by, 196-199 allosteric effectors for, 206 concerted (MWC) model of, 202-203, 203f, 214–216, 216f cooperative nature of, 200-201, 201f, 203f quantitative models of, 213-216, 214f, 216f sequential model of, 203, 203f, 216 structural changes in, 201-202, 202, 204, 204f

oxygen release by, 200, 201f, 206-208 Bohr effect and, 206-208, 207f carbon dioxide in, 207-208, 208f in oxygen transport, 200–201, 207-208, 208f oxygen-binding curve for, 199–200, 200f, 201f with concerted model, 214-216, 216f for Hill coefficients, 214, 214f R state of, 202-204, 203f sequence homology with myoglobin, 175-181, 176f-182f, 199 structure of, 48, 48f, 199 oxygen-binding-induced changes in, 201-202, 202, 204f oxygen-binding-induced changes in, 204 quaternary, 201-202, 202 tertiary, 183, 183f T state of, 202–204, 203f, 207–208, 207f vs. myoglobin, 195-196 Hemoglobin A, 199 Hemoglobin gene, introns in, 131-132, 132f Hemoglobin H, 210 Hemoglobin S, 209, 209-210, 209f Hemolytic anemia, G6PD deficiency and, 609-610, 611 Hemophilia, 311 Henderson-Hasselbalch equation, 16-17 Henseleit, Kurt, 685 Heparin, 312, 331, 332 Hepatic gluconeogenesis, 481 Hepatic glycolysis, 475-477 Hepatitis, alcoholic, 811 Hepatotoxicity, of acetaminophen, 1036, 1037f Heptad repeats, in coiled-coil proteins, 43, 44f Heptoses, 320 Her2 receptor, 417, 421 Herbicides, mechanism of action of, 584, 584, 719-720 Hereditary nonpolyposis colorectal cancer, 842-843 Hers disease, 634t Heterolytic cleavage, 651 Heterotrophs, 566, 590 Heterotropic effect, 296 Hexokinase in glycogen metabolism, 620 in glycolysis, 455-457, 456f, 457f in liver, 476 in muscle, 473-474 induced fit in, 457, 457f Hexose monophosphate pathway. See Pentose phosphate pathway Hexose monophosphate pool, 594 Hexose sugars, formation of, in Calvin cycle, 594-597, 594f Hexoses, 320-321 High-density lipoproteins, 773, 773t, 774-775, 776, 778-779. See also Lipoprotein(s)

INDEX

High-energy bonds, 433-434 High-pressure liquid chromatography, 71.71f High-protein-low-carbohydrate diets, 798 High-throughput screening, 1041 Hill, Archibald, 213 Hill coefficients, 214 Hill plot, 213-214, 214 Histamine, synthesis of, 726, 726f Histidine in carbon dioxide hydration, 269, 270, 270f in catalytic triads, 258-260. See also Catalytic triads conversion to glutamate, 692, 692f distal, 198, 198 histamine from, 726, 726f in oxygen release from hemoglobin, 207.207f proximal, 197 structure of, 31, 31f Histidine operon, 932-933, 932f Histone acetyltransferase (HAT), 949, 950 Histone deacetylase, 950 Histones, 939-941, 940f acetylation of, 298t, 949-951, 950f amino-terminal tails of, coactivator modification of, 949-951, 950f in gene regulation, 939-941, 940f, 949-951,950f sequence homology of, 940-941, 940f structure of, 940-941, 940f HIV infection. See Human immunodeficiency virus infection H^+ – K^+ pump, 374 HMG-CoA (3-hydroxy-3-methylglutaryl CoA), 653, 653, 654 in cholesterol synthesis, 767–768, 770-771 metabolic fates of, 768f regulation of, 770-771 HMG-CoA reductase, in cholesterol synthesis, 767-768 HMG-CoA reductase inhibitors, 779, 779, 1040, 1040-1041, 1041 Hodgkin, Alan, 382 Hoffmann, Felix, 1039-1040 Hogness box, 126f, 127 Holliday junctions, 845-846, 845f Holliday, Robin, 845 Holoenzymes, 221 Homeostasis, caloric, 791-798. See also Caloric homeostasis Homocysteine in activated methyl cycle, 717-718, 717f in amino acid degradation, 693, 693 in amino acid synthesis, 717–718 in cysteine synthesis, 719 synthesis of, 717 in vascular disease. 719 Homocysteine methyltransferase, 717 Homocystinuria, 698t Homogenate, centrifugation of, 67, 75t Homogentisate, 695, 696

Homogentisate oxidase, 696 in alcaptonuria, 697 Homologous recombination, 970b in color blindness, 970 in gene knockout, 164–165 Homologs, 174-175, 175f Homology, 174-181, 175f sequence, 175-181, 176f-182f. See also Sequence homologies Homolytic cleavage, 651, 651f Homopolymers, 328 Homotropic effects, 294 Hood, Lerov, 987 Horizontal gene transfer, 278–279 Hormone(s). See also specific hormones in citric acid cycle regulation, 514 eicosanoid, 664-665, 665, 665f in fatty acid metabolism, 643–644, 666-667 in gene expression, 946-949 in gluconeogenesis, 488 in glucose metabolism, 488 in glycogen metabolism, 624-626, 632-633, 632f local. 665 metabolic functions of, 446-447 sex regulation of, 946 synthesis of, 780, 780f in signal transduction, 405–406. See also Signal transduction steroid. 780–782 anabolic, 785, 948 cardiotonic, 377, 377f hydroxylation of, 781-782 structure of, 781, 781 synthesis of, 780, 780f, 783 Hormone receptor(s), 946–948, 947f–949f, 948f, 949f DNA-binding domains of, 946, 947f drug binding to, 948-949, 949f ligand-binding domains of, 946-947, 947f nuclear localization of. See Nuclear hormone receptors Hormone-receptor complexes, in signal transduction, 405-406 Human immunodeficiency virus, 999-1000.999f Human immunodeficiency virus infection aspartyl protease in, 264, 264f helper T cells in, 996-998 immune system in, 999-1000, 999f protease inhibitors for, 264-265, 265, 266f development of, 1043, 1043f, 1044f resistance to. 1050 vaccine for, 1003 Humoral immune response, 979-980 Huntingtin, 842 Huntington disease, 55-56, 842 Hurler disease, 332, 332f Huxley, Andrew, 382 Hyaluronate, 331, 332 Hybridization, DNA, 121

Hybridoma cells, in monoclonal antibody production, 87-88, 87f Hydrogen bonds, 5, 8, 9, 9f, 11, 38-40. See also Bonds in α helix, 38–40, 39f in antigen-antibody binding, 984, 985, 985f. 986 in β sheet, 40–42, 40f, 41f in double helix, 5, 5f, 10, 114, 118f in enzyme-substrate complex, 228, 228f in water, 9-10, 9f Hydrogen ion(s). See also Acid(s) in aqueous solutions, 13-17 oxygen affinity of hemoglobin and, 206-207, 206f pH and, 13–14. See also pH sour taste of, 962, 962f, 965-966 Hydrogen ion channels, in taste, 965-966 Hydrogen-potassium ion pump, 374 Hydrolases, 249t. See also Enzyme(s) Hydrolysis in metabolism, 444-445 of peptide bonds. See Peptide bonds, cleavage of Hydronium ions, 13 Hydropathy plot, 360, 361f Hydrophilic side chains, 30, 30f. See also Amino acid side chains protein folding and, 46, 46f, 47 Hydrophobic effect, 9-10, 9f, 28 Hydrophobic interactions, 10 Hydrophobic side chains, 28, 29f. See also Amino acid side chains protein folding and, 45-46, 46f, 47 Hydroxide ions, pH and, 13-14 3-Hydroxy-3-methylglutaryl CoA. See HMG-CoA (3-hydroxymethylglutaryl CoA) L-3-Hydroxyacyl CoA dehydrogenase, 647 3-Hydroxyanthranilate, 696, 696 D-3-Hydroxybutyrate, 653, 653-655. See also Ketone bodies D-3-Hydroxybutyrate ACP, 658t, 659, 659 Hvdroxvethvl-TPP in amino acid synthesis, 723-724, 723f in citric acid cycle, 500, 500 Hydroxyl groups, 30, 30f, 44, 57-58 Hydroxylation cytochrome P450 in, 782-783, 782f protein, 30, 30f, 57-58 steroid, 781-782 p-Hydroxymercuribenzoate, 291, 291, 293 Hydroxyphenylpyruvate, 696 in amino acid degradation, 695, 696f in amino acid synthesis, 720, 721f Hydroxyphenylpyruvate hydroxylase, 696 17α-Hydroxyprogesterone, 784, 784 Hydroxyproline, 44, 57 synthesis of, 812f, 813 Hyperbaric oxygen therapy, 206 Hypercholesterolemia, 759, 774-775 familial, 759, 776-779

C25 Index

Hyperlysinemia, 698t Hypersensitive sites, 944 Hyperuricemia, 753-754 Hypervariable loops, 984, 984f Hypochromism, 120, 120f Hypomethylation, 945 Hypoxanthine, 838, 838 degradation of, 752, 753f Hypoxanthine-guanine phosphoribosyltransferase (HGPRT), 744 deficiency of, 754 Hypoxia inducible factor 1 (HIF-1), in cancer, 515 Hypoxia, tumor, 479 Hypoxia-inducible transcription factor (HIF-1), 479, 479f, 479t I band, 1016, 1016f Ibuprofen, 358 I-cell disease, 336 Ig. See Immunoglobulin(s) Ig-α-Ig-β, 989, 989f Imatinib mesylate, 421 Imidazole, 31, 31 4-Imidazolone 5-propionate, 692, 692 Immune response, 977-1004. See also Antibodies; Antigen(s) antigen binding in, 984-986 in autoimmune diseases, 1001 B cells in, 989-990. See also B cell in cancer, 1001-1002 cellular, 980 class switching in, 990-991, 991f evolution of, 978-981 humoral, 979-980 overview of, 977-978 selectins in, 339, 339f to self-antigens, 1001 self-tolerance in, 1001 signaling in, 989–990, 998 T cells in, 980–981. See also T cell(s) Immune-modulating drugs, 990 Immunity adaptive, 977, 978, 979-981 cell-mediated, 961 innate, 978–979 Immunization. See Vaccine(s) Immunodeficiency, gene therapy for, 167-168 Immunoelectron microscopy, 91, 91f Immunogens, 980 Immunoglobulin(s). See also Antibodies cleavage of, 982, 982f constant regions of, 984, 984f definition of, 83 heavy chains of, 981-982, 981f, 987-989, 988f in class switching, 991 formation of, 991 light chains of, 981-982, 981f, 987-989, 987f, 988f in class switching, 991

properties of, 982t segmental flexibility of, 982, 982f sequence diversity in, 983f, 984 variable regions of, 983, 984f Immunoglobulin α , 989 Immunoglobulin A (IgA), 982t, 983 Immunoglobulin β , 989 Immunoglobulin D (IgD), 982t, 983, 983f Immunoglobulin domains, 981–982 Immunoglobulin E (IgE), 982t, 983, 983f Immunoglobulin fold, 984, 984f Immunoglobulin G (IgG), 981f-983f, 982, 982t, 983f Immunoglobulin M (IgM), 982t, 983, 983f B cells and, 989, 989f Immunologic techniques antibody production for, 86-88, 87f hybridoma cells in, 87-88, 87f monoclonal antibodies in, 86-88, 86f-88f polyclonal antibodies in, 86, 86f for protein study, 84-91 Immunological memory, 1002 Immunoreceptor tyrosine-based activation motif (ITAM), 989 Immunosuppressants, 990 Immunotoxins, genetically engineered, 157 IMP. See Inosine monophosphate (IMP) Inactivated vaccines, 1002 Inborn errors of metabolism, 697-698, 698t Indian muntjac, genome of, 113, 113f Indinavir, 264-265, 265, 266f development of, 1043, 1043f Indirect ELISA, 88, 89f Indole, substrate channeling and, 722-723 Indole-3-glycerol phosphate, 721, 722 Induced pluripotent stem cells, 944 Induced-fit model, of enzyme-substrate complex, 228, 228f, 254, 457, 457f Infant respiratory distress syndrome, 765-766 Influenza virus, 339-340 Information science, 17 Inhibition constant (K_i) , 1031 Initiation factors, 902 Initiator tRNA, 907 In-line displacement, 272-274 Innate immunity, 977. See also Immune response Inosinate. See Inosine monophosphate (IMP) Inosine in anticodons, 891 degradation of, 752, 753f Inosine monophosphate (IMP), 742f, 743 AMP from, 743-744, 743f GMP from, 743–744, 743f synthesis of, 743, 743f, 744, 751 Inositol 1,4,5-trisphosphate (IP₃), 403, 408-409, 409f Inositol, in phospholipids, 349 Insertional inactivation, 1494 Insertions, production of, 157 Insig, 771, 772f

Insulin amino acid sequence of, 35, 35f in appetite regulation, 796, 797 biosynthesis of, 303 in diabetes, 798-803. See also Diabetes mellitus exercise and, 803 in fatty acid synthesis, 667 in glucose metabolism, 488, 798-800 glucose-stimulated secretion of, 801-802 in glycogen metabolism, 632–633, 632f mass spectrometry of, 91-95, 93f in obesity, 798, 800-802 regulation of, 798-800 secretion of, 799, 799f, 801-802 in signal transduction, 411–415, 412f-415f, 798-800, 799f structure of, 35, 35f, 412, 412 Insulin receptor, 412-415, 412f-415f, 799, 799f Insulin receptor substrates, 412-415, 414f, 799.800 Insulin receptor tyrosine kinase, 412-415 Insulin resistance, 798-802, 799-802 Insulin-dependent diabetes mellitus, 802-803. See also Diabetes mellitus Integral membrane proteins, 355, 355f Intercalation, 862 Intermediate-density lipoproteins, 773, 773t, 774. See also Lipoprotein(s) Internal aldimine, 682, 682 in amino acid degradation, 682, 682f in amino acid synthesis, 712, 712f Internal guide sequence, 881, 881f Internal repeats in amino acid sequences, 79 sequence homology of, 184, 185f Internet databases, of amino acid sequences, 65-66, 181, 182f Intestinal microvilli, 328f Intestines, protein digestion and absorption in, 674, 674f Intrinsic pathway, coagulation, 308, 308f Intrinsically unstructured proteins, 54-55 Introns, 110, 131-132, 132f, 869 evolution of, 133 group I, 880 removal of, 132-133, 132f. See also Splicing Inverted repeats, as recognition sites, 275 Ion channels, 372, 382–393 acetylcholine receptor as, 389-390, 390f, 391f action potentials and, 382, 382f, 391-392, 391f calcium, 367, 384f, 386 cell-to-cell, 393-394, 393f, 394f definition of, 382 equilibrium potential and, 391-392, 391f gap junction, 372, 393-394, 393f, 394f in hearing, 972-973 inactivation of, 388-389, 389f, 392 ligand-gated, 389-390, 391f

INDEX

Ion channels—(continued) nerve impulses and, 382 opening and closing of, 389-390, 391f, 392 patch-clamp studies of, 383, 383f potassium, 364-393, 382. See also Potassium ion channels rate of transport in, 387 responsive to mechanical impulses, 972-973 selectivity filter in, 385-386, 386f, 387f sequence homologies of, 384, 384f, 386 sodium, 382, 383-384, 384f, 387-392. See also Sodium ion channels specificity of, 385-387 in taste, 965–966, 965f **TRP. 973** voltage-gated, 387, 388f Ion, dipolar, 27-28, 27f Ion gradients, 373-374 in ATP synthesis, 437-438, 437f Ion pumps, 371-372, 374-381 Ca²⁺, 374–377, 375f, 376f evolution of, 374, 378-379 H⁺-K⁺, 374 Na⁺-K⁺, 374, 377 Ion-exchange chromatography, 69-70, 70f, 75,75t in amino acid identification, 80-82,80f IP₃ (inositol 1,4,5-trisphosphate), 408, 408-409, 409f IPGT (isopropylthiogalactoside), 926, **926** IRE (iron-response element), 951-952, 952f IRE-binding protein, 951–952 Iron in heme, 730. See also Heme oxygen binding to, 196-198, 197f metabolism of, 951-953 Iron center, of ribonucleotide reductase, 746 Iron protein, in nitrogen fixation, 707, 708-709 Iron-molybdenum cofactor, 708, 708-709 Iron-response element (IRE), 951–952 Iron-response element-binding protein (IRP), 951-952, 953f Iron-sulfur proteins, 506 in nitrogen fixation, 708-709 in oxidative phosphorylation, 534, 536 IRP (iron-response element-binding protein), 951-952, 953f Isocitrate dehydrogenase, in citric acid cycle, 506, 514, 514f Isocitrate, in citric acid cycle, 506-507. 507f Isocitrate lyase, in glyoxylate cycle, 518-519, 519f Isoelectric focusing, 73, 73f with SDS-PAGE, 74, 74f

Isoelectric point, 73 Isoenzymes, 289–290, 296–297, 297f Isoleucine degradation of, 694–695 in maple syrup urine disease, 697 structure of, 28, 29f synthesis of, 723-724 Isomerases, 249t. See also Enzyme(s) Isomerization, in metabolism, 443t, 444 Isomers, 27 carbohydrate, 321 constitutional, 321, 321f 3-Isopentenyl pyrophosphate, 742f, 768, **768** Isopeptide bonds, 675, 675f Isopropylthiogalactoside (IPTG), 926, 926 Isovalervl CoA, 694, 694 Isozymes, 289–290, 296–297, 297f ITAM (immunoreceptor tyrosine-based activation motif), 989

J genes, 987–988 in antibody switching, 991 Jacob, François, 924 Jagendorf, André, 577 Jenner, Edward, 1002 Joining (J) genes, 987–988 in class switching, 991 Joule (J), 7, 223b

K⁺ ions. See Potassium к chains, 982, 983f, 987, 989 k_{cat} (turnover number), 234. 234t k_{cat}/K_M ratio, 234–235 K_d (dissociation constant), 1030–1031 for enzyme-substrate complex, 233-234 for ligand binding, 1030-1031, 1030f Kendrew, John, 196 Kennedy, Eugene, 526, 644 Keratan sulfate, 331, 332 α -Keratin, α helices of, 43–44, 44, 44f Ketimine, 682, 682, 712 Ketoacidosis, in maple syrup urine disease, 697 3-Ketoacyl CoA, 647 α -Ketobutyrate, 719 in amino acid degradation, 693, 693 in amino acid synthesis, 719, 723 Ketogenic amino acids, 690. See also Amino acid(s) Ketogenic diet, 656 α -Ketoglutarate, 692 in amino acid degradation, 692 in amino acid synthesis, 709–710, 712 in citric acid cycle, 507, 507f, 692 in double-displacement reactions, 237 in glutamate synthesis, 709, 710 in 4-hydroxyproline synthesis, 812f, 813 in nitrogen fixation. 709 α-Ketoglutarate dehydrogenase complex in citric acid complex, 499-500, 507 in citric acid cycle, 507, 514–515, 514f α-Ketoisocaproate, 693-694, 694

