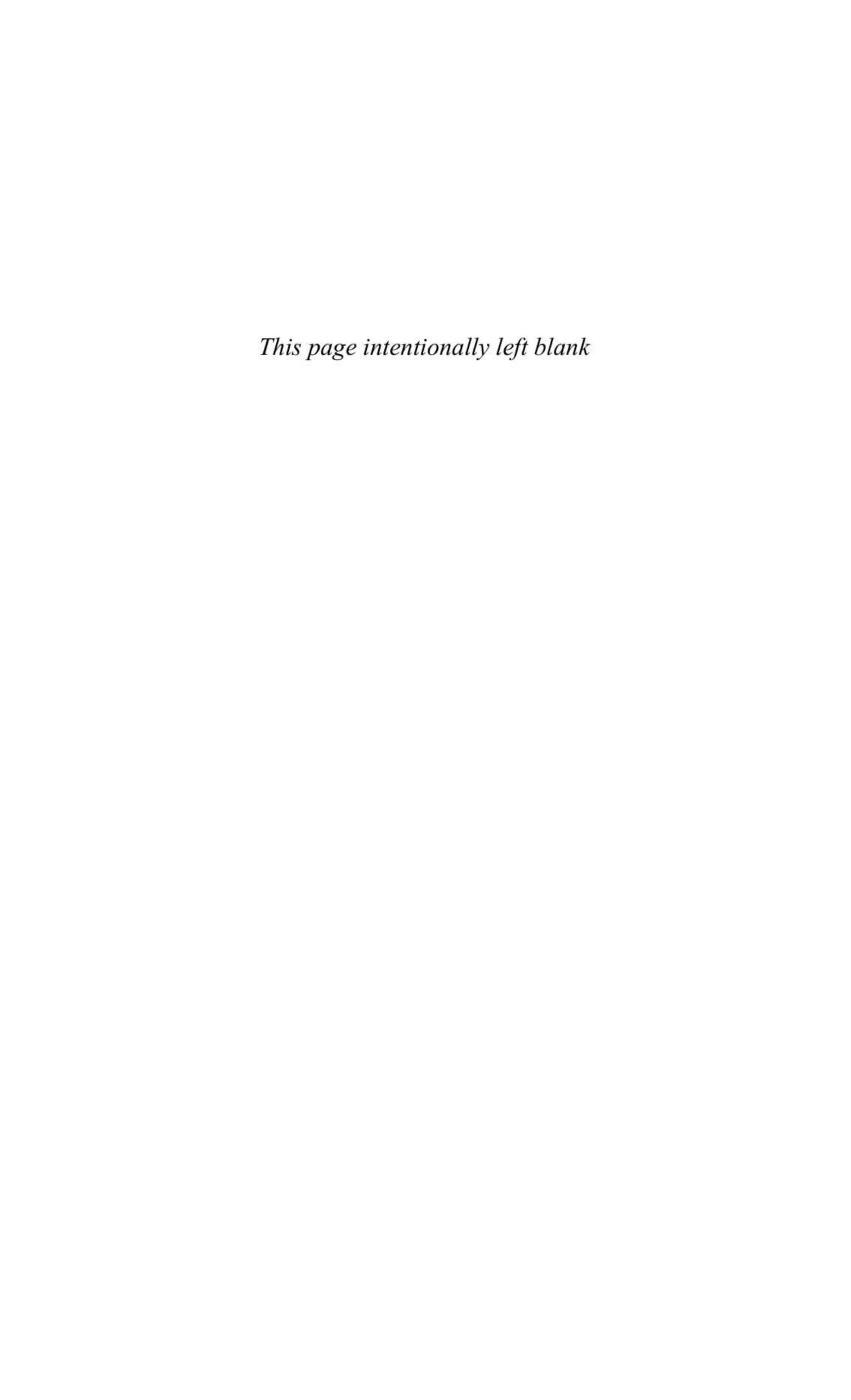
MODERNORS CORGANIC
SYNTHESIS
IN THE

A COLLECTION OF STANDARD EXPERIMENTAL PROCEDURES

JIE JACK LI CHRIS LIMBERAKIS DEREK A. PFLUM

# Modern Organic Synthesis in the Laboratory



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A Collection of Standard Experimental Procedures