Ketone bodies, 653-656 acetyl CoA formation from, 633f, 653-655 in citric acid cycle, 810, 810f in diabetes, 633f, 653, 655, 803 in ethanol metabolism, 811 as fuel source, 654-655, 654f in starvation, 809f, 810 synthesis of, 810, 810f Ketose, 320, 320f in Calvin cycle, 594 in transaldolase reaction, 605, 606f in transketolase reaction, 605, 605f Ketosis, diabetic, 633f, 653, 655 3-Ketosphinganine, 764 in sphingolipid synthesis, 737 β-Ketothiolase, 647, 647t Keyhole limpet hemocyanin, 990 Khorana, Har Gobind, 128 K_{i} (inhibition constant), 1031 Kidnev drug metabolism in, 1035 gluconeogenesis in, 481 Killed vaccines, 1002 Killer T cells, 980, 994–995, 997–998, 997f. See also T cell(s) Kilobase (kb), 124b Kilocalorie (kcal), 7, 223b Kilojoule (kJ), 7, 223b Kinase(s). See also specific types activity and properties of, 457 definition of, 457 induced fit in, 457, 457f magnesium and, 457 in phosphorylation, 457 substrate-induced cleft closing in, 457,457f Kinase cascade, in signal transduction, 406-407, 408-409, 409f, 418 Kinesin, 1008, 1018-1022 ATP binding to, 1012, 1012f, 1021, 1021f conventional, 1010 disorders of, 1018 movement along microtubules, 1020-1022, 1020f, 1021f myosin and, 1008 relay helix of, 1012, 1012f structure of, 1010, 1010f Kinetic energy, 11. See also Energy Kinetic perfection, 235 Kinetically perfect enzymes, 234, 235, 460 Kinetics. See also Bioenergetics; Energy; Reaction rates definition of. 229 enzyme, 229-237. See also Enzyme kinetics Kirschner, Marc, 1019 Klenow fragment, 821, 821f Klug, Aaron, 939 K_M (Michaelis constant), 231–235 calculation of, 233 definitions of, 232, 234 for selected enzymes, 233t

C27 Index

Köhler, Geörges, 87 Kornberg, Arthur, 121 Kornberg, Roger, 853 Koshland, Daniel E., Jr., 228 Krebs cycle. See Citric acid cycle Krebs, Hans, 437, 518b, 685 Kringle domain, in prothrombin, 310, 310f Kynurenine, 696, 696 L amino acids, 27, 27f L chains. See Light (L) chains Laborit, Henri, 1038 lac operator, 926, 926f, 927f lac operon, 925-928, 925f, 926f lac regulatory site, nucleotide sequence of, 922f lac repressor, 922f, 923, 925-928, 926f. 927 lac repressor-DNA complex, 922f Lactase, 328, 454 deficiency of, 471 Lactate, 236 in Cori cycle, 490, 490f formation of, 236, 489-490, 490f from pyruvate, 467-468 in gluconeogenesis, 453, 479 in glycolysis, 453 in muscle contraction, 489-490, 490f, 806 Lactate dehydrogenase, 236 evolution of, 490-491 in gluconeogenesis, 479 in glycolysis, 467-468 isozymes of, 297, 297f, 490-491 Lactic acid fermentation, 453, 467-468 Lactobacillus, 471f β-Lactoglobulin, mass spectrometry of, 93, 93f Lactose, 328 metabolism of, 923-924 Lactose intolerance, 471 Lactose operon. See lac operon Lactose permease, in membrane transport, 381, 381f Lagging strand, 823f, 824, 832-833, 833f λ chains, 982, 983f λ phage, 150–151, 150f, 151f λ repressor, 928–929, 928f, 929f Cro protein and, 929 genetic switching and, 928-931 self-regulation by, 928-929 Lanosterol, 769-770, 770 Lansoprazole, 1046, 1046 Lariat, in splicing, 874, 874f, 875f Lateral diffusion, of membrane lipids, 361-364, 362f Laws of thermodynamics, 11-13. See also Thermodynamics LDLs. See Low-density lipoprotein(s) Leader sequences, 931, 932, 932f Leading strand, 823f, 824, 832-833, 833f Leber hereditary optic neuropathy, 559 Lectins, 337-339, 338 Leder, Philip, 987

Leghemoglobin, 180, 181f, 183, 183f, 707. See also Hemoglobin evolution of, 187-188, 187f tertiary structure of, 183, 183f Lehninger, Albert, 526, 644 Leloir, Luis, 627 Leptin, 795-797, 796f Leptin resistance, 797 Lesch-Nyhan syndrome, 754 Leucine degradation of, 693-695 in maple syrup urine disease, 697 structure of, 28, 29f synthesis of, 723-724 Leucine-rich repeats, 978, 978f Leukemia, chronic myelogenous, treatment of, 421 Leukotrienes, 664-665, 665f Lever arm, of myosin, 1011-1012, 1011f, 1018, 1018f Levinthal, Cyrus, 52 Levinthal's paradox, 53 Libraries cDNA, 154 genomic, 151-153, 152f, 153f screening, in drug development, 1039-1042, 1042f Licensing factors, 835 Li-Fraumeni syndrome, 843 Ligand, 402, 1030 Ligand binding dissociation constant for, 1030-1031, 1030f in drug development, 1030-1031, 1030f in gene expression, 926, 946-947 Ligand-binding curve, 1030, 1030f Ligand-gated ion channels, 389-390, 391f. See also Ion channels Ligase, 249t, 824, 841. See also Enzyme(s) Ligation reactions, in metabolism, 443-444, 443t Light (L) chains, 981-982, 981f, 987-989, 987f, 988f in class switching, 991 Light, electromagnetic spectrum of, 966, 966f Light meromyosin, 1009, 1009f Light perception. See Vision Light reactions, 566-567, 566f, 572-584, 589. See also Photosynthesis in photosystem I, 567, 572, 572f, 574f, 575-577, 575f, 583, 583f in photosystem II, 567, 572-577, 572f-575f, 583, 583f Light-harvesting complexes, 583, 583f Limit dextrin, 454 LINES, 160 Lineweaver-Burk plot, 233, 233f, 240, 240f Linker DNA, 148-149, 149f, 939 Linking number, 826, 827f Linoleate, 664. 664 oxidation of, 648-649, 649f Linolenate, 664, 664 Linoleoyl CoA, 648-649, 649 Lipases, 641-642, 642f, 643-644

Lipid(s) dietary, solubilization of, 779-780 energy storage in, 641-642 fatty acids in, 346-348, 347f, 347t. See also Fatty acid(s) hydrolysis of, 643-644, 643f membrane. See Membrane(s); Membrane lipids metabolism of. See Fatty acid metabolism mobilization of, 643-644, 643f transport of, 642, 642f Lipid A, 978, 978f Lipid bilayer. See Membrane(s), lipid bilaver of Lipid diffusion lateral, 361-364, 362f transverse, 362 Lipid kinases, in insulin signaling, 414 Lipid phosphatases, in signal transduction, 415 Lipid rafts, 363 Lipid vesicles, 353-354, 353f Lipin 1, 766-767 Lipinski's rules, 1032-1033, 1032f Lipoamide, in citric acid cycle, 501, 501, 502-503 Lipoic acid, in citric acid cycle, 500 Lipolysis, 643-644, 644f. See also Fatty acid metabolism in starvation, 809, 809f, 810 Lipophilic molecules, 372 Lipopolysaccharide (LPS), 978-979, 978f Lipoprotein(s) classification of, 773, 773t high-density, 773, 773t, 774-775, 776, 778-779 intermediate-density, 773, 773t, 774 low-density, 559, 744-748, 773-774, 773t, 775-778 in familial hypercholesterolemia, 776-778 receptor-mediated endocytosis of, 775-776, 776f metabolism of, 775f transport function of, 773-774 very-low-density, 773, 773t Lipoprotein lipases, 773 Lipoprotein particles, 773 Liposomes, 353-354, 353f Lipoxygenase, 665 Liquid chromatography, high-pressure, 71, 71f L isomers, monosaccharide, 320 Live attenuated vaccines, 1002 Liver acetaminophen-related injury of, 1036, 1037f alcoholic injury of, 811-812 in amino acid metabolism, 680 cholesterol synthesis in, 770, 774f cirrhosis of, 811-812 drug metabolism in, 1035-1036, 1036f ethanol metabolism in, 811-812

INDEX

Liver—(continued) fatty, 811 fatty acid metabolism in, 761 gene expression in, 939t glucose metabolism in, 621, 633-634, 633f. 806 gluconeogenesis in, 490, 490f glycolysis in, 475-477, 487, 807 ketone body synthesis in, 654-655, 654f, 810, 810f, 810t metabolism in interchanges with muscle in, 490, 490f, 684-685 in starvation, 810, 810t triacylglycerol synthesis in, 761, 773, 774f Local hormones, 665 Lock-and-key model, of enzyme-substrate complex, 228, 228f Long interspersed elements (LINES), 160 Long QT syndrome, 392–393 Loops, 42, 43f Lou Gehrig's disease, 147-148 Lovastatin, 779, 779, 1040, 1040-1041 Low-carbohydrate-high-protein diets, 798 Low-density lipoprotein(s), 773-774, 773t, 774f-777f, 775-778, 779. See also Lipoprotein(s) in familial hypercholesterolemia, 776-778 receptor-mediated endocytosis of, 775-776.776f Low-density lipoprotein receptor, 777–778, 777f Low-phenylalanine diet, 674 Lungs, oxygen transport to, 200–201, 208, 208f LuxR, 930 Lyase-catalyzed reactions, 445 Lyases, 662b. See also Enzyme(s) Lymphocytes. See B cell; T cell(s) Lymphotactin, 55, 55 Lynch syndrome, 842-843 Lysine structure of. 30-31. 31f synthesis of, 724, 724f Lysophosphatidate, 760, 760 Lysosomal proteins, 911 Lysosomes definition of, 336 function of. 336 in I-cell disease, 336 in receptor-mediated endocytosis, 776 Lysozyme, 227 antibody binding of, 986, 986f molecular models of, 61-62, 61f, 62f M isozyme, 297, 297f MacKinnon, Roderick, 384, 388 Macromolecules, biological, 2 Mad cow disease, 56-57, 56f MADLI-TOF mass spectrometry, in

oligosaccharide sequencing, 337

Magnesium ions in ATP hydrolysis, 280 in Calvin cycle, 592-593, 598, 598f in DNA cleavage, 274, 277 kinases and, 457 Magnetic resonance imaging, functional, 187f, 197–198 Main olfactory epithelium, 958 Major facilitator superfamily, 477-478 Major histocompatibility complex (MHC), 961-1000, 980-981 Major histocompatibility complex proteins, 961-1000 class I. 961-996, 993f, 998-999 class II, 996-999, 996f diversity of. 998-999 peptide presentation by, 992–994, 992f. 993f structure of, 992–994. 994f T-cell receptors and, 994–998, 994f. See also T-cell receptors in transplant rejection, 998 Malaria G6PD deficiency and, 611 infectious mechanisms in, 340 sickle-cell trait and, 210, 210f Malate, 510 in Calvin cycle, 599, 600f in citric acid cycle, 510 Malate dehydrogenase, 510 Malate synthase, in glyoxylate cycle, 519.519f Malate-aspartate shuttle, 552, 552f MALDI (matrix-assisted laser desorption/ ionization), 91-92, 92f, 93f MALDI-TOF mass spectrometry, 92, 92f. 93f 4-Maleylacetoacetate, 696, 696 Malonyl ACP, 658-659, 658t, 659 Malonyl CoA, 657, 666 Maltase, 328, 454 Maltose, 327, 328 Manganese in ATP hydrolysis. 280 in photosystem II, 574, 574f Manganese center, 574, 574f Mannose 6-phosphate, 336, 336 Mannose 6-phosphate receptor, 336, 338 Mannose, structure of, 321-322, 321f, 322f Maple syrup urine disease, 697 Maps active site, 241-242 electron-density, 99-100, 100f Margoliash, Emanuel, 543 Mass spectrometry, 91-95, 92f, 93f, 95f in oligosaccharide sequencing, 336-337, 337f tandem, 93–94, 94f vs. Edman degradation. 93 Mast cells, heparin in, 312, 312f Matrix-assisted laser desorption/ionization (MALDI), 91-92, 92f, 93f McArdle disease, 634t, 635

MDR (multidrug-resistance) protein, 378, 378f Mechanism-based (suicide) inhibition, 242, 750.754 Megabase (Mb), 937b Melanin, synthesis of, 726 Melanocyte-stimulating hormone, 796 Melting, of double helix, 120-121, 120f, 824-825, 825f Membrane(s), 345-368 in Archaea, 350-351, 350f asymmetry of, 362, 363-364, 364f in bacteria, 363, 364–365 chloroplast, 567-568, 567f cholesterol in, 363 diffusion across. See Membrane diffusion in eukaryotes, 363, 364-366, 364f, 365f fatty acids in, 362-363, 363f flip-flop in, 362, 362f fluidity of, 361-364, 362f key features of, 346 lipid bilayer of, 352-355 formation of, 352-353 functions of, 362 noncovalent forces in, 353 as permeability barrier, 362 as solvent for membrane proteins, 362 melting temperature of, 362-363, 362f, 362t mitochondrial, 526, 527f overview of, 345-346 permeability of, 354, 354f, 371 liposomic evaluation of, 353-354, 353f phospholipid, 352-355, 352f-354f planar bilayer, 354, 354f synthetic, 354, 354f thylakoid, 567-568, 567f, 574-575 proton gradient across, 577-581 stacked vs. unstacked regions of, 583.583f vesicle formation from, 365 vesicle fusion to, 366, 366f Membrane channels ion. 371-372. 372f. 382-393. See also Ion channels water. 394 Membrane diffusion active, 371, 373 facilitated, 371, 373 lipid lateral, 361-364, 362f transverse, 362 simple, 372 Membrane lipids. See also Membrane(s), lipid bilayer of amphipathicity of, 351, 352 archaeal, 350-351, 350f asymmetric distribution of, 362, 363-364 cholesterol, 350, 350 fatty acids in, 346-348, 347f, 347t flip-flop of, 362, 362f glycolipids, 349-350 hydrophilicity of, 351, 352

Index

hydrophobicity of, 351, 352-353 lateral diffusion of, 361-364, 362f phospholipids, 348, 348-349, 352-355 synthesis of, 760-767, 760f polar head group of, 351 sphingolipids, synthesis of, 763-765 synthesis of, 759-787 in thylakoid membranes, 567-568 transverse diffusion of, 362 triacylglycerols, synthesis of, 760-767, 760f types of, 348-350 Membrane polarization, action potential and, 382 Membrane potential, 373, 382 action, 382, 382f, 391-392, 391f generation of, 392, 393f current generated by, 392 Nernst equation for, 391 Membrane proteins α helix of, 356, 358, 359–360, 360f, 360t, 362f amounts of, 355 bacteriorhodopsin, 356, 356f β strands of, 357, 357f channel. See Membrane channels functions of, 355 hydropathy plots for, 360, 362f hydrophobicity of, 45-46, 46f, 47f integral. 355, 355f interaction with membrane. 357-359 membrane anchors in. 359f peripheral, 355-356, 355f prostaglandin H₂, 356, 358f pump. See Ion pumps SDS-polyacrylamide gel electrophoresis, 355, 355f solubility of, 355 structure of, 355-356, 358f transporter. See Membrane transport; Transporters Membrane transport ABC transporters in, 378-380, 378f, 379f across endoplasmic reticulum, 911-914 active, 371 ATP hydrolysis in, 374-381 concentration gradient in, 373-374 free energy in, 373-374, 373f gap junctions in, 393-394, 393f, 394f ion channels in, 372, 382-393. See also Ion channels ion pumps in, 371, 374-381. See also Ion pumps membrane potentials in, 373, 382, 382f, 392. See also Membrane potential passive, 371-373 rate of, 382, 387 signaling in, 911-913 transport vesicles in, 913-914, 914f transporters in, 371-372, 380-381. See also Transporters water channels in, 394

Membrane-spanning proteins, 911 Memory B and T cells, 1002 Menten, Maud, 230 β-Mercaptoethanol, in ribonuclease denaturation, 49, 49, 49-50, 50f 6-Mercaptopurine, 1047-1048, 1048 Mercury poisoning, 517 Meromysin, 1009, 1009f Merrifield, R. Bruce, 98 Meselson, Matthew, 119 Messenger RNA. See mRNA (messenger RNA) Metabolons, 512 Metabolic pathways, 428–430, 428f. See also specific pathways amphibolic, 429 branched, regulation of, 723 committed step in, 474, 723 coordinated action of, 662 in fatty acid synthesis, 654f, 662, 666f in purine synthesis, 744 degradative vs. synthetic, 429 evolution of, 447 salvage, 740 definition of, 740 in nucleotide synthesis, 736, 736f, 744-745 Metabolic syndrome, 800 Metabolism, 427-447 activated carriers in, 438-442 allosteric interactions in, 291, 446 anabolic reactions in, 428-429 bioenergetics of, 429-435 of carbohydrates, 454 catabolic reactions in, 428, 435-438 electron carriers in, 438-439 common motifs in, 438-447 definition of, 427 drug, 1034-1036 electron carriers in, 438-439, 440t energy charge in, 446-447, 446f evolution of, 447 first-pass, 1035 free energy in, 429-430 functional group addition in, 445 group-transfer reactions in, 436-437, 443t, 444 hydrolysis in, 444-445 interconnected reactions in, 428-430, 428f isomerization in, 443t, 444 ligation reactions in, 443-444 lyase-catalyzed reactions in, 445 overview of, 427-428 oxidation-reduction reactions in, 443. 443t regulation of, 445-447 via catalytic activity, 446-447 via enzyme amounts, 446 via substrate availability, 447 substrate cycles in, 489, 489f thermodynamics of, 429-430. See also Energy; Thermodynamics

Metabolites, 2 Metal ion catalysis, 221, 221t, 253-254, 254 - 255in ATP hydrolysis, 280 in carbon dioxide hydrolysis, 221, 221t, 267-269, 267f-269f in DNA cleavage, 274, 277 in DNA replication, 821-822, 821f Metalloproteases, in peptide bond cleavage, 263, 264, 264f Metals, coenzyme, 221, 221t Metamorphic proteins, 55, 55f Metarhodopsin II (R*), 968, 968f Methionine, 693 in activated methyl cycle, 717-718, 717f conversion to succinvl CoA, 693, 693f in emphysema, 307 structure of, 28, 29f synthesis of, 650, 716-718, 724 Methionine sulfoxide, 307, 307f Methionine synthase, 717 Methionyl-tRNA, 901-902, 902f Methotrexate, 750 Methyl cycle, 717-718, 717f Methylase, in DNA cleavage, 277-278 Methylation in amino acid synthesis, 716–718, 717f, 718f of DNA, 718, 718f, 945-946 in amino acid synthesis, 716-718, 718f in cleavage, 277-278, 278f in gene regulation, 945-946, 951 hypomethylation and, 945 of tRNA, 889-890 3-Methylbutane-1-thiol, 958, 958 Methylcobalamin, 717 Methylcrotonyl CoA, 694, 694 5-Methylcytidine (mC), 890 5-Methylcytosine, 945, 945-946 N^5, N^{10} -Methylenetetrahydrofolate, 715, 716 Methylglutaconyl CoA, 694, 694 Methylmalonyl CoA, 649, 650, 693 Methylmalonyl CoA mutase, 650–652, 652,693 N^5, N^{10} - Methyltetra
hydrofolate, 749f, 750 N^5 -Methyltetrahydrofolate, 715, **716**, 717 in thymidylate synthesis, 748 Metmyoglobin, 198 Metoprolol, 1047, 1047 Mevalonate, in cholesterol synthesis, 767-768 Meyerhof, Otto, 454 MHC. See Major histocompatibility complex (MHC) Mice, transgenic, 164, 164f Micelle, 352, 352f Michaelis constant ($K_{\rm M}$), 231–235 calculation of, 233 definitions of, 232, 234 for selected enzymes, 233t Michaelis, Leonor, 230 Michaelis-Menten equation, 229-237, 231f

INDEX

Michaelis-Menten kinetics, 229-237, 231f Microarrays, DNA, 162-163, 162f Microbodies, 593 fatty acid metabolism in, 653f Microinjection, of recombinant DNA, 163, 163f MicroRNA (miRNA), 124, 872, 872f, 953-954 Microscopy fluorescence, 90-91, 90f immunoelectron, 91, 91f Microsomal ethanol-oxidizing system (MEOS), 811 Microtubules, 1018-1020 in cilia, 1019 dynamic instability of, 1019 in flagella, 1019 functions of, 1019 kinesin movement along, 1020-1022, 1020f, 1021f structure of, 1018-1019, 1019f Microvilli intestinal, 328f in taste buds, 963, 963f Milstein, César, 87 Mineralocorticoids. See also Steroid hormones synthesis of, 780, 780f Minor groove, 822, 822f Minoxidil sulfate, 1034, 1034 miRNA (microRNA), 124, 842, 872f, 953-954 Mismatch repair, 840, 840f Mitchell, Peter, 544-545 Mitchison, Tim, 1019 Mitochondria in apoptosis, 559 ATP synthase in, 526, 544-550, 558 brown fat, 556-557 citric acid cycle in, 498, 498f endosymbiotic origin of, 527-528 fatty acid metabolism in, 652 genome of, 527-528, 527f, 528f oxidative phosphorylation in, 525-528 properties of, 526-527 structure of, 526, 527f Mitochondrial ATPase. See ATP synthase Mitochondrial diseases, 558-559 Mitochondrial DNA, 527-528 genetic code of, 131, 131t sequencing of, 158 Mitochondrial outer membrane permeabilization (MOMP), 559 Mitochondrial porins, 526-527 Mitochondrial transporters, in oxidative phosphorylation, 551-553, 551f-553f Mixed-function oxygenases, 695, 781 Mobile genetic elements, 159-160 Molecular models, 60-62, 61f, 62f ball-and-stick, 22, 23f, 61, 61f space-filling, 21, 23f Molecular motors, 1007–1026. See also Motor proteins

homologous, 174-181, 175f three-dimensional structure of, 21-22 Moloney murine leukemia virus vectors, 163 Molybdenum-iron protein, 708, 708-709 Monoacylglycerol, 642 Monoamine oxidase, suicide inhibition of, 242, 242f Monoclonal antibodies for cancer. 421 preparation of, 86-88, 87f Monod, Jacques, 202, 924 Monooxygenases, 695, 781 Monosaccharides. 320-327. See also Carbohvdrates abbreviations for, 330b with alcohols. 326 aldoses, 320, 321 with amines, 326 boat form of, 324, 325f chair form of, 324, 325f definition of, 320 D-isomers of, 320, 322f envelope form of, 325, 325f envelope forms of, 325f furanose, 323-325, 323f isomers of, 321 ketoses, 320, 322 linkage of, 327-329. See also Disaccharides modified, 326, 326f nomenclature for, 323 nonreducing sugars, 325 phosphorylated, 327 pyranose, 322-325, 323f reducing sugars, 325 ring forms of, 322-325, 323f, 325f stereoisomers of, 321, 321f structure of, **320,** 320–321 Montagnier, Luc, 999 Morphine glucuronidate, 1034, 1034 Morphine, oral bioavailability of, 1032-1033, 1032f MotA-MotB pairs, 1023, 1023f, 1024f Motifs. 47. 47f repeating, 79 sequence homology of, 184, 185f Motor proteins, 1007-1026 ATP binding to, 1010–1012, 1011f, 1012f dynein, 1008, 1010, 1011f kinesin, 1008, 1010-1012, 1018-1022, 1020-1022 myosin, 1007-1018. See also Myosin overview of, 1007-1008 P-loop NTPase superfamily, 1008–1012 structure of, 1009-1012, 1009f-1012f Mouse, transgenic, 164 mRNA (messenger RNA), 109, 123-124, 123t. See also RNA in attenuation. 931–932 complementarity with DNA, 126, 126f 5' cap of, 870-871, 871f functions of, 123-124

Molecules

hairpin turns in, 42, 43f, 127, 127f in transcription termination, 859-860.859f iron-response elements in, 952-953 polycistronic, 901 precursor of, processing of, 870-872 in protein synthesis, 123-124, 130, 130f reductase, in cholesterol regulation, 771 splicing of, 869 structure of, in eukaryotes, 908 3' poly(A) tail of, 871-872, 871f transferrin-receptor, 951, 952f in translation, 123-124, 130, 130f. See also Translation translocation of, 904-906 translation of, 130, 130f MS ring, 1023, 1023f, 1024f μ chains, 938f, 982 Mucins, 330, 333, 333f Mucopolysaccharidoses, 332 Mucoproteins, 330, 333, 333f Muller, Hermann, 839 Mullis, Kary, 145 Multidrug resistance, 378 Multidrug-resistance (MDR) protein, 378, 378f Multienzyme complexes, 660-661 substrate channeling by, 722-723, 722f Multiple myeloma, 87-88 Muscle amino acid degradation in, 684 cardiac, H and M isozymes in, 297, 297f drug-related injury of, 1040 energy storage in, 641, 805t, 806 glycolysis in, 473-474, 475f, 489-490 metabolism in, 804-806, 804f interchanges with liver, 490, 490f, 684-685 in starvation, 809-810 triacylglycerols in, 641 Muscle contraction. See also Exercise actin in, 1012-1018. See also Actin/actin filaments ATP formation during, 434-435, 435f. 489-490, 805-806, 805t ATP hydrolysis in, 1009–1012, 1011f, 1012f, 1015, 1016f creatine phosphate and, 434-435, 435f gluconeogenesis in, 489-490, 490f lactate formation during, 435f myosin in, 1012-1018. See also Myosin pyruvate in, 489-490, 490f sliding filament model of, 1016-1017, 1017f Mushroom poisoning, α -amanitin in, 865, 865-866 Mutagenesis in recombinant DNA technology, 156-157, 156f, 157f site-directed, in catalysis investigation, 262-263 Mutagens, 838 Mutases, 464

C31 Index

Mutations base-pair substitution, tests for, 843-844 deletion, production of, 156 disease-causing, 157. See also Diseases and disorders functional effects of, 157 identification of, 147-148 frameshift, tests for, 843-844 insertion, production of, 157 point, production of, 157 production of, 156-157, 156f, 157f in recombinant DNA technology, 156-157 somatic, antibody diversity and, 989 substitution, production of, 156, 156f MWC (concerted) model of allosteric enzyme kinetics, 296, 296f of hemoglobin oxygen binding, 202-203, 203f. 214-216. 216f myc gene, in cancer, 869 Mycobacterium tuberculosis, 679 Myelin, protein content of, 355 Myeloma, 87-88 Myofibrils, 1016 sarcomere of, 1016-1017, 1017f Myogenin, 165, 165f Myoglobin electron-density map of, 100f evolution of, 187-188, 187f forms of, 196 Hill plot for, 213-214, 214f overview of, 195-196 oxygen binding by, 196-198, 198f-200f, 200 oxygen-binding curve for, 200-201, 200f, 201f sequence homology with hemoglobin, 175-181, 176f-182f, 199 structure of, 46, 46–48, 46f, 196, 196–197 tertiary, 183, 183f vs. hemoglobin, 195-196 Myopathy, drug-related, 1040 Myosin, 279-284, 1007-1018 ATP binding to, 1010-1012, 1011f, 1012f ATP hydrolysis and. 279-283. 281f-283f ATPase domains of, 279-280, 280f functions of, 279 kinesin and, 1008. See also Kinesin lever arm of, 1011-1012, 1011f, 1018, 1018f movement along actin, 1012-1018. See also Actin/actin filaments direct observation of, 1014-1015 power stroke in, 1015 rate of, 1018, 1018f P-loop of, 283-284 relay helix of, 1012, 1012f S1, 1009, 1009f, 1011f S2, 1009, 1009f structure of, 279, 279f, 1008-1010, 1009f, 1010f, 1011f conformational changes in, 280-283, 282f

in thick filaments, 1016, 1017f in thin filaments, 1016, 1017f Myosin-actin complex, 1014, 1014f Myristovlation, 298t Na⁺. See under Sodium NAD⁺ (nicotinamide adenine dinucleotide) in catabolism, 438, 438f in citric acid cycle, 501-502, 512 dehydrogenase binding sites for, 469, 469f in fatty acid metabolism, 646, 647, 647f in glycolysis, 469 NADH (nicotinamide adenine dinucleotide reduced) ADP units in, 447, 447f in ATP synthesis, 439, 589 in citric acid cycle, 498, 498f. 502. 503. 512. 514f ethanol and, 811 in fatty acid metabolism, 646-647, 647f, 664 in gluconeogenesis, 483-484 in glycolysis, 466-468 in oxidative phosphorylation, 525 electron-transfer potential of, 530-531 reduction potential of, 528-530 transport of, 551-552, 551f, 552f vs. NADPH, 589-590 NADH dehydrogenase, in respiratory chain, 531-535, 532f NADH-cytochrome b₅ reductase, 664, 664f NADH-Q oxidoreductase (Complex I), in respiratory chain, 531-535, 532f NADP⁺ (nicotinamide adenine dinucleotide phosphate), 438, 439 as electron carrier, 439 in pentose phosphate pathway, 601, 602f, 606-607 in photosynthesis, 576-577 in reductive biosynthesis, 439 NADPH (nicotinamide adenine dinucleotide phosphate reduced), 438, 439 in Calvin cycle, 589-590, 590f, 596f, 599 as electron carrier, 439 pathways requiring, 601t in pentose phosphate pathway, 589-590, 601-603, 604t, 607-609 in photosynthesis, 566-567, 576-577, 579-581, 589-590, 599 in reductive biosynthesis, 439, 589-590 in steroid hydroxylation, 781-782 Nalidixic acid, 831, 831 Nasal anatomy, in olfaction, 958, 959f. See also Olfaction Nathans, Daniel, 141 Neanderthals, DNA sequencing for, 188-189, 189f Neck linker, 1012, 1012f Neher, Erwin, 383 Nernst equation, 391 Nerve gases, 241 Nerve impulse, propagation of, 382, 382f

Neuberg, Carl, 454 Neural-tube defects, 755 Neuraminidase, 339, 340f Neuroglobin, 212 Neurological diseases. See Diseases and disorders Neuropeptide Y, 796 Neurotransmitters definition of, 389 ion channels and, 389-390 release of, 366f Neutral fat. See Triacylglycerol(s) Niacin, 441, 441t Nicolson, Garth, 362 Nicotinamide adenine dinucleotide. See NAD Nicotinamide adenine dinucleotide phosphate. See NADP⁺; NADPH Nicotinamide, synthesis of, 726f, 727 Nicotinic acid, 441t 9 + 2 array, 1019 19S regulatory unit, 667f, 677 Ninhydrin, in Edman degradation, 81, 81 Nirenberg, Marshall, 128 Nitric oxide in signaling, 728 synthesis of, 727-728, 728f Nitric oxide synthase, 728 Nitrogen excretion ammoniotelic organisms in, 690 glucose-alanine cycle in, 684-685, 685f transamination in, 680-685 transport to liver in, 684-685 urea cycle in, 673, 685-690 ureotelic organisms in, 685, 690 Nitrogen fixation, 706-710, 707f Nitrogen, in ammonia, 706. See also Ammonia Nitrogenase, 707 iron-molybdenum cofactor of, in nitrogen fixation, 708-709 Nitrogenase complex, 707 Nitrogen-starvation promoter, 858, 858f N-linked oligosaccharides, 330, 330f, 336 NMP kinases. See Nucleoside monophosphate (NMP) kinases Nociceptors, 973 NOESY (nuclear Overhauser enhancement spectroscopy), 101-104, 102f, 103f NompC, 972 Noncompetitive inhibition, 238, 238f, 239-240, 239f, 240f Noncovalent bonds, 7-8. See also Bonds electrostatic interactions, 7-8 in enzyme-substrate complex, 228, 228f hydrogen bonds, 8, 9, 9f. See also Hydrogen bonds hydrophobic effect and, 9-10, 9f surface complementarity and, 11 van der Waals interactions, 8, 8f, 10, 10f, 11,46 Nonessential amino acids, 711, 711t, 712f synthesis of, 712-713

INDEX

Nonheme iron proteins. See Iron-sulfur proteins Nonhomologous end joining, 841 Nonpolar molecules, 9 Nonreducing sugars, 325 Nonshivering thermogenesis, 556-557 Northern blotting, 142 Nose. See also Olfaction anatomy of, 958, 959f electronic, 961-962, 962f Novobiocin, 831 N-terminal rule, 676 NTP. See Nucleoside triphosphates (NTPs) NTPase domains, P-loop, 284, 284f NTPases, P-loop. See P-loop NTPases Nuclear envelope, 365 Nuclear hormone receptors, 946–948, 947f-949f, 948f, 949f, 950 DNA-binding domains of, 946, 947f drug binding to, 948-949, 949f ligand-binding domains of, 946-947, 947f Nuclear localization signal, 79 Nuclear magnetic resonance (NMR) spectroscopy, 101-104, 101f-103f Nuclear Overhauser enhancement spectroscopy (NOESY), 101-104, 102f, 103f Nuclear pores, 365 Nucleation-condensation model, 53 Nucleic acids, 109. See also DNA; RNA abbreviations for, 112, 112f evolution of, molecular studies of, 188-191, 190f sequence of, notation for, 112, 112f Nucleoside(s), 111, 111 nomenclature of, 736t Nucleoside diphosphate (NDP), 739 phosphorylation of, 431 Nucleoside diphosphate kinases, 431, 739 Nucleoside diphosphokinase, 629 in citric acid cycle, 508 Nucleoside monophosphate, 739 phosphorylation of, 431 Nucleoside monophosphate kinases, 248-249, 431, 739 P-loop of. See P-loop(s) structure of, 284 Nucleoside phosphorylases, 752 Nucleoside triphosphate-magnesium complex, 280 Nucleoside triphosphates (NTPs), 739. See also P-loop NTPases hydrolysis of, energy from, 431 Nucleosome core particle, 939 Nucleosomes, 939-941, 940f Nucleotidases, 752 Nucleotide(s), 110, 110, 111-112. See also Bases/base pairs: Deoxyribonucleotide synthesis; Purine nucleotides; Pyrimidine nucleotides definition of, 735 disorders of, 752-755

nomenclature of, 736t synthesis of, 735–755 de novo pathways in, 736-740, 736f deoxyribonucleotide, 745-750, 752, 752f disorders of, 753-755 inosinate in, 743-744, 743f purine, 740-745, 751, 751f pyrimidine, 736-740, 736f, 751 regulation of, 750-752 salvage pathways in, 736, 736f, 744-745 substrate channeling in, 738 transformation to amino acids. See Translation Nucleotide-excision repair, 840-841, 841f Nucleotides, nomenclature for, 112 NusA, in transcription termination, 861 Nutrition, 20, 20f. See also Diet; Food; Starvation starved-fed cycle and, 807-808 Obesity, 792-803 causes of, 792-794 definition of, 792 diabetes and, 798-803 dieting and, 797–798 evolution and, 792-793 fatty acids in muscle and, 800-801 health consequences of. 793t insulin resistance and, 798–800, 800 metabolic syndrome and, 800 pancreatic function in. 801–802 prevalence of, 792-794 Obligate anaerobes, 468–469 pathogenic, 468-469, 469t Odorant receptors, 958-960, 960f, 961f Okazaki fragments, 823-824, 823f, 833 Okazaki, Reiji, 823 Oleate, 347, 347, 664, 664 Olfaction, 958-962, 964, 964f anatomic structures in, 958, 959f combinatorial mechanisms in, 960–962 by electronic nose, 961–962, 962f evolution of, 959-960, 959f impaired, 958 odorant receptors in, 958-960, 960f, 961f 7TM receptors in, 959 signal transduction in, 959-960, 960f Oligomerization, in B-cell activation, 989-990 Oligonucleotide-directed mutagenesis, 156, 156f Oligopeptides, 34. See also Peptide(s) Oligosaccharides, 327. See also Disaccharides; Polysaccharides abbreviations for, 330b in gangliosides, 764, 765f in glycoproteins, 329-337 N-linked, 330, 330f, 336 O-linked, 330, 330f, 336 sequencing of, 336-337 O-linked oligosaccharides, 330, 330f, 336 Ω loops, 42, 43f

functions of, 735

Omeprazole, 1046, 1046 Oncogenes, 420 One-carbon units in amino acid degradation, 716 in amino acid synthesis, 715–716, 715t, 716f Operator DNA, 922f Operon(s), 924-928 histidine, 932-933, 932f lac, 925-928, 925f, 926f, 927. See also under lac phenylalanine, 932-933, 932f structure of, 924-925, 925f threonine, 932-933, 932f trp, 931 Opsin, 967 Optical trap, 1015, 1015f Oral bioavailability, 1032-1033, 1032f oriC locus, 834, 834f Origin of replication in bacteria, 834, 834f in eukaryotes, 835-836 Origin of replication complexes, 835 Ornithine, 686, 686 Ornithine transcarbamovlase, 686 deficiency of, 689 Orotate, 738, 738-739 Orotidylate, 739, 739 Orotidylate decarboxylase, 739 Orthologs, 174-175, 175f Orthophosphate in gluconeogenesis, 483 resonance structures of, 433, 433f Osteoarthritis, 332 Osteogenesis imperfecta, 45 Osteomalacia, 786 Oster, George, 548 Overlap peptides, 82, 83f Overweight, 792, 793f Oxaloacetate, 237, 443 in amino acid degradation, 692 in amino acid synthesis, 712 in Calvin cycle, 599, 600f in citric acid cycle, 498, 498f, 504, 505f, 509-510, 515, 516f, 653 in gluconeogenesis, 482, 483-484, 484f in glyoxylate cycle, 518-519, 519f synthesis of, 443-444 transfer from mitochondria to cytoplasm, 662-663 in urea cycle, 687, 687f Oxidation, of xenobiotic compounds, 1035 Oxidation-reduction potential, 528-530, 529t measurement of, 528f, 529-530 in oxidative phosphorylation, 528-531 Oxidation-reduction reactions free energy change in, 530 in metabolism, 443, 443t reduction potential of, 529-530, 529t Oxidative damage defenses against, 540-542 in DNA, 838