Jie Jack Li
Chris Limberakis
Derek A. Pflum

Pfizer Global Research & Development



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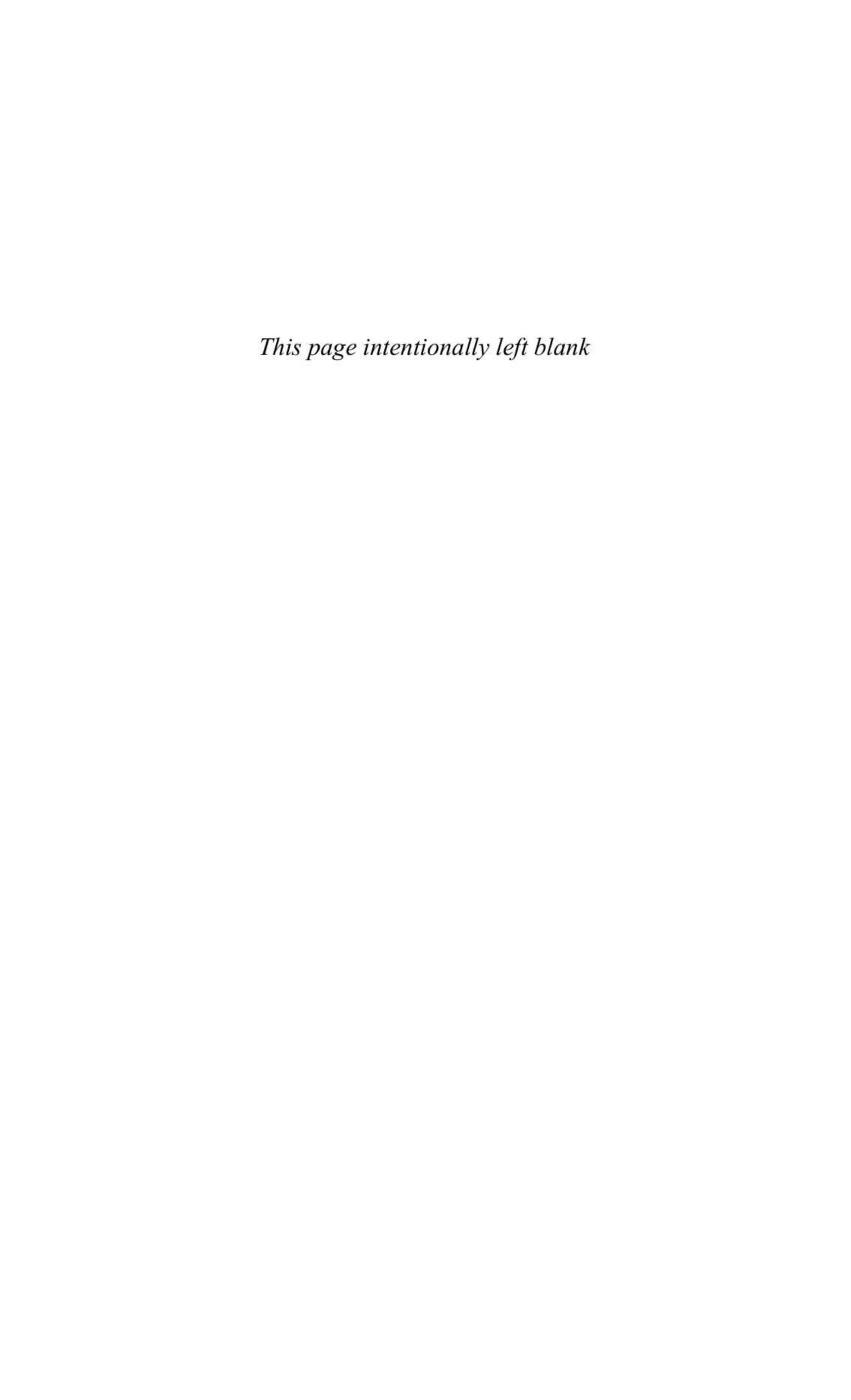
The material in this text is intended to provide general guidelines and is for informational purposes only. Although the materials, safety information, and procedures described in this book are designed to offer current, accurate and authoritative information with respect to the subject matter covered, the information and these procedures should serve only as a starting point for laboratory practices. They do not purport to specify minimal legal standards or to represent the policy of the authors or Oxford University Press.

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# To Sherry, Rachele, and Mary Kay For Their Love, Support, and Patience



## Foreword

Those who do not want to imitate anything, produce nothing