C33 Index

Oxidative phosphorylation, 437, 525-561. See also ATP, formation of ATP in. 525-526 ATP vield in, 554-555, 555t ATP-ADP translocase in, 552-553, 553f, 554f cellular respiration in, 526 chemiosmotic model of, 544-545, 544f, 545f in citric acid cycle, 498 electron transfer in, 528-531. See also Respiratory chain rate of, 542-543, 542f during exercise, 806 glycerol 3-phosphate shuttle in, 551, 551f heat from, 556-557 inhibition of, 558, 558f malate-aspartate shuttle in, 552, 552f NADH in, 525 electron-transfer potential of, 530-531 reduction potential of, 528-530 transport of, 551-552, 551f, 552f overview of, 525-526, 526f, 551f oxidative damage and, 540-542, 541t, 542f proton gradients in, 530-531, 543-550, 559-560, 560f proton pumps in, 525 proton-motive force in, 525, 544-545, 544f, 545f rate of, 555-556, 556f reduction potential in, 528-531 respiratory chain in, 525, 531-543. See also Respiratory Chain in thermogenesis, 556-557 transporters in, 551-553, 551f-553f uncoupling of, 556-557, 558 vs. photosynthesis, 578, 579f Oxidoreductases, 249t Oxidosqualene cyclase, 769 8-Oxoguanine, mutagenicity of, 838, 838f Oxvanion hole, 259, 259f of chymotrypsin, 261, 261f Oxygen fractional saturation of, 199 hemoglobin affinity for, 204-205, 204f hemoglobin binding of, 196-199. See also Hemoglobin, oxygen binding by hemoglobin release of, 200, 201f, 206-208, 208f carbon dioxide in, 207-208, 208f myoglobin binding of, 196-198, 198f-200f, 200 partial pressure of, 200 photosynthetic production of, 572-574, 574f reactive. See Reactive oxygen species (ROS) in steroid hydroxylation, 781-782 Oxygen therapy, hyperbaric, 206 Oxygen transport, 200-201, 207-208, 208f

in amino acid degradation, 695-696 mixed-function, 695, 781 Oxygenation, in amino acid degradation, 695-696 Oxygen-binding curve, 199-200, 200f, 201f with concerted model, 214-216, 216f for Hill coefficients, 214, 214f Oxymyoglobin, 196, 198. See also Myoglobin P clusters, 708, 708f P site, ribosomal, 900, 900f, 903 p53,843 p160 family, 948 P680, 573-574, 573f P700, 576 P960. 571 Pain management, capsaicin in, 974 Pain sensation, 973-974 PALA (*N*-phosphonacetyl-L-aspartate), 293, 293-294 Palade, George, 526 Palindrome, 141b Palindromic sequences, 141 Palmitate, 347, 347 oxidation of, 647-648, 648f synthesis of, 661-662 Palmitoleate, 664, 664 oxidation of, 648, 648f Palmitoleovl CoA, 648, 648 Palmitovl CoA, 647-648, 666 in sphingolipid synthesis, 764, 764 Palmitoylcysteine, 359, 359 Pamaquine, hemolytic anemia and, 610 PAMP (pathogen-associated molecular pattern), 978-979, 979f Pancreas in diabetes, 801-802. See also Diabetes mellitus disorders of. See Diabetes mellitus gene expression in, 939t Pancreatic β cells, 802 Pancreatic enzymes, 255-260, 303-307, 303t, 304f, 674 Pancreatic lipases, 641-642, 642f, 643-644 Pancreatic trypsin inhibitor, 306 Pancreatic zymogens, 303-307, 303t, 304f Panthothenic acid (coenzyme A), **440.** 441t as acyl group carrier, 440 ADP units in, 447, 447f in fatty acid metabolism, 644-645 Papain, 220-221 Paralogs, 174, 175f Paraquat, 584 Pardee, Arthur, 291 Park, James, 1038 Parkinson disease, 55-56, 242, 676 Parnas, Jacob, 454 Partition coefficient, in drug absorption, 1032

Oxygenases

Passive transport, 371, 373. See also Membrane transport Pasteur, Louis, 454, 1002 Patch-clamp technique, 372f, 383, 383f Pathogen-associated molecular pattern (PAMP), 978–979, 979f Pathological conditions. See Diseases and disorders Pauling, Linus, 38, 197, 209, 226b, 243 pBR322 plasmid, 149, 149f PCR. See Polymerase chain reaction (PCR) Pectin, 329, 329f Penicillin discovery and development of, 1037-1038 mechanism of action of, 244-245, 244f, 245f, 1038 resistance to, 1050 structure of, 1037, 1037 Pentose phosphate pathway, 589-590, 601-611 Calvin cycle and, 609 functions of, 608t glycolysis and, 601-609 modes of, 607-609, 607f nonoxidative phase of, 601-606, 603f, 604t oxidative phase of, 601-603, 603f, 604t rate of, 607-609 reactions of, 602f, 604t regulation of, 607-609 in selected tissues, 608t stoichiometry of, 607-609, 607f transaldolase in, 601-604, 605, 606f transketolase in, 601-605, 605f Pentoses, 320 Pepsin, 674 Peptide(s). See also Protein(s) definition of, 34 synthetic, 95-98, 96f, 97f Peptide bonds, 33, 33f, 36-37, 36f, 37f. See also Bonds angles of rotation for, 37, 37f, 38f cis configuration of, 36-37, 36f, 37f cleavage of, 58, 220-221, 255, 258-260, 259f acyl-enzyme intermediate in, 257, 257f, 259f chromogenic substrate for, 256-257, 257f deacylation in, 259-260, 259f, 260f proteases in, 255-266 aspartyl, 263, 264, 264f cysteine, 263, 263-264, 264f metal ion, 263, 264, 264f serine, 255-266 tetrahedral intermediate in, 258-259, 259f double-bond character of, 36 formation of, 903-904, 904f hydrolysis of, 220-221, 255-260 length of, 36, 36f planarity of, 36, 36f torsion angles for, 37, 37f, 38f trans configuration of, 36-37, 36f, 37f

INDEX

Peptide ligation, 96 Peptidoglycans, 244, 244f Peptidyl transferase center, 903-904 Perforin, 996 Perilipin A, 643 Peripheral membrane proteins, 355-356, 355f Periplasm, 365 Peroxide bridge, in cytochrome c oxidase, 539f, 540, 540f Peroxide detoxification, 727 Peroxisomes, 593 fatty acid metabolism in, 652, 653f Perutz, Max, 199 PFK. See Phosphofructokinase (PFK) P-glycoprotein, 378 pН buffers and. 15-17 definition of, 13 oxygen affinity of hemoglobin and, 206–207, 206f physiological, 16 Phagocytes, 978 Pharmacology. See also Drug development definition of, 1030b Phase I transformations, in oxidation, 1035 Phase II transformations, in conjugation, 1035 Phenyl isothiocyanate, in Edman degradation. 81. 81 Phenylacetate, 689 Phenylalanine degradation of, 695-696 in phenylketonuria, 697-698 structure of, 28, 29f, 696 synthesis of, 719-720, 720f, 721f Phenylalanine hydroxylase, 695 deficiency of, in phenylketonuria, 697-698 Phenylalanine operon, 932–933, 932f Phenylketonuria (PKU), 674, 697-698 Phenylpyruvate, 720, 721 Pheromones, 974 Phi (ϕ) angle of rotation, 37–38, 37f, 38f Phosphatase(s) deficiency of, 514 in signal transduction, 415 Phosphatase and tensin homolog, 799 Phosphate carrier, 554, 554f Phosphate, in myosin movement, 1015 Phosphate solutions, as buffers, 16 Phosphatidate, 348, 348, 762 in membrane lipid synthesis, 760, 761, 761f synthesis of, 760 Phosphatidic acid phosphatase (PAP), 761, 766-767, 766f Phosphatidylcholine, 349, 349 melting temperature of, 362t synthesis of, 763, 763f Phosphatidylethanolamine, 349, 349 synthesis of, 763, 763 Phosphatidylethanolamine methyltransferase, 763 Phosphatidylglycerol, 349, 349

Phosphatidylinositol, 349, 349, 762, 762.763 Phosphatidylinositol 3,4,5-trisphosphate (PIP₃), in signal transduction, 414, 799. 799f Phosphatidylinositol 4,5-bisphosphate (PIP₂), 762 in signal transduction, 408, 409f, 414, 799, 799f Phosphatidylserine, 349, 349 synthesis of, 763 3'-Phosphoadenosine-5'-phosphosulfate (PAPS), 1035 Phosphocreatine, synthesis of, 236-237 Phosphodiester bridges, 110, 110, 111 hydrolysis of, 272-274 Phosphoenolpyruvate, 434, 445 in amino acid synthesis, 719, 720f formation of, 445, 465 in gluconeogenesis, 480f, 481, 482, 483-484 in glycolysis, 464f, 465 NAD^+ regeneration in, 466–468 phosphoryl-transfer potential of, 434, 434t, 465 Phosphoenolpyruvate carboxykinase, 488. 488f Phosphofructokinase (PFK), 473 in gluconeogenesis, 487 in glycolysis, 455f, 458 in liver, 475-476, 475f, 476f, 487, 488 in muscle, 473–474, 473f Phosphofructokinase 2, 487 Phosphoglucomutase, 471, 620, 621f 6-Phosphogluconate, in pentose phosphate pathway, 601, 603 6-Phosphoglucono- δ -lactone, in pentose phosphate pathway, 601-603, 603 Phosphoglucose isomerase, 457 2-Phosphoglycerate, 445 in glycolysis, 445, 464, 466t 3-Phosphoglycerate, 464 in Calvin cycle, 591-592 in cysteine synthesis. 714-715 in glycine synthesis, 714-715 in glycolysis, 463-464, 464f, 466t in serine synthesis, 714–715, 714f, 723 3-Phosphoglycerate dehydrogenase, 723, 723, 724, 724f Phosphoglycerate kinase, in glycolysis, 463-464 Phosphoglycerate mutase in glycogen metabolism, 464, 620 in glycolysis, 464, 464f Phosphoglycerides, 348, 349, 351, 351f Phosphoglycolate, in Calvin cycle, 593, 593-594 3-Phosphohydroxypyruvate, 714 in proline synthesis, 714, 714f Phosphoinositide cascade, 408-409, 409f in glycogen metabolism, 626 Phospholipase C, in signal transduction, 408.409f

Phospholipid(s). See also Lipid(s) membrane, 352-355. See also Membrane(s); Membrane lipids phosphoglycerides, 348, 349 structure of, 348, 348-349 synthesis of, 760-767, 760f 5-Phosphomevalonate, 768 N-Phosphonacetyl-L-aspartate (PALA), 293, 293-294 Phosphopentose epimerase, 596, 596f in Calvin cycle, 596, 596f in pentose phosphate pathway, 603, 604t Phosphopentose isomerase, in Calvin cycle, 596, 596f Phosphoribomutase, 752 5-Phosphoribosyl-1-pyrophosphate (PRPP), 720-721, 721, 755 in purine synthesis, 741, 751 in pyrimidine synthesis, 738-739 Phosphoribosylamine, 741, 742 5-Phosphoribosylanthranilate, 721, 722 Phosphoribulose kinase, 596, 596f Phosphorolysis, in glycogen metabolism, 617-621 Phosphorothioates, in DNA cleavage, 273-274, 274f Phosphoryl group transfer, 432-434, 444 active carriers in, 438-442, 438f, 439f ATP and, 434-435, 434f, 434t, 436-437, 437f in glycolysis, 444, 444f Phosphorylase. See Glycogen phosphorylase Phosphorylase a, 622-623 Phosphorylase b, 622–623 Phosphorylase kinase, in glycogen metabolism, 623-624, 624f, 629 Phosphorylaspartate, 374, 374 Phosphorylation, 298-301 in amino acid synthesis, 724 ATP in, 298-301, 432-434, 436-437, 437f, 444 in chemotaxis, 1024, 1025f in cholesterol regulation, 772 in citric acid cycle, 513-514 in enzyme regulation, 298-301 in fatty acid metabolism, 666-667, 666f in gluconeogenesis, 483-484, 485 in glycogen metabolism, 617-618 in glycogen synthesis, 629-630 in glycolysis, 455-457, 456f, 457, 458 in immune response, 989-990, 989f kinases in, 457 in metabolism, 432-434, 444 oxidative, 436-437, 525-561. See also Oxidative phosphorylation of phosphorylase kinase, 623-624, 624f phosphoryl-transfer potential and, 434-435, 434f, 434t protein kinases in, 298-301 in purine synthesis, 741-743, 742, 742f regulatory functions of, 300-301

C35 Index

in signal transduction, 299t, 406-407, 407f. 408-409, 409f, 411-415, 413f, 417, 418 substrate-level, 463-464, 507-508 of sugars, 326-327 Phosphorylation potential, 447 Phosphorylcholine, antibody binding of, 985-986, 985f Phosphorylethanolamine, synthesis of, 763, 763 Phosphoryl-group-transfer potential, 434f, 434t Phosphoryl-transfer potential, 434-435 Phosphoserine, 57, 58, 714, 714 Phosphothreonine, 58 Phosphotriesters, in DNA synthesis, 144 Phosphotyrosine, 58 Photoinduced charge separation, 569-572, 569f Photophosphorylation, cyclic, 580-581, 580f Photoreceptors, 966 cones, 966, 969-970 evolution of, 970, 970f rods, 966-967, 966f Photorespiration in C₄ pathway, 599-600 in Calvin cycle, 593f, 594 Photosynthesis, 565-586 accessory pigments in, 581-584, 582f, 583f ATP synthase in, 578-579, 583, 583f in autotrophs, 590 basic equation of, 565 C₄ pathway in, 599-600, 600f carotenoids in, 582-583 chlorophyll a in, 568-572, 568f, 569f, 582f, 583, 592 chlorophyll b in, 582, 582f, 583 in chloroplasts, 567-568, 567f components of, location of, 583-584, 583f cyclic photophosphorylation in, 580-581, 580f dark reactions of, 566, 580, 589, 590-601. See also Calvin cycle coordination with light reactions, 599 definition of, 565 electron transfer in, 568-572, 569f, 571f energy conversion in, 566-567 evolution of, 568, 584, 600 herbicide inhibition of, 584, 584 light reactions of, 566-567, 566f, 572-584, 589 coordination with dark reactions, 599 in photosystem I, 567, 572, 572f, 574f, 575-577, 575f, 583, 583f in photosystem II, 567, 572-577, 572f-575f, 583, 583f stoichiometry of, 580-581 yield of, 580-581 light-harvesting complexes in, 583, 583f photoinduced charge separation in, 569-572, 569f proton-motive force in, 566, 566f, 578-579

O cycle in, 575, 575f reaction centers in, 569-572, 570f energy transfer to, 581-582, 582f resonance energy transfer in, 581-582, 582f in thylakoid membranes, 567-568, 567f, 574-575, 577-581, 583, 583f in tropical plants, 599-600 vs. oxidative phosphorylation, 578, 579f yield in, 566b Z scheme of, 577, 577f Photosynthetic bacteria, 584, 584t reaction center of, 569-572, 570f Photosynthetic catastrophe, 567b Photosynthetic vield, 566b Photosystem I, 567, 572, 572f, 574f, 575-577, 575f location of, 583, 583f Photosystem II, 567, 572-577, 572f-575f location of, 583, 583f Phototrophs, 428 Pigments accessory, in photosynthesis, 581-584, 582f, 583f visual, 969-970, 970f. See also Color vision in cones. 969–970 in rods, 966-967 Ping-pong reactions, 237 Pinopsin, 970, 970f PIP₂ (phosphatidylinositol 4,5trisphosphate), in signal transduction, 414, **414** PIP₃ (phosphatidylinositol 3,4,5bisphosphate), in signal transduction, 414, 414 pK_a values, 15, A2 PKU (phenylketonuria), 674, 697-698 Planar bilayer membrane, 354, 354f Plants C₃, 599 $C_4, 600$ crassulacean acid metabolism in, 600-601, 600f genetically engineered, 166-167, 167f glyoxylate cycle in, 518-519, 519f lectins in, 339 nitrogen fixation in, 706-710 photosynthesis in. See Photosynthesis starch in, 597 sucrose synthesis in, 597, 598f tropical, photosynthesis in, 599-600 Plaque, atherosclerotic, 776, 777f Plaques, in genomic library screening, 153, 153f Plasma cells, 979 Plasmid(s), 148 in drug resistance, 1050 pBR322, 149, 149f pUC18, 149f tumor-inducing, 166-167, 167f

Plasmid vectors, 149-151, 149f, 150f Plasmin, 312 Plasmodium falciparum glycan-binding proteins of, 340 mitochondrial genome of, 527 Plastocyanin, 575, 584 Plastoquinol, 573, 573, 575 Plastoquinone, 572-573, 573, 573, 584 Pleckstrin homology domain, 414, 419 P-loop, 283–284, 284f P-loop(s), in G proteins, 405 P-loop NTPases ABC proteins as, 378-380 in G proteins, 405 in microtubules, 1019, 1019f in motor proteins, 1008–1012, 1009f PLP. See Pyridoxal phosphate (PLP) Pluripotent stem cells, 944 Point mutations, production of, 157 Poisons mushroom, 865, 865–866 protein inhibition by, 910-911 Polar head group, 351, 352 Polarization, membrane, action potential and, 382 Polyacrylamide gel electrophoresis, 71–73, 72f, 73f Polyampholytes, in isoelectric focusing, 73 Polycistronic mRNA, 901 Polyclonal antibodies, 86, 86f Polygalacturonase, 167 Polylinkers, 150, 150f Polymerase chain reaction (PCR), 145–147, 145f-147f in ancient DNA amplification, 147, 188, 189f procedure for, 145-146, 145f quantitative, 161-162, 162f real-time, 161-162, 162f uses of, 146-148 Polymerase switching, 835 Polymorphisms, 147 Polypeptide chains, 34-35, 34f, 35f, 36-38. See also Protein(s) cleavage of, 82-83, 82f cross-linked, 35, 35f disulfide bonds of, 35, 35f formation of, 903-904, 904f. See also Translation side chains of, 34, 34f subunits of, 48, 48f Polypyrimidine tract, 873, 873f Polyribosomes, 906, 906f Polysaccharides, 328, 328f, 329f abbreviations for, 330b cellulose, 328-329 glycogen, 328, 329f starch, 328, 329f Polysomes, 906, 906f Poly(A) tail, 871, 872 Polyunsaturated fatty acids. See also Fatty acid(s) eicosanoids from, 664-665, 665, 665f

INDEX

Pompe disease, 634t, 635, 635f Pores, nuclear, 365 Porins, 47, 47f, 357, 357f mitochondrial, 526-527 Porphobilinogen, 728, 729 Porphobilinogen deaminase, 728-729 Porphyrias, 730-731 Porphyrins disorders of, 730-731 synthesis of, 728-730, 729f Potassium ion channels, 364-393, 382. See also Ion channels inactivation of, 388-389, 389f rate of transport in, 387 selectivity filter in, 385-386, 386f, 387f sequence homologies of, 384, 384f, 386 specificity of, 385-387 structure of, 384-385, 388f voltage-gated, 387-388, 388f Potassium ions, in Na⁺–K⁺ pump, 374, 377. See also Ion pumps Potential energy, 11. See also Energy Power stroke, in myosin, 1015 pppA, 854, 854 pppG, 854, 854 Predisposition, to disease, 19-20 Pregnenolone corticosteroids from, 783 progesterone from, 783 structure of, 784 synthesis of, 783 Pre-mRNA, processing of, 870–872, 871f, 878-879 splicing in, 873-879, 875f-878f. See also Splicing Prephenate, 720, 721 Prepriming complex, 834, 834f Pre-rRNA, processing of in bacteria, 863 in eukaryotes, 869-870, 870f Pressure, sensation of, 973 Pre-tRNA, processing of in bacteria, 871f in eukarvotes, 870-872, 871f in prokarvotes, 870 Prevacid (lansoprazole), 1046, 1046 Pribnow box, 126f, 127 Prilosec (omeprazole), 1046, 1046 Primary active transport, 371 Primary messengers, 402 Primary protein structure, 33-38. See also Amino acid sequences Primase, 823, 823f Primers in DNA replication, 122, 145, 821 in polymerase chain reaction, 145 Prions, 56-57, 56f Probes, DNA, 80, 142, 144-145, 151 - 153generation of, 144-145, 151-153 solid-phase approach in, 144-145, 144f for genomic library, 151-153, 152f Procaspases, 303

Processivity definition of, 832b in DNA replication, 831–832 Procollagen, 303 Procollagenase, 303 Proenzymes. See Zymogens Progestagens. See also Steroid hormones synthesis of, 780, 780f Progesterone aldosterone from, 783 cortisol from, 783, 784 synthesis of, 783, 784 Programmed cell death mitochondria in. 559 T cell, 1000, 1000f zymogens in, 303 Proinsulin, 303, 802 bacterial synthesis of, 155, 155f Prokaryotes, 3. See also Bacteria Proliferating cell nuclear antigen, 836 Proline hydroxyl groups of, 44, 57, 57-58 structure of, 28, 29f synthesis of, 714, 714f Proline racemase, inhibition of, 243, 243f Prolyl hydroxylase, 812-813 Prolyl hydroxylase 2, in cancer, 515 Promoters, 126-127, 126f, 852 bacterial, 852, 856-858, 857f eukarvotic, 866–869, 866f for RNA polymerase I, 866, 866f for RNA polymerase II, 866-867, 866f, 867-868 for RNA polymerase III, 866, 866f heat-shock, 858, 858f nitrogen-starvation, 858, 858f Proofreading evolution of, 895 in replication, 839-840, 839f in transcription, 856 in translation, 895 Proopiomelancortin (POMC), 796 PROP (6-n-propyl-2-thiouracil), 963 Propionyl CoA, 649, 649-650, 693, 693, 694 conversion to succinyl CoA, 649-650, 650f 6-n-Propyl-2-thiouracil (PROP), 963 Prostacyclins, 664-665, 665, 665f Prostaglandin E2, 665, 665 Prostaglandin H₂, 357–358, 358, 358f, 665, 665f Prostaglandin H₂ synthase-1, 357–358, 358, 358f Prostaglandin inhibitors, 358, 665, 1040 aspirin as, 358, 665, 1040 Prostaglandins, 664–665, 665, 665f Prosthetic groups, 221 Protease(s) active sites of, 185–186, 185f, 257–258 convergent evolution of. 185. 185-186 aspartyl, in peptide bond cleavage, 263, 264, 264f cysteine, in peptide bond cleavage, 263, 263-264, 264f

metal ion, in peptide bond cleavage, 263, 264,264f in peptide bond cleavage, 255-266. See also Peptide bonds, cleavage of in carbohydrates, 337 serine, in peptide bond cleavage, 255-266 site-directed mutagenesis of, 262-263, 262f specificity of, 260, 260f Protease inhibitors, 306-307 α_1 -antitrypsin, 307 pancreatic trypsin inhibitor, 306 pharmaceutical, 264-266, 265, 358, 665 development of, 1043, 1043f resistance to, 1050 Proteasomes, 677-678, 677f, 678f, 992. See also Amino acid degradation eukaryotic, 677-678, 678f evolution of. 677-678, 678f, 679 of M. tuberculosis, 679 prokaryotic, 678, 678f Protein(s), 2. See also Peptide(s) acetylation of, 57 adaptor, 414 in signal transduction, 414, 417 aggregated, in neurological diseases, 56-57, 56f allosteric, 289, 290-296. See also Allosteric enzymes α helix of. 38-40. **39.** 39f. **40.** 40f amino acids of. See Amino acid(s) antibody-tagged, 88 β sheet of, 40–42, 40f–42f carbohydrate units of, 57, 329-337. See also Glycoproteins cleavage of, 49, 58, 82-83, 82f, 83f coiled-coil, 43, 44f covalent modification of, 57-58, 57f, 297-302, 298t. See also Phosphorylation definition of. 34 degradation of, 673-700. See also Amino acid degradation denatured, 49-50, 49f, 50f, 52, 52f dietary, digestion and absorption of. 255-260. 303-307. 303t. 304f. 674, 674f. See also Amino acid degradation evolution of, 133 experimental studies of. See Protein studies fibrous. 43-45 flexibility of, 26-27, 26f folding of, 18, 19f, 38, 48-49, 52-57 cooperative transition in, 52 cumulative selection in, 53 denaturation and, 49f, 50, 50f, 52, 52f heat shock protein 70 in, 183, 183-184 hydrophobicity and, 45-46, 46f, 47, 47f intermediates in. 52-54, 53f Levinthal's paradox in, 53 mistakes in, in neurological diseases, 56-57 nucleation-condensation model of, 53

C37 Index

prediction of, 54 into regular structures, 38-45 steps in, 53-54, 53f function of, structural correlates of, 25-26, 26f, 57-59 functional groups of, 26 genes encoding, 19 genetically engineered, 157 glycan-binding, 337-338 glycosylation of, 329-337 congenital disorders of, 336 sites of, 333-334, 337 half-life of, 675, 676t hemoglobin as model of, 195-216. See also Hemoglobin hydroxylation of, 30, 30f, 57-58 identification of, 65-66 immunological methods for, 86 intrinsically unstructured, 54-55 as linear polymers, 33-35 lysosomal, 911 mass of, 34 measurement of electrophoresis in, 74, 74f mass spectrometry in, 91–95, 92f, 93f, 95f membrane. See Membrane proteins membrane-spanning, 911 metabolism of, 437f metamorphic, 55, 55f molecular models of, 21-22, 23f, 60-62, 61f, 62f molecular weight of, 34 multiple conformations of, 54-55, 55f Ω loops in, 42, 43f peptide bonds of, 33, 33f, 36-37, 36f, 37f polypeptide chains of, 33-35, 34f, 35f, 36-38 post-translational modification of, 57-59, 58f properties of, 25-26 random-coil conformation of, 49, 49f refolding of, 50, 50f regulation of covalent modification in, 298t phosphorylation in, 298-301. See also Phosphorylation reverse turns in, 42, 43f ribosomal. See also Ribosome(s) in translation, 900 rigidity of, 26, 26f secretory, 911 signal sequences in, 79 similarity among, 2, 2f sorting of, in Golgi complex, 334, 334f structure of, 2f, 3 amino acid sequences and, 18-19, 19f, 25, 35-36, 49-59 functional correlates of, 25-26, 26f, 57 - 59models of, 21-22, 23f, 60-62, 61f, 62f primary, 25, 33-38. See also Amino acid sequences

secondary, 25, 38-45, 47 supersecondary, 47, 47f tertiary, 45-47, 46f, 47f, 183-184. See also Protein(s), folding of subunits of, 48, 48f synthesis of. See also Translation initiation of, 130 mRNA in, 123-124, 130, 130f synthetic, 96-98, 96f, 97f, 157 translocation of, 911-914 unfolding of, 52, 52f Protein blots, 89, 142 Protein Data Bank, 60, 104 Protein domains, 47, 47f exon encoding of, 133, 133f Protein kinase(s), 298 calmodulin-dependent, 410-411 cyclin-dependent, 836 dedicated, 299 in enzyme regulation, 298-301 multifunctional, 299 in phosphorylation, 298-301 regulation of, cAMP in, 301, 301f serine, 298-299, 299f, 299t in signal transduction, 299t, 406-407, 408, 409, 409f, 419 specificity of, 299 threonine, 298-299, 299f, 299t tyrosine, 298-299, 299f in cancer, 420 Protein kinase A pseudosubstrate binding to, 302, 302f regulation of, 301, 301f in signal transduction, 406-407, 419 in glycogen metabolism, 624, 625f, 626 structure of, 302, 302f Protein kinase B (Akt), in signal transduction, 414, 799, 799f Protein kinase C, in signal transduction, 409.409f Protein kinase inhibitors, for cancer, 421 Protein phosphatase 1 (PP1), in glycogen metabolism, 626-627, 631-634, 631f Protein phosphatases, 300 Protein purification, 65, 66-79 affinity chromatography in, 70-71, 70f assays in, 67 autoradiography in, 73 dialysis in, 69, 69f differential centrifugation in, 67-68, 68f fractionation in, 67-68, 68f gel electrophoresis in, 71-73, 72f, 73f gel-filtration chromatography in, 69, 69f gradient centrifugation in, 77-78, 77f high-pressure liquid chromatography in, 71, 71f homogenization in, 67, 75t ion-exchange chromatography in, 69-70, 70f, 80-82, 80f isoelectric focusing in, 73, 73f with SDS-PAGE, 74, 74f monoclonal antibodies in, 87-88

guaternary, 26, 48, 48f

protein concentration in, 67 guantitative analysis of, 75-76, 75f, 75t recombinant DNA technology in, 78-79 salting out in, 68, 75t SDS-PAGE in, 74, 74f, 75, 75f with isoelectric focusing, 74, 74f sedimentation-equilibrium technique in. 78 two-dimensional electrophoresis in, 74, 74f Protein regulation covalent modification in, 298t phosphorylation in, 298-301. See also Phosphorylation Protein sequencing. See Amino acid sequences, determination of Protein sorting, 911-914 Protein studies amino acid sequencing in, 79-84. See also Amino acid sequences, determination of electron-density maps in, 99-100, 100f electrospray ionization in, 91-92 enzyme-linked immunosorbent assay in, 88-89, 89f fluorescence microscopy in, 90-91, 90f genomic analysis in, 84 immunoelectron microscopy in, 91, 91f immunologic techniques in, 84-91. See also Monoclonal antibodies mass spectrometry in, 91-95, 92f, 93f, 95f nuclear magnetic resonance spectroscopy in, 101-104, 101f-103f nuclear Overhauser enhancement spectroscopy in, 101-104, 102f, 103f proteomic analysis in, 84, 94-95, 95f purification methods for, 65, 66-79. See also Protein purification recombinant DNA technology in. See Recombinant DNA technology synthetic peptides in, 96-98, 96f, 97f time of flight analysis in, 92, 92f western blotting in, 89, 89f, 142 X-ray crystallography in, 98-100, 98f-100f Protein targeting, 334, 334f, 911-914 Protein transport, 911-914. See also Membrane transport Protein turnover, 255, 673. See also Amino acid degradation; Amino acid synthesis Protein tyrosine kinases, 298–299, 299f, 412 in cancer, 420 receptor, 412 Proteins, synthesis of, DNA technology in, 78-79 Proteoglycans, 329-330, 331f Proteolytic enzymes, 220–221, 302–313 activation of, 302-313 in blood clotting, 307-312 chymotrypsinogen as, 303-305 in digestion, 303-307 functions of, 303 inhibitors of, 306-307 in starvation, 809, 810

C 3 8

INDEX

Proteomes, definition of, 66 Prothrombin, 309f, 310, 310f calcium binding by, 310-311 Proton gradients, 437 free energy from, 559-560, 560f in oxidative phosphorylation, 530-531, 543-550, 559-560, 560f in photosynthesis, 571-572, 578-581 Proton pumps in oxidative phosphorylation, 525 in respiratory chain, 525 Proton shuttle, in carbon dioxide hydration, 269-270, 270f Proton transport, by flagella, 1023, 1024f Proton-motive force, 525 in oxidative phosphorylation, 525, 544-545, 544f, 569 in photosynthesis, 566, 566f, 578-579 Protons, chemical, 540f Proto-oncogenes, 420 Protoplasts, 167 Protoporphyrin, 196, 196-197 Protoporphyrin IX, 729, 730 Provitamin D₃, 785, 785 Proximal histidine, 197 PRPP. See 5-Phosphoribosyl-1pyrosphosphate (PRPP) Pseudo-first-order reactions, 230 Pseudogenes, 159 Pseudosubstrate sequence, 302 Pseudouridylate, 863, 863 Psi (Ψ) angle of rotation, 37–38, 37f, 38f Psoralen, 876 PTEN, 799 P-type ATPases, 374 evolution of, 378-379 functions of, 378 in membrane transport, 374-381. See also Ion pumps types of, 378 pUC18 plasmid, 150f Puffer fish genome of, 161, 161f tetrodotoxin of, 384, 384 Pumps. See Ion pumps; Proton pumps pur repressor, 927, 927f Purine nucleotides, 111, 111–112 degradation of, 752-755, 753f derivatives of, 111, 111 synthesis of, 726, 740-745, 740f, 751. See also Nucleotide(s), synthesis of phosphorylation in, 741-743, 742f steps in, 740-743, 741t, 742f Purinosomes, 744, 744f Puromycin, 909t, 910, 910 Pyranose, 322-325, 323f Pyridoxal phosphate (PLP), 674 in amino acid degradation, 657, 681, 681-684 in amino acid synthesis, 712f, 713 in glycogen metabolism, 618-619, 618f

Pyridoxal phosphate enzymes in amino acid degradation, 681–684, 683f in amino acid synthesis, 712f, 713 catalytic activity of, 683-684, 683f Pyridoxal phosphate-dependent transaminases, 712-713 Pyridoxamine phosphate, 682, 682, 712 Pyridoxine (vitamin B₆), **441**, 441t, 681 Pyrimidine(s), 111, 111 derivatives of, 111, 111 synthesis of, 726 aspartate transcarbamoylase in, 290-296, 738 Pyrimidine nucleotides, 111, 111 recycling of, 740 synthesis of, 736–740, 736f, 751. See also Nucleotide(s), synthesis of Pyrimidine phosphoribosyltransferase, 739 Pyrophosphate, hydrolysis of, 627-628, 645 in fatty acid metabolism, 645 5-Pyrophosphomevalonate, 768 Pyrrole-2-carboxylic acid, 243, 243 Δ^1 -Pyrroline 5-carboxylate, in proline synthesis, 714 Pvruvate, 236 in amino acid degradation, 684-685, 685f, 691 in amino acid synthesis, 712 in citric acid cycle, 500-501 conversion to acetyl coenzyme A, 500-501, 500f, 501f in Cori cycle, 490, 490f, 685 in ethanol formation, 466–467, 467f formation of, 691, 691f in gluconeogenesis, 453, 479-485 in glucose–alanine cycle, 685, 685f in glycolysis, 453, 464–466, 464f–466f metabolic fates of, 466-468, 466f in nitrogen transport, 684-685, 685f oxaloacetate synthesis from, 443-444, 443f Pyruvate carboxylase in citric acid cycle, 516 in gluconeogenesis, 481 Pvruvate carrier, 554, 554f Pyruvate decarboxylase, 466 Pyruvate dehydrogenase complex in citric acid cycle, 499-503, 499f, 499t, 502f, 503f components of, 499, 499t phosphorylation of, 513-514 reactions of, 502-503, 503f regulation of, 513-514 structure of, 502, 502f Pyruvate dehydrogenase kinase, in cancer, 515 Pyruvate kinase, in glycolysis, 465 in liver, 476–477, 476f in muscle. 474 Pyruvate-P; dikinase, in C4 pathway, 599 Q (coenzyme Q), 531, 532–533, 533f in fatty acid metabolism, 646

O cycle, 575, 575f in oxidative phosphorylation, 536-537 in photosynthesis, 575 Q pool in oxidative phosphorylation, 533 in photosynthesis, 571f, 572 Q_A, 571–572, 571f, 573 Q_B, 571, 571f, 573 Q-cytochrome c oxidoreductase (Complex III), 531, 532f, 532t, 535-536, 536f QH₂, 571–572 Quantitative polymerase chain reaction, 161-162, 162f Ouaternary protein structure, 26, 48, 48f Ouinine, 962 taste of, 962 Ouinones in oxidative phosphorylation, 531, 533. 533f in photosynthesis, 570, 571 Quinonoid dihydrobiopterin, 695, 695 Quorum sensing, 930-931, 930f biofilms and, 930-931 R* (metarhodopsin II), 968, 968f R groups. See Amino acid side chains R state, of hemoglobin, 202-204, 203f Radiation, mutagenicity of, 839 Raloxifene, 948, 948 Ramachandran diagrams for angles of rotation, 37, 38f for helices, 39-40, 40f Ramachandran, Gopalasamudram, 37 Random-coil conformation, 49, 50f Ranitidine (Zantac), 1045, 1045 Ras farnesylation of, 298t in signal transduction, 417-418, 418f Rate constant (k), 230 Rate, reaction. See Reaction rates RBP4, 796 Reaction(s) activation energy for, 222, 226 anabolic, 428-429 anaplerotic, 516-517 biomolecular, 230 bisubstrate, 236 catabolic, 428, 435-438 electron carriers for, 438-439 catalysis of. See Catalysis; Enzyme(s) chemical modification, 256 Cleland notation for, 236, 237 coupled, thermodynamics of, 429-430, 431-433 double-displacement (ping-pong), 237 electrocyclic, 720 endergonic, 222 enzyme-catalyzed. See Catalysis; Enzyme(s) exergonic, 222 first-order, 229 free energy change of, 12, 222-225 group-transfer, 436-437, 443t, 444

interconnected, 428-430, 428f ligation, 443-444 lvase-catalyzed, 445 metabolic, 428-430. See also Metabolism multiple-substrate, 235-237 oxidation-reduction, 443, 443t free energy change in, 530 in metabolism, 433t, 443 reduction potential of, 528-530, 529t second-order, 230 sequential, 236-237 thermodynamics of, 222-225 transition state in, enzymes and, 225-226, 225f, 227-228 Reaction centers, in photosynthesis, 569-572, 570f, 582, 582f energy transfer to, 581-582, 582f Reaction equilibrium, enzymes and, 224-225 Reaction rates, 229-237. See also Kinetics catalytic acceleration of, 220, 220t, 225-229, 226f definition of, 229 in enzyme-catalyzed reactions, 229-237. See also Enzyme kinetics in first-order reactions, 229-230 maximal, calculation of, 233 Michaelis constant (K_M) for, 231-235 in pseudo-first-order reactions, 230 in second-order reactions, 230 Reactive oxygen species (ROS), 541-542, 541t cytochrome P450 and, 782-783, 782f glucose 6-phosphate and, 610 mutagenicity of, 838 Reactive substrate analogs, 241-242 Reading frame, 130 Reagents, group-specific, 241 Real-time polymerase chain reaction, 161-162, 162f *RecA*, 845 Receptor(s). See also specific types ligand binding to, 926, 947 in signal transduction, 402 G-protein-coupled, 404-406 insulin, 412-415 serpentine, 404 7TM, 404-406, 404t Receptor tyrosine kinases, in signal transduction, 412 in cancer, 421 Reclinomonas americana, genome of, 528 Recognition helix, 923 Recognition sites (sequences), 141, 141f, 272, 275-277 in cognate vs. noncognate DNA, 275–277, 276f distortion of, 276-277, 276f inverted repeats as, 275 structure of, 275, 275f Recombinant DNA production of, 154-155, 154f transfer of, by microinjection, 163, 163f vectors for, 163-164. See also Vectors

Recombinant DNA technology, 139-169 in amino acid sequencing, 84 autoradiography in, 73, 143 bacterial artificial chromosomes in, 151 bombardment-mediated transformation in. 167 cloning in, 149-151, 149f-151f cohesive-end method in, 148-149, 149f complementary DNA in, 154-155, 154f, 162f designer genes in, 157 for disease-causing mutations, 157 DNA microarrays in, 162-163, 162f DNA microinjection in, 163, 163f DNA probes in, 80, 142, 144-145, 151-153 DNA sequencing in, 143-144, 143f DNA synthesis in, 144-145, 148-149 electroporation in, 167, 167f expression vectors in, 154-155 fluorescence detection in, 143-144, 143f gel electrophoresis in, 141-142, 142f gene guns in, 167 gene manipulation in, 161-168 gene-expression analysis in, 161–162, 162f, 163-164 genomic libraries in, 151-153, 152f, 153f lambda phage in, 150-151, 150f, 151f medical applications of, 167-168 mutations in, 156-157 overview of, 139-141 in plants, 166-167, 167f plasmid vectors in, 149-151, 149f, 150f polymerase chain reaction in, 145-147, 145f-147f, 188 in protein purification, 78-79 restriction-enzyme analysis in, 141-142 solid-phase approach in, 144-145, 144f transgenic animals in, 164, 164f vectors in, 149-151, 149f-151f, 154-155 yeast artificial chromosomes in, 151, 151f Recombinases, 845-846 Recombination in antibodies, 987-991, 988f DNA. See DNA recombination homologous, 970b in color blindness, 970 in gene knockout, 164-165 Recombination signal sequences (RSSs), 988 Recombination synapse, 846 Red blood cells. See also Hemoglobin 2,3-bisphosphoglycerate in, oxygen affinity of hemoglobin and, 204-205 carbohydrates in, 335 disorders of, 209-211 Heinz bodies in, 610, 610f life span of, 730 sickled, 209-210, 209f Red photoreceptors, 969-970, 969f, 970f

Redox couples, 529 redox potential of, 529-530, 529t Redox potential. See Reduction potential Redox reactions. See Oxidation-reduction reactions Reducing sugars, 325 Reductase, in nitrogenase complex, 708-709 Reductase mRNA, in cholesterol regulation, 770, 771 Reduction potential measurement of, 528f, 529-530 in oxidative phosphorylation, 528-531 Reed, Randall, 959 Regulatory domains. See Domains Regulatory sites, 291 Rejection, in transplantation, 998 Relaxed DNA, 117, 117f Relay helix, 1012, 1012f Release factors (Rfs), 130, 906, 907f Renal function. See Kidney Renal gluconeogenesis, 481 Repeating motifs, 79 sequence homology of, 184, 185f Replication. See DNA replication Replication factor C, 836 Replication fork, 823, 823f, 832-833, 833f Replication protein A, 835 Replicon, 835 Repressors, 924-925 corepressors, 927 lac, 922f, 923, 925-928, 926f, 927 λ, 928–929, 928f, 929f pur, 927, 927f Resistin, 796 Resonance structures, 7 of ATP, 433 improbable, 433f of orthophosphate, 433 Respiration, cellular, 498, 499f, 593f, 594 definition of, 526b in photosynthesis, 593f, 594 regulation of, 554-560 Respiratory chain, 525, 531-543 coenzyme Q in, 531 components of, 531-532, 532f, 533f cvtochrome c oxidase (Complex IV) in, 531, 532f, 532t, 535-536 electron carriers in, 438-439, 438f, 439f linkage to ATP synthase, 544–545, 544f, 545f NADH-Q oxidoreductase (Complex I) in, 531-535, 532f, 532t Q cycle in, 536–537, 537f Q-cytochrome c oxidoreductase (Complex III) in, 531, 532f, 532t, 535-536. 536f succinate-Q reductase (Complex II) in, 531, 532f, 532t, 535 Respiratory control, 556 Respiratory distress syndrome, 765-766

INDEX

Restriction enzymes (endonucleases), 141-142, 271-279 binding affinity of, 275-277 in DNA cleavage, 141-142, 271-279. See also Restriction fragment(s) in E. coli, 272–279, 274f–277f evolution of, 278-279 recognition sites for, 141, 141f, 271-272, 275-277. See also Recognition sites (sequences) in restriction-modification systems, 272 specificity of, 141, 141f, 272, 275-277, 275f type II, 278–279, 278f Restriction fragment(s) separation of, gel electrophoresis in, 141-142 in Southern blotting, 142 Restriction fragment length polymorphisms, 147 Restriction-modification systems, 277 11-cis-Retinal, 967, 967, 968f all-trans-Retinal, 967, 968 Retinitis pigmentosa, 877-878 Retinol (vitamin A), 442, 442, 442t deficiency of, alcohol-related, 812 Retroviruses, 122–123, 123f, 999–1000 as vectors, 163 Reverse cholesterol transport, 778-779 Reverse transcriptase, 123, 123f in complementary DNA production, 154-155 Reverse turns, 42, 43f amino acid residues in, 50-51, 51t Reversible covalent modification, 446, 725-726, 725f RFs (release factors), 130, 906, 907f Rhizobium, in nitrogen fixation, 706-707 Rho, 127 in transcription termination, 860-861 Rhodopseudomonas viridis, photosynthetic reaction center in, 569-570, 570f, 584 Rhodopsin, 404, 966–968 absorption spectrum of, 967f cone photoreceptors and, 969-970, 970f structure of, 404, 404f Rhodopsin kinase, 968 Ribbon diagrams, 62, 62f Riboflavin (vitamin B₂), 441, 441t Ribonuclease amino acid sequences in, 49-50, 49f reduction and denaturation of, 49-50, 49f, 50f scrambled, 50, 50f sequence comparison for, 173, 174 structure of, 49-50, 49f, 50f in transcription, 863 Ribonuclease P, 863 Ribonuclease III. 863 Ribonucleic acid. See RNA Ribonucleoproteins small nuclear, 852, 875-876, 876t small nucleolar, 870, 875-877

Ribonucleotide reductase, 718, 745-750, 745f, 746, 752, 752f Ribose, 110, 110, 111, 321, 322, 322f ring form of, 323, 323f Ribose 1-phosphate, 752 Ribose 5-phosphate, 596 in Calvin cycle, 595f, 596, 596f in pentose phosphate pathway, 601, 602, 602f, 603f, 607-609 Ribosomal RNA. See rRNA (ribosomal RNA) Ribosome(s), 130, 897-907 in codon-anticodon interactions. 902 definition of. 887 endoplasmic reticulum-bound, 911-914 in eukaryotes vs. bacteria, 907 evolution of, 900 functions of, 900 initiation sites in, 902, 903f polyribosomal, 906, 906f as ribozymes, 887 structure of, 887f, 897-898, 898f subunits of, 897-898, 898f formation of, 902 tRNA binding by mechanism of, 900f, 903-904 sites of, 900, 900f, 902-904 Ribosome release factor, 906, 907f Riboswitches, 860, 860f Ribosvlation, 298t Ribothymidylate, 863, 863 Ribozymes, 879, 887 Ribulose 1,5-bisphosphate carboxylase/ oxygenase. See Rubisco Ribulose 1,5-bisphosphate, in Calvin cycle, 591-597, 592, 595f, 596, 596, 596f Ribulose 5-phosphate in Calvin cycle, 596, 596 in pentose phosphate pathway, 602, 602f Ricin, 911 Rickets, 786 Rickettsia prowazekii, genome of, 527-528 Rieske center, 536 Rifampicin, 861-862, 862 Rise, in α helix, 39 RNA, 111 ATP-binding region of, 190–191, 191f backbone of, 110, 110f catalytic activity of, 879-881, 900 evolution of, molecular studies of, 188-191, 190f in gene expression, 123-128 messenger. See mRNA micro, 124 nucleosides in, 111 post-transcriptional processing of in bacteria, 863 in eukaryotes, 869-879 pre-mRNA. 870–872. 871f. 878–879 splicing in, 873-879, 875f-878f pre-rRNA, 869-870, 870f in bacteria, 863 in eukaryotes, 869-870, 870f

pre-tRNA in bacteria, 870, 871f in eukaryotes, 870-872, 871f ribosomal. See rRNA self-splicing in, 879-881, 880f-882f sequence homologies of, 186-187, 187f small interfering, 124 small nuclear, 124, 875-876 in splicing, 875-876, 876t splicing of, 132-133, 132f, 851-852. See also Splicing stem-loop motif in, 117f structure of, 110, 111-112, 117-118, 117f, 186-187, 187f sugars in, 110f, 111 synthesis of, 124-127, 125f, 126f, 851-882. See also Transcription transcriptional modification of, 127, 127f transfer. See tRNA types of, 123-124 RNA blots, 142, 142f RNA editing, 852, 872-873, 873f RNA interference, 165-166, 167f RNA polymerase(s), 124–127, 125f, 852, 852,853-858 ADP ribosylation of, 298t backtracking and, 856, 856f bacterial, 853-858 active site of, 853, 854f promoter recognition by, 856-857 structure of, 852, 852, 853, 853t subunits of, 853, 853t, 856-858 catalytic action of, 853-858 in DNA replication, 823, 823f eukaryotic, 852, 864 promoter elements in, 866-869, 866f in proofreading, 856 RNA-directed, 122 structure of, 852, 853 RNA polymerase holoenzyme complex, 857, 857-858, 857f RNA polymerase I, 865, 865t, 866 rRNA processing by, 869-870, 870f RNA polymerase II, 865, 865t, 866-867, 943 bromodomains and, 949-950, 950f in transcription initiation, 950f RNA polymerase III, 865, 865t, 866 RNA processing, 132–133. See also Transcription in bacteria, 863 in eukaryotes, 869-879 of pre-rRNA, 869-870, 870f RNA polymerase I in, 869-870, 870f of tRNA, 870, 871f in prokaryotes, of tRNA, 870 RNA splicing. See Splicing RNA viruses, 122-123, 123f RNA world, 447, 881 RNA-directed RNA polymerase, 122 RNA-inducing silencing complex (RISC), 166 RNase P, 881 Roberts, Richard, 131

Index

Rods, 966-967, 966f Rofecoxib (Vioxx), 1044, 1044-1045, 1049 Rossman fold, 469, 469f, 509, 509f Rossman, Michael, 469 Rotation, flagellar, 1022-1025 Rotational catalysis, in ATP synthesis, 547-550, 547f-549f Rotenone, 558 Rough endoplasmic reticulum, 911. See also Endoplasmic reticulum Roundup, 719-720 Rous sarcoma virus, 420 rRNA (ribosomal RNA), 123t, 124. See also RNA in base-pairing, 892 bases in, 863 folding of, 898, 898f functions of, 124, 900 16S, in translation, 892, 901 transcription and processing of, 863, 869-870, 870f transcription of, 863 in translation, 892, 898-900, 901, 903-904 RsrI endonuclease, 278 RSS (recombination signal sequence), 988 Rubisco, 591, 591-597 evolution of, 600 light activation of, 598 Rubisco activase, 592 Saccharomyces cerevisiae, genome of, 158 Sakmann, Bert, 383 Salmonella test, for mutagens, 843-844 Salt bridges, oxygen release from hemoglobin and, 206-207, 207, 207f Salting out, 68, 75, 75t Salty taste, 962, 965 Salvage pathways, 740 purine, 744-745 pyrimidine, 736, 736f Sandwich ELISA, 89, 89f Sanger dideoxy method, 143-144, 143f Sanger, Frederick, 35, 143, 158 Sarcomere, 1016-1017, 1016f, 1017f Sarcoplasmic reticulum Ca²⁺ ATPase (SERCA), 374 SARS, drug therapy for, 1046–1047, 1047f Satiation signals, 795-797, 796f Saturated fatty acids. See Fatty acid(s) SCAP (SREBP cleavage activating protein), 770-771,771f Schiff bases, 681-682, 709 in amino acid degradation, 681-682, 709 in amino acid synthesis, 709 in aminoacrylate, 722, 722 in glycogen phosphorolysis, 619, 619f in retinal, 967-968, 968f in transaldolase reaction, 605-606, 606f in transamination, 618-682 Scrapie, 56 Screw sense, of helix, 39, 40f Scurvy, 57, 812-813

SDS (sodium dodecyl sulfate), in gel electrophoresis, 73, 78 SDS-PAGE, 74, 74f, 75-76, 75f with isoelectric focusing, 74, 74f SDS-polyacrylamide gel electrophoresis, of membrane proteins, 355, 355f Second Law of Thermodynamics, 11-12, 792. See also Thermodynamics Second messengers, 402-403, 403f, 409-411, 409f, 419 Secondary active transport, 371 Secondary protein structures, 38-45, 50-52, 51t alternative conformations of, 51-52, 51f prediction of, 51 Secondary transporters, 381, 381f Second-order reactions, 230 rate of. 230 Secretory proteins, 911 Sedimentation coefficients, 76, 76t, 77f Sedimentation-equilibrium technique, 78 Sedimentation-velocity technique, 78 Sedoheptulose 1,7-bisphosphate, in Calvin cycle, 595 Sedoheptulose 7-phosphate, 595 in Calvin cycle, 595f, 596f in pentose phosphate pathway, 602, 602f, 603, 603f Segmental flexibility, 982, 982f Seizures, ketogenic diet for, 656 Selectins, 339 Selective estrogen receptor modulators (SERMs), 948 Selectivity filters, 385-386, 386f, 387f Selenolate, 727, 727f Self-antigens, immune response to, 1001 Self-splicing, 879-881, 880f-882f. See also Splicing Self-tolerance, 1001 Semiconservative replication, 118-120, 120f Sense strand, 845, 854 Sensory systems, 957-975 hearing. 971-973 overview of. 957-958 smell. 958–962 taste, 962-966 touch. 973-974 vision, 966-971 yet-to-be-studied, 974 Sequence alignment, 175-181, 176f-182f Sequence homologies. See also Evolution analysis of, 175-181, 176f-182f of cytochrome c, 543, 543f evolutionary trees and, 187-188, 187f of hemoglobin and myoglobin, 175-181, 176f-182f, 199 of histones, 940-941, 940f identification of databases for, 181, 182f sequence alignment in, 175-181, 176f-182f shuffling in, 177-178, 177f

statistical analysis in, 175-177 substitution matrices in, 178-181 of ion channels, 384, 384f, 386 repeated motifs and, 184, 184 RNA secondary structures and, 186–187, 187f three-dimensional structure and, 183-184 of tubulin, 1019 of urea cycle enzymes, 688 Sequence specifies conformation, 50 Sequence template, 184 Sequential model of hemoglobin oxygen binding, 202–203, 203f, 216 of substrate binding, 294 Sequential reactions, 236-237 SERCA, 375-377 SERCA (sarcoplasmic reticulum Ca²⁺ ATPase), 374 Serine in catalytic triads, 258-260. See also Catalytic triads in chymotrypsin, 255-256, 256f in cysteine synthesis, 714-715, 719 from glycine, 692 in glycine synthesis, 714–715 in glycoproteins, 330 pyruvate formation from, 691, 691f in sphingolipid synthesis, 764, 764 sphingosine from, 726, 726f structure of, 30, 30f synthesis of, 714-715, 714f, 716, 723 Serine dehydratase, in amino acid degradation, 684, 691 Serine hydroxymethyltransferase, 715 Serine kinases, 298–299, 299f, 299t Serine proteases active sites on, convergent evolution of, 185, 185–186 catalytic strategies of, 253, 255-256 SERMs (selective estrogen receptor modulators), 948 Serotonin, synthesis of, 726f, 727 Serpentine receptors, 404 Serum albumin, in drug distribution, 1003f, 1033 7TM (seven-transmembrane-helix receptors), 404-406, 404f, 404t, 408, 408-409 for drugs, 404, 1045 in olfaction, 959 in phosphoinositide cascade, 408-409 in taste, 963-965 of bitterness, 962, 963-964 of sweetness, 964-965 in vision, 967-969, 968f, 969 70S initiation complex, 902, 903f 70S subunit, of ribosome, 897-898, 898f formation of, 902, 903f Severe acute respiratory syndrome (SARS), drug therapy for, 1046-1047, 1047f

INDEX

Severe combined immunodeficiency (SCID), 753 gene therapy for, 167–168 Sex hormones. See also Hormone(s) regulation of, 946 synthesis of, 780, 780f, 784-785, 784f SH2 domain, in signal transduction, 414, 417, 419, 420 SH3 domain, in signal transduction, 417, **420** Shaker channel, 388, 389f shaker gene, 384 Sharp, Philip, 131 Shemin, David, 728, 728b Shikimate 3-phosphate, 720 Shikimate, in amino acid synthesis, 719,720f Shine-Dalgarno sequences, 130, 901 Short interspersed elements (SINES), 160 Sialic acid, influenza virus and, 339, 340f Sialidase, 339, 340f Sickle-cell anemia, 209-210, 209f, 210f Side chains amino acid. See Amino acid side chains polypeptide, 34, 34f Side effects, 1031 genetic variations in, 1047-1048 σ subunit, of RNA polymerase, 853t, 857-858.868 Sigmoid binding curve, 200, 294-295.295f Sigmoidal kinetics, enzymatic, 200, 291, 291f, 294, 295f Signal peptidase, 912 Signal sequences, 79 nuclear localization signal, 79 in protein targeting, 912, 912f Signal transduction, 401-423 abnormalities in in cancer, 420-421 in cholera, 421-422 in whooping cough, 422 adaptor proteins in, 414, 417 β-adrenergic receptor in, 404-405.404f calcium in, 409-411, 409f-411f calmodulin in, 410-411, 411f in caloric homeostasis, 794–797, 795f-797f, 795t cAMP in, 406-407 in chemotaxis, 1024, 1025f cross talk in, 403 cytokines in, 998 definition of, 401 epidermal growth factor in, 402f, 415–418, 416f-418f epinephrine in, 402-410, 402f, 406f evolution of, 419 G proteins in, 417-418, 418t. See also G protein(s) gap junctions in, 393 in gene expression/regulation, 929-930

GTPases in, 298, 298t, 407-408 in hearing, 927f, 972 hormone-receptor complexes in, 405-406 in immune response, 989–990, 998 insulin in, 402f, 411–415, 798–800, 799f ligands in. 402 molecular circuits in, 402-403 nitric oxide in, 728 in olfaction, 959-960, 960f overview of, 401-402, 419 phosphoinositide cascade in, 408-409, 409f phosphorylation in, 298-299, 299t, 406-407, 408-409, 409f, 411-415, 417.419 primary messengers in, 402 principles of, 402f protein kinase A in. 406–407, 419 protein kinase C in. 408, 409 protein kinases in, 298-299, 299t, 419 in protein sorting, 911–914 Ras in, 417-418, 418f reactive oxygen species in, 542 receptor protein kinases in, 411 receptor tyrosine kinases in, 412, 415 receptors in, 402, 405-406 second messengers in, 402-403, 403f, 409-411, 409f, 419 7TM receptors in, 404-406, 404t specialized domains in, 419 Src homology domains in, 414, 414f, 417, 419, 420, 420 steps in. 402–403 in taste, 963–966 termination of, 407f in touch, 973-974 in vision, 968-969, 968f Signal-recognition particle (SRP), 124, 911, 912f receptor for, 911 Sildenafil, 1039 discovery of, 1039 Simple diffusion, 372 **SINES**, 160 Singer, S. Jonathan, 362 Single-strand-binding protein, 832 siRNA (small interfering RNA), 124. See also RNA Site-directed mutagenesis, in catalysis investigation, 262-263 16S rRNA, in translation, 892, 901 Sjöstrand, Fritjof, 526 Slack, C. Roger, 599 Sliding DNA clamp, 831-832, 831f, 834 Sliding filament model, 1016-1017, 1017f Small G proteins. See also G protein(s) in signal transduction, 418, 418t Small interfering RNA (siRNA), 124. See also RNA Small nuclear ribonucleoproteins (snRNPs). 852, 875-876, 876t Small nuclear RNA (snRNA), 124, 875-876. See also RNA in splicing, 875-876, 876t

Small nucleolar ribonucleoproteins (snoRNPs), 870 Smallpox, 1003 Smell, 958-962. See also Olfaction Smith, Emil, 543 Smith, Hamilton, 141 Smooth endoplasmic reticulum, 911. See also Endoplasmic reticulum SNARE proteins, 366, 914 snoRNPs (small nucleolar ribonucleoproteins), 870 snRNA (small nuclear RNA), 124, 875-876 in splicing, 875-876, 876t snRNPs (small nuclear ribonucleoproteins), 852, 862t, 875-876 Snurps, 875 SOD1 gene, 152, 157 Sodium dodecyl sulfate (SDS). See also under SDS in gel electrophoresis, 73, 78 Sodium ion channels, 382, 384, 384f, 387-392. See also Ion channels amiloride-sensitive, 965, 965f inactivation of, 388-389, 389f sequence homologies of, 384, 384f, 386 in taste, 962, 965 Sodium ion, salty taste of, 962, 965 Sodium-potassium pump, 364, 364f, 374, 377, 392 digitalis inhibition of, 377 evolution of, 378 Solid-phase method, 144f Solid-phase methods, 97f, 98, 144-145, 144f Solid-phase peptide synthesis, 97f, 98 Somatic mutations, antibody diversity and, 989 Sonication, 353 Sos, in signal transduction, 417 Sound, perception of, 971-973 Sour taste, 962, 962f, 965-966 Southern blotting, 142, 142f Southern, Edwin, 142 Space-filling molecular models, 21, 23f, 61.61f Special pairs, 571, 573, 573f Specific activity, 67 Sphingolipids functions of, 765 gangliosides, 764-765, 765f synthesis of, 764-765 Sphingomyelin, 349, 349, 351, 351f synthesis of, 764, 764 Sphingosine, 348, 349, 349, 764, 764 synthesis of, 726, 726f Spicy foods, capsaicin in, 973-974, 973f Spin, 101, 101t Spina bifida, 755 Spleen tyrosine kinase (Syk), 989–990 Spliceosomes, 132, 874, 875-877, 875f Splicing, 132-133, 132f, 851-852, 869, 873-879 alternative, 133, 133f, 878-879, 878f branch points in, 874, 875, 875f

C43 Index

consensus sequences in, 132f, 133, 873-874, 873f definition of, 873 errors in, 877-878 internal guide sequence in, 881, 881f mutations affecting, 877–878 self-splicing and, 879-881, 880f-882f sites of, 873-874, 873f, 875-876, 877, 881, 881f spliceosomes in, 132, 874, 875-877, 875f transesterification in, 874-875, 880 Splicing factors, 875 Split genes, 131-132, 132f, 133f evolutionary advantages of, 133 Split-pool synthesis, 1041, 1042f Squalene, 768-770 Squalene epoxide, 769, 770 Squalene synthase, 769 SR Ca²⁺ ATPase, 375–377 Src in cancer, 420 myristoylation of, 298t Src homology domains, 414, 414f, 417, 419, 420, **420** SREBP (sterol regulatory element binding protein), 770-771, 771f SREBP cleavage activating protein (SCAP), 770-771, 771f SRP (signal-recognition particle), 124, 911, 912f, 913f SRP receptor, 912, 913f SRP54, 911, 913 Stadtman, Earl, 725 Stahl, Franklin, 119 Staining, in gel electrophoresis, 73, 73f Staphylococcus aureus, peptidoglycan of, 244, 244f Starch, 328, 329f. See also Carbohydrates in plants, 597 Starvation, metabolic adaptations in, 809-810 Starved-fed cycle, 807-808 Statins, 779, 779, 1040, 1040–1041, 1041 Steady-state assumption, 231 Steatorrhea, 642 Steitz, Tom, 821 Stem cells, pluripotent, 944 Stem-loop motif, 117, 117f Stereochemistry, 21 notation in, 27 Stereocilia, 971-973, 971f Stereoisomers, 27 carbohydrate, 321, 321f Steric exclusion, 37 Steroid hormones. See also Hormone(s) anabolic, 785, 948 cardiotonic, 377, 377f functions of, 780 hydroxylation of, 781-782 structure of, 781, 781 synthesis of, 780f, 783-785 Steroid receptors, nuclear localization of, 90-91, 90f. See also Nuclear hormone receptors

Sterol regulatory element, 770 Sterol regulatory element binding protein (SREBP), 770–771, 771f Sticky ends, 148, 149f Stoma, in Crassulacea, 600f, 601 Stomach. See also under Gastric protein digestion in, 674, 674f Strand invasion, 845, 845f Streptomycin, 909, 909-910, 909t Stroma, 567 Strominger, Jack, 1038 Structural formulas, 21-22, 23f Structure-based drug development, 1042-1045, 1043f, 1044f Substitution(s) in amino acid sequences, 178-181, 179f-181f conservative, 178 production of, 156, 156f Substitution matrices, 178-181, 179f-181f Substrate(s), 220. See also Enzymesubstrate complex binding of, enzyme specificity and, 275-277 chromogenic, 256 Substrate binding. See also Enzymesubstrate complex concerted mechanism in. 294-296 cooperative, 200-201, 201f, 294 energy for, 229, 254, 275-277. See also Binding energy sequential model of, 294 specificity of, 220-221, 221f, 275-277 Substrate channeling in amino acid synthesis, 722, 722f in citric acid cycle, 512 in nucleotide synthesis, 737-738 Substrate cycle, 489 Substrate-level phosphorylation, 463–464, 507-508 Subtilisin, 186, 261, 261f active sites on, 186, 186f site-directed mutagenesis of, 262, 262f Subunit vaccines, 1002 Subunits, protein, 48, 48f Succinate dehydrogenase, in citric acid cycle, 509 - 510Succinate, in 4-hydroxyproline synthesis, 812f, 813 Succinate thiokinase, in citric acid cycle, 507-508 Succinate-Q reductase (Complex II), 531, 532f, 532t, 535-536 Succinvl CoA, 649 in amino acid degradation, 693 in citric acid cycle, 507-508, 508f, 514, 514f formation of, 649-652, 650f, 651f, 693 porphyrins from, 728-730, 729f Succinyl CoA synthetase, in citric acid cycle, 507-508 Sucrase, 327, 454

Sucrose, 327 plant synthesis of, 597, 598f Sugar(s). See also Carbohydrates amino, 330 disaccharide, 327-329. See also Disaccharides monosaccharide, 320-327. See also Monosaccharides in DNA, 110-111, 110f in RNA, 110f, 111 nonreducing, 325 reducing, 325 Sugars, phosphorylated, 326-327 Suicide inhibition, 242, 750, 754 Sulfate, conjugation of, 1034, 1035 Sulfation, 298t Sulfolipids, in thylakoid membrane, 567 Supercoiled DNA, 117, 117f, 826, 827f, 828f DNA condensation from, 828 linking number and, 826, 827f negative supercoiling in, 826, 829-831, 941 positive supercoiling in, 826 relaxation of, 828-829, 829f topoisomerases and, 828-831 twist in, 826-827, 827f writhe in, 826–828, 827f Supercompensation, 806 Supernatant, in centrifugation, 67, 68f Superoxide anion, 198, 198f Superoxide dismutase, 541-542, 542f Superoxide ion, 540, 542f Supersecondary structures, 47, 47f, 82–83, 83f, 174, 175f. See also Motifs Suppressors of cytokine signaling (SOCS), 797, 797f, 800 Surface complementarity, noncovalent bonds and, 11 Svedberg units (S), 76, 76t Syk (spleen tyrosine kinase), 989-990 Symmetry matching, 922 Symporter, 380-381, 380f, 381f Synapses, 389, 390f Synaptic cleft, 389, 390f Synonyms, 129, 129t Synthetic analog system, 269 T cell(s), 980–981 activation of, 995–996 cytotoxic (killer), 980, 994–995, 997–998 helper, 980, 996-998 in HIV infection, 999-1000 major histocompatibility complex and, 980-981 memory, 1002 negative selection of, 1000-1001, 1000f positive selection of, 1000, 1000f T state, of hemoglobin, 202-204, 203f, 207-208, 207f T1R proteins, as sweet receptors, 964–965 T2R proteins, as bitter receptors, 963–965 Tactile sensation, 973-974 Takepron (lansoprazole), 1046, 1046 Tamoxifen, 948, 948

INDEX

Tandem mass spectrometry, 93-94 TAP protein, 992 Taste anatomic structures in, 963, 963f tastants in, 962, 962-963 Taste buds, 963, 963f Taste papillae, 963 TATA box, 126f, 127, 866-867, 867f, 868f TATA-box-binding protein (TBP), 867-868, 867f, 868f, 950 repeated motifs in, 184, 185 TATA-box-binding protein associated factor. 950 Taurocholate, 642 synthesis of, 779-780, 780 Taxol, 1019–1020 Tay-Sachs disease, 765-766 TBP. See TATA-box-binding protein (TBP) T-cell receptors, 980-981, 994-998 CD3 and, 995, 995f CD4 and, 980 CD8 and, 994-995, 995f cytotoxic T cells and, 980, 994-995, 997-998 domains of, 994, 994f genes of, 994 helper T cells and, 980, 996-998 MHC complex proteins and. 994-998.994f number of, 980 structure of. 993f. 994, 994f in T-cell activation, 995-996, 995f T-cell selection, 1000-1001, 1000f T-DNA, 166 Telomerase, 124, 837 Telomerase inhibitors, 837 Telomeres, 836-837, 836f, 837f Temperature perception, 973 Template in replication, 121-122, 122f, 820-821 sequence, 184 in transcription, 124, 125f, 126, 126t, 854.855f in translation, 123-124 -10 sequence, 857, 857f Terminal transferase, 154 Tertiary protein structure, 45-47, 46f, 47f, 183-184, 183f. See also Protein(s), folding of Testosterone, 784 functions of, 784-785 reduction of, 785 synthesis of, 780, 780f, 784-785, 784f Tetracycline, 909t Tetrahydrobiopterin, 695, 695 Tetrahydrofolate, 715 in amino acid synthesis, 715–716, 716-717, 716f in deoxyribonucleotide synthesis, 748-749. 749 Tetrahymena, self-splicing in, 880f, 881 Tetrodotoxin, 384, 384

Tetroses, 320 TFII, in transcription, 867-868 TFPI (tissue factor pathway inhibitor), 312 Thalassemia, 210-211 Therapeutic index, 1036 Thermodynamics, 11–13. See also Energy of coupled reactions, 429-430, 431-433 enzymes and, 222-225 in metabolism, 429-435 Thermogenesis, oxidative phosphorylation in, 556-557 Thermogenin, 557 Thiamine (vitamin B_1), 441t biosynthesis of, 678, 679f deficiency of, 517 Thiamine pyrophosphate (TPP), 500 in citric acid cycle, 500, 500f, 501f deficiency of, 517 in pentose phosphate pathway, 605, 605f Thiamine pyrophosphate (TPP), 594 Thick filaments, in myosin, 1016, 1017f ThiF, 678, 679f Thin filaments, in myosin, 1016, 1017f Thioester intermediate, in glycolysis, 461-463, 462f Thiogalactoside transacetylase, 924 6-Thioguanine, 1047-1048, 1048 Thiolase, 647-648, 647t Thioredoxin, 598-599, 598t, 599 Thioredoxin reductase, 747 30S initiation complex, 902, 903f 30S subunit, of ribosome, 892, 897-898, 898f, 902, 903f -35 sequence, 126f, 127, 857, 857f ThiS, 678, 679f Thorazine (chlorpromazine), 1038 discovery of, 1038 mechanism of action of, 1038, 1039f 3' poly(A) tail, 871-872, 871f 3' splice sites, 842-843, 873f Threonine in glycoproteins, 330 pyruvate formation from, 691, 691f structure of. 30, 30f synthesis of, 724, 724f Threonine deaminase, 723-724, 723f Threonine kinases, 298-299, 299f, 299t Threonine operon, 932-933, 932f Threonyl-tRNA synthetase, active site of, 894, 894-895 Threonyl-tRNA synthetase complex, structure of, 896, 896 Thrombin, 221, 221f in blood clotting, 308–310, 308f, 312 γ-carboxylation of, 298t inhibitors of, 312 Thromboxanes, 664–665, 665, 665f Thylakoid membrane, 567-568, 567f, 574-575 proton gradient across, 577-581 stacked vs. unstacked regions of, 583, 583f Thylakoid spaces, 567, 567f Thylakoids, 567, 567f, 574-575

Thymidine, 111 Thymidine kinase, 740 Thymidine phosphorylase, 740 Thymidine triphosphate (TTP), 752 Thymidylate, 111, 748 synthesis of, 748, 748f salvage pathway for, 740 Thymidylate inhibitors, for cancer, 749-750 Thymidylate synthase, 748 Thymine, 4, 4, 111, 111, 748 in DNA repair, 841 synthesis of, 716 Thymocytes, 1000-1001, 1000f Thymus, T-cell selection in, 1000-1001, 1000f Thyroxine, synthesis of, 726, 726f Ti plasmids, 166-167, 167f Time of flight (TOF) mass analyzer, 92, 92f Time, sense of, 974 Tip links, 972, 972f Tissue factor, in blood clotting, 308 Tissue factor pathway inhibitor (TFPI), 312 Tissue-type plasminogen activator (TPA), 312-313, 313f Titin, 34 Titration, 15-16 Tobacco mosaic virus, 122 α-Tocopherol (vitamin E), 442, 442, 442t in lipid peroxidation prevention, 542 Toll receptors, 978-979 Toll-like receptors, 978–979, 978f, 979f Tonegawa, Susumu, 987 Tongue, taste buds of, 963, 963f Topoisomerases, 828-831 bacterial, 831 type I, 828-829 structure of, 828, 828-829 in supercoiled DNA relaxation, 828-829, 829f type II, 829-831, 830f structure of, 829-831, 830 Topoisomers, 826-828, 828f Torpedo marmorata, acetylcholine receptor in. 389. 390f Torr. 200b Torsion angles, 37, 37f, 38f Tosyl-L-phenylalanine chloromethyl ketone (TPCK), 241, 241-242 Touch, 973-974 Toxoid vaccines, 1002 TPA (tissue-type plasminogen activator), 312-313, 313f TPI. See Triose phosphate isomerase (TPI) TPP. See Thiamine pyrophosphate (TPP) TΨC loop, of tRNA, 890f, 891 Trace elements, 20 Trans configuration, of peptide bonds, 36-37, 36f, 37f Transacetylase component, of pyruvate dehydrogenase complex, 502, 502f Transaldolase, in pentose phosphate pathway-glycolysis linkage, 601-605, 605,606f

C 4 5

Index

Transaminases in amino acid degradation, 680 evolution of, 713 pyridoxal phosphate-dependent, 712-713 Transamination in amino acid degradation, 680-685 in amino acid synthesis, 709-710, 712-713, 712f pyridoxal phosphate enzymes in, 680-685 reaction mechanism of, 682f, 712f Schiff-base intermediates in, 681-682 stereochemistry of proton addition in, 713, 713f Transcription, 124-127, 125f, 851-869 in archaea, 869 in bacteria, 852-863 attenuation in, 931-932, 931f, 932f chain elongation in, 855-856, 855f DNA rewinding in, 859, 859f helix unwinding in, 858, 858f initiation of, 854-858 promoters in, 856-858 proofreading in, 856 regulation of, 922-928. See also Gene expression/regulation rho in, 860-861, 861f ribonucleases in, 863 riboswitches in, 860, 860f RNA processing and, 133, 863. See also RNA processing RNA-DNA hybrid in backtracking of, 856, 856f separation of, 855, 856f, 859f translocation of, 855, 855f of rRNA, 863 termination of, 859-860, 859f transcript processing in, 863 transcription bubble in, 858-859, 859f transition from open to closed promoter complex in, 858 translation and, 864, 864f of tRNA, 863 vs. in eukaryotes, 864, 864f coordination with translation, 864, 864f definition of, 851 direction of, 125, 854-855, 855f DNA template in, 124, 125f, 126, 126f, 854.855f enhancer sequences in, 127 in eukaryotes, 864 carboxyl-terminal domain in, 868, 877, 877f chain elongation in, 867-868 coactivators in, 947-948, 948f, 949-951,950f enhancers in, 127, 868-869, 943-944, 943f histone acetylation in, 949-951, 950f initiation of. 866-869, 950 nuclear membrane in, 864 polyadenylation in, 871, 871f pre-mRNA processing and, 870-872, 871f, 878-879. See also Splicing

promoters in, 866-869, 866f regulation of, 864, 941-944. See also Gene expression/regulation repression of, 950-951 RNA editing in, 872-873, 873f RNA polymerases in, 865-867, 866f, 869-872. See also RNA polymerase(s) RNA processing and, 133, 864, 869-879. See also RNA processing splicing in, 132f, 133, 851-852, 873-879. See also Splicing TATA box in, 866-867, 867f, 868f transcript processing in, 869-879 transcription factors in, 867-868, 867f. See also Transcription factors vs. in bacteria, 864, 864f evolution of, 869 fundamental reaction of, 853 in humans, pre-mRNA processing and, 878-879 inhibition of, by DNA-binding proteins, 924-927 initiation of, 126-127, 126f de novo, 854 RNA chain elongation in, 858-859, 859f RNA chain formation in, 858-859 intron removal in, 132-133, 132f overview of. 851-852 promoters/promoter sites in, 126-127, 126f RNA modification in, 127, 127f RNA polymerase in, 124-127, 125f, 852, 853-858. See also RNA polymerase splicing in, 132-133, 132f, 851-852, 873-879. See also Splicing stages of, 852-853 termination of, 127 Transcription bubble, 853-854, 854f, 858-859, 859f Transcription factors in eukaryotes, 867, 941-944, 943. See also DNA-binding proteins and specific factors chromatin structure and, 946 definition of, 164, 941 DNA-binding domains of, 941, 942-943 in gene regulation, 942-943 nuclear hormone receptor, 946-948, 947f regulatory domains of, 942-943 in gene knockout, 164–165 in prokaryotes, 941, 943 Transcriptome, 162 Transducin P-loop NTPase domains in, 284, 284f rhodopsin and, 968 Transesterification, in splicing, 874-875 Transfer RNA. See tRNA Transferase(s), 249t. See also Enzyme(s) in glycogen metabolism, 620, 620f Transferrin, 730, 951 Transferrin receptor, 951

Transgenic animals, 164, 164f Transglutaminase, 309 Transition state catalytic stabilization of, 225-226, 225f, 227, 243-244 enzymes and, 225-226, 225f, 227 Transition-state analogs, 243, 243f Transketolase, 594 in Calvin cycle, 594, 595f in pentose phosphate pathway-glycolysis linkage, 601-605, 605f Translation, 130, 130f. See also Gene expression/regulation; Protein(s), synthesis of accuracy of, 888-889, 889t, 895 activating enzymes in, 893 adenvlation in, 893-895 in α helix. 39 aminoacyl-tRNA synthetases in, 888, 893-897 activation sites of, 894, 894-895 amino acid activation by, 893-895 classes of, 897, 897t editing sites of, 895, 895 proofreading by, 895 specificity of, 893-894 tRNA recognition by, 896 anticodons in, 889, 891-893, 892f attenuation and, 931-932 in bacteria transcription and, 864, 864f vs. in eukaryotes, 907-908 base pairing in, 891-893, 901 codon-anticodon interactions in, 891-893 wobble in, 891-893 chain elongation in, 902-904, 903f, 904f codons in, 889, 891-893, 892f coordination with transcription, 864, 864f definition of, 123, 887 direction of, 888, 905-906 elongation factors in, 902-903, 903f in eukarvotes, 908 error rate in, 888-889, 889t, 895 in eukarvotes transcription and, 864, 864f vs. in bacteria, 907-908 fidelity of, 888-889, 889t, 895 formylmethionyl-tRNA in, 901-902, 902, 902f free energy of, 893-894 in gene regulation, 951–953 inhibition of by antibiotics, 909-910 by diphtheria toxin, 910-911 by ricin, 911 initiation of, 130, 893-894, 900-902.903f in eukaryotes, 907–908, 907f, 908f sites of, 900-901, 901f in iron metabolism, 951-953 mechanisms of, 905f

Translation—(Continued) mRNA in, 123-124, 130, 130f. See also mRNA (messenger RNA) in initiation, 900-901 in termination, 901 translocation of, 904-906, 905f overview of, 887-888 peptide bond formation in, 903-904, 904f peptidyl tRNA in, 903-904 proofreading in, 895 protein sorting/transport in, 911-914 release factors in, 906, 907f ribosome in, 897–906 initiation sites in, 902, 903f RNA editing in, 872-873, 873f rRNA in, 892, 898-900, 901, 903-904 Shine-Dalgarno sequences in, 130, 901 signaling in, 911-914 termination of, 130, 906, 907f in eukaryotes, 908 translocation in, 904-906 tRNA in, 127-128, 889-893, 901-906. See also tRNA Translocase, 904-906, 905f Translocon, 911–912 Transmissible spongiform encephalopathies, 55 - 56Transpeptidase, penicillin inhibition of. 245.245f Transplant rejection, 998 Transport vesicles, 913-914, 914f Transporters, 372 ABC, 378-380, 378f, 379f antiporters, 380, 380f ATP-ADP translocase, 552-553, 553f, 554f dicarboxylate, 554, 554f glucose, 372, 414, 477, 477t glucose 6-phosphate, 635 inhibitors of, 558, 558f lactose permease, 381, 381f in metabolism, 438-442, 438f-440f in oxidative phosphorylation, 438-442, 438f-440f, 439f, 531-543, 532t. 551-553. 551f-553f phosphate, 554, 554f in photosynthesis, 568-572, 569f, 596f pyruvate, 554, 554f secondary, 381, 381f symporters, 380-381, 380f, 381f tricarboxylate, 554, 554f uniporters, 380, 380f Tree of life, 3f Triacylglycerol(s), 639, 641-642 energy storage in, 641-642 hydrolysis of, 643-644, 643f. See also Fatty acid metabolism mobilization of, 643-644, 643f synthesis of, 760-767, 760f, 761 hepatic, 773 transport of, 642, 642f, 773-774 Triacylglycerol synthetase complex, 761 Tricarboxylate carrier, 554, 554f

Tricarboxylic acid cycle. See Citric acid cycle Triglycerides. See Triacylglycerol(s) Trimethoprim, 750 Trimethoprim-sulfamethoxazole, 750 Trinucleotide repeats, 842 Triose kinase, 470 Triose phosphate isomerase (TPI), 242, 242, 459 in glycolysis, 456f, 458-460 Triose phosphates, in sucrose synthesis, 597. 598 Trioses, 320, 320f Triple helix, of collagen, 44-45, 44f tRNA (transfer RNA), 123t, 124, 127-128, 128f. See also RNA anticodons of, 889, 890f, 891-893. 891f. 896 base-pairing in, 890–893, 890f wobble in, 891-893 charged, 891f, 893 codons of, 889 functions of, 124 initiator, 907 inosine in. 892 key features of, 889-890 post-transcriptional processing of, 863 precursor of, processing of, 870, 871f ribosomal binding of mechanism of, 903-904, 905f sites for, 900, 902-904, 905 sequencing of, 889, 890f structure of, 128f, 889-890, 890f-892f transcription and processing of, 863, 870, 871f in translation, 889–893, 901–906 as adaptor molecule, 889 binding sites for, 900, 901f in chain elongation, 903-904, 904f, 905f function of, 889 in initiation, 901–902 translocation of, 904-906, 905f Trojan horse strategy, 245 Trombone model, 833, 833f Tropical plants, photosynthesis in, 599-600 Tropomyosin, 1017 Troponin complex, 1016 TRP channels, 973 trp operon, 931 Trypsin, 221, 221f, 305-306 catalytic triad in, 260-261, 261f chymotrypsin and, 260-261, 261f in protein cleavage, 80, 82f, 306 structure of, 260, 261f in zymogen activation, 306, 306f Trypsin inhibitor, 306, 307f Trypsinogen, 306 Tryptophan, 696 degradation of, 696, 696f nicotinamide from, 726f, 727 pyruvate formation from, 691 serotonin from, 726f, 727

structure of, 28, 29f synthesis of, 720-722, 721f, 722f Tryptophan operon, 931 Tryptophan synthase, 722-723, 723 Ts elongation factor, 903 t-SNARE, 366, 914 TTP (thymidine triphosphate), 752 Tu elongation factor, 902-903, 903f Tuberculosis, proteasome inhibitors for, 679 Tubulin in microtubules, 1018-1019, 1019f sequence homology of, 1019 Tumor-inducing (Ti) plasmids, 166-167 Tumor-suppressor genes, 420, 842 Turnover number (k_{cat}), 234, 234t 20S proteasome, 677, 677f -25 region, 126f, 127 26S proteasome, 677, 677f Twist, in DNA, 826-827, 827f Two-dimensional electrophoresis, 74, 74f Typing-monkey analogy, 53, 53f Tyrosine degradation of, 695-696 epinephrine from, 726, 726f melanin from, 726 structure of, 30, 30f, 696 synthesis of, 719-720, 721f thyroxine from, 726, 726f Tyrosine kinases, 298–299, 299f, 412 in cancer, 420 receptor, 412 Tyrosine phosphatase IB, 799 Tvrosinemia, 698t Tyrosyl radical, 745-747 U1 snRNA, 875-876, 875f, 876t U2 snRNA, 875f, 876, 876t U4 snRNA, 875f, 876, 876t U5 snRNA, 875f, 876, 876t U6 snRNA, 875f, 876, 876t Ubiquinol, 535 Ubiquinone (coenzyme Q), 532, 533 in fatty acid metabolism, 646 Ubiquitin. 675-678 structure of, 675, 675–676 Ubiquitin-activating enzyme (E1), 675-676, 675f Ubiquitination, 298t, 675-678 Ubiquitin-conjugating enzyme (E2), 675-676, 675f Ubiquitin-protein ligase (E3), 675-676, 675f UCP-1, 557 UCP-2, 557 UCP-3, 557 UDP (uridine diphosphate), hydrolysis of. 431 UDP-α-glucuronic acid, 1035 UDP-galactose, 470, 471 UDP-glucose, 470, 470-471, 627 in glucose synthesis, 627-630 in glycogen synthesis, 627-630 structure of, 627, 627 in sucrose synthesis, 597, 598

Index

UDP-glucose phosphorylase, 627 Umami, 962, 962f, 965 UMP (uridine monophosphate), synthesis of. 739 Uncompetitive inhibition, 238, 238f, 239, 239f, 240, 240f Uncoupling proteins, 557, 558f Unfolded protein response, 802 Uniporters, 380, 380f Units of measure for distance, 7 for energy, 7 Unsaturated fatty acids, 648-649, 664-665. See also Fatty acid(s) Upstream promoter elements in bacteria, 857 in eukarvotes, 866 Uracil, 111, 111 in DNA repair, 841 thymidylate from, 748 Uracil DNA glycolase, 841, 841f Urate degradation of, 752, 753f in gout, 753-754 Urea, 49 Urea cycle, 673, 685-690 carbamoyl phosphate formation in, 685-687 disorders of, 688-689 evolution of, 688 gluconeogenesis and, 687 reactions in, 685f stoichiometry of, 687 Ureotelic organisms, 685, 690 Uric acid degradation of, 752, 753f in gout, 753-754 Uricotelic organisms, 690 Uridine, 111 Uridine diphosphate. See UDP (uridine diphosphate) Uridine monophosphate (UMP), synthesis of, 739 Uridine triphosphate. See UTP (uridine triphosphate) Uridylate, 739, 739, 863, 863 Urine, as bleach, 687b Uroporphyrinogen III, 729, 729 UTP (uridine triphosphate) in glycogen synthesis, 617 synthesis of, 739 V genes, 987-988 in antibody switching, 990-991 Vaccine(s), 1002-1003 HIV, 1003 synthetic, 157 Vaccinia virus vectors, 163-164 Valine degradation of, 694-695 in maple syrup urine disease, 697 structure of, 28, 29f

synthesis of, 723-724

See also Bonds in antigen-antibody binding, 984, 985, 985f. 986 in double helix, 10, 10f in enzyme-substrate complex, 228, 228f in space-filling models, 61, 61f Vanishing white matter, 908–909, 909f Variable (V) genes, 987-988 in class switching, 990-991 Variable number of tandem repeats (VNTR) region, 333, 333f Variable regions, 983, 984f Vasopressin, 96, 96 Vectors cloning, 148, 149 expression, 149, 154-155 lambda phage, 150-151, 150f, 151f plasmid, 149-151, 149f, 150f viral, 150-151, 150f, 151f, 163-164 Velcade (bortezomib), 678, 679 Very-low-density lipoproteins, 773, 773t. See also Lipoprotein(s) Vesicles formation of, 365 transport, 913-914, 914f Viagra (sildenafil), 1039 discovery of, 1039 Vibrio cholerae ABC transporter of, 378, 378f, 379f infection with, 421-422 Vibrio fischeri, quorum sensing in, 930-931, 930f Vioxx (rofecoxib), 1044, 1044-1045, 1049 Viral infections, carbohydrates in, 339-340 Viral vectors, 150-151, 150f, 151f, 163-164 Viruses cell entry by, 340f, 365-366 HIV, 999-1000, 999f infectious mechanisms of, 339-340, 340f, 365-366 influenza, 339, 340f protein structure of, 48, 48f retroviruses, 999-1000 RNA, 122–123, 123f Vision, 966-971 color, 966, 969-971 in animals, 970, 970f evolution of, 970, 970f cones in, 966, 969-970 evolution of, 970, 970f photoreceptors in, 966 rhodopsin in, 966-968 rods in, 966-967, 966f signal transduction in, 968-969, 968f Visual pigments, 969–970, 970f. See also Color vision in cones, 969-970 in rods, 966-967 Vitamin(s), 20 B, 441, 441–442, 441t coenzyme, 441, 441-442, 441t deficiencies of, 441t, 442

Van der Waals interactions, 8, 8f, 11, 46.

electron carrier derived from, 441-442 electron carriers derived from, 440t evolution of, 441-442, 441t noncoenzyme, 442, 442, 442t Vitamin A (retinol), 442, 442, 442t deficiency of, alcohol-related, 812 Vitamin B₁ (thiamine), 441t biosynthesis of, 678, 679f deficiency of, 517 Vitamin B₂, 441t Vitamin B₆ (pantothenic acid), **441**, 441t Vitamin B₁₂ (cobalamin), 441t, 649 in amino acid synthesis, 650-652, 650f, 651f. 717 as coenzyme, 650-652, 650f, 651f, 717 in fatty acid metabolism, 649-652 structure of, 650, 650-651 Vitamin C (ascorbic acid), 442, 442t, 813.813 deficiency of, alcohol-related, 812-813 forms of, 813, 813 Vitamin D (calciferol), 442, 442t deficiency of, 786 sources of, 786 synthesis of, 785, 785f Vitamin E (a-tocopherol), 442, 442, 442t in lipid peroxidation prevention, 542 Vitamin K, 442, 442t in blood clotting, 310-311 deficiency of, 57, 301 structure of, 310 VJ recombination, 987-988, 988f V(D)J recombination, 988, 988f V_{max}, 232–234 calculation of, 233 turnover number and, 234, 234t VNTR region, 333, 333f VO4³⁻, 1011 Voltage-gated ion channels, 387-388, 388f. See also Ion channels Von Gierke disease, 634-635, 634t VR1 (capsaicin receptor), 973-974, 973f v-SNARE, 366, 914 v-Src, in cancer, 420 Wang, James, 828 Warburg effect, 478 Warburg, Otto, 454, 478 Warfarin, 310, 310 Water dissociation of, equilibrium constant for. 14 equilibrium constant for, 14 hydrogen bonds of, 9–10, 9f hydroxide ions in, 13-17 molecular models of, 23f polarity of, 8-9 properties of, 8-9 Watson, James, 5, 113 Watson-Crick base pairs. See Bases/base pairs

INDEX

Watson–Crick DNA model, 5, 113–115, 114f. See also Bases/base pairs; DNA; Double helix
Weight, regulation of, 791–798. See also Caloric homeostasis
Wernicke–Korsakoff syndrome, 812
Western blotting, 88f, 89, 142
White adipose tissue, 556
Whooping cough, 422
Wiley, Don, 992
Wilkins, Maurice, 113
Withering, William, 377
Wobble hypothesis, 891–893
Writhe, in DNA, 826–828, 827f
Wyman, Jeffries, 202

Xanthine, degradation of, 752, 753f Xanthine oxidase inhibitors, 754 Xanthomas, 776 Xanthylate, **743**, 744 Xenobiotic compounds, 1034
Xeroderma pigmentosum, 842
X-ray crystallography, 98–100, 98f–100f of enzyme–substrate complexes, 227, 227f
X-ray diffraction photographs, of DNA, 113, 113f
Xylulose 5-phosphate, 596 in Calvin cycle, 596, 596 in pentose phosphate pathway, 602f, 604

Yamanaka, Shinya, 944 Yanofsky, Charles, 931 Yeast artificial chromosomes, 151 Yeast, genome of, 158, 937, 938f

Z line, 1016, 1016f Z scheme of photosynthesis, 577, 577f Zantac (ranitidine), 1045, 1045 ZAP-70, 995-996 Z-DNA, 116, 116f, 116t Zellwegger syndrome, 652 Zinc finger, 109f Zinc proteases, 263, 264, 264f Zinc sites, in carbonic anhydrase, 267-269, 267f-269f evolution of, 271 Zinc-based domains, 946 Zingiberene, 958 Zonal centrifugation, 77–78,77f Zwitterions, 27-28, 27f Zymogens, 303-313, 674 in blood clotting, 307-312 functions of, 303 gastric, 303, 303t, 674 pancreatic, 303-307, 303t, 304f, 674

ACIDITY CONSTANTS

pK_a values of some acids

Acid	pK' (at 25°C)	Acid	р <i>К</i> ′ (at 25°С)
Acetic acid	4.76	Malic acid, pK_1	3.40
Acetoacetic acid	3.58	pK_2	5.11
Ammonium ion	9.25	Phenol	9.89
Ascorbic acid, p K_1 p K_2	4.10 11.79	Phosphoric acid, pK ₁ pK ₂	2.12 7.21
Benzoic acid	4.20	pK_3	12.07
n-Butyric acid	4.81	Pyridinium ion	5.25
Cacodylic acid	6.19	Pyrophosphoric acid, pK_1	0.85
Citric acid, p K_1 p K_2	3.14 4.77	pK_2 pK_3 pK_4	1.49 5.77 8.22
pK_3	6.39	Succinic acid, pK_1	4.21
Ethylammonium ion	10.81	nK ₂	5.64
Formic acid	3.75	Trimethylammonium ion	9.79
Glycine, p K_1 p K_2	2.35 9.78	Tris (hydroxymethyl) aminomethane	8.08
Imidazolium ion	6.95	Water*	15.74
Lactic acid	3.86		
Maleic acid, p K_1 p K_2	1.83 6.07		

 $*[H^+][OH^-] = 10^{-14}; [H_2O] = 55.5 \text{ M}.$

Typical pK_a values of ionizable groups in proteins

Group	Acid		Base	Typical pK_a	Group	Acid	\rightarrow	Base	Typical pK_a
Terminal α-carboxyl group	° ⊂_o_H		° C C	3.1	Cysteine	—s´ ^H	`	—S-	8.3
Aspartic acid	0		O 		Tyrosine		<u> </u>		10.4
Glutamic acid	_ ^с _ ₀ _н		_ د ا	4.1	Lysine	+ H —N	`	—N	10.0
	H				Lysine	H	、	H	10.0
Histidine		`		6.0	Arginine	H + N~H N==-C	<u> </u>	H N—C	12.5
Terminal α-amino	+ H —N	`	—N.	8.0		,Ñ−H H		,Ň−H H	
group	H	<u>`</u>	V _н						

Note: pK_a values depend on temperature, ionic strength, and the microenvironment of the ionizable group.

STANDARD BOND LENGTHS

Bond	Structure	Length (Å)	
С—Н	R ₂ CH ₂ Aromatic RCH ₃	1.07 1.08 1.10	
С—С	Hydrocarbon Aromatic	1.54 1.40	
C=C	Ethylene	1.33	
C≡C	Acetylene	1.20	
C—N	RNH_2 O=C-N	1.47 1.34	
С—О	Alcohol Ester	1.43 1.36	
C=0	Aldehyde Amide	1.22 1.24	
C—S	R_2S	1.82	
N—H	Amide	0.99	
O—H	Alcohol	0.97	
0—0	O ₂	1.21	
Р—О	Ester	1.56	
S—H	Thiol	1.33	
S—S	Disulfide	2.05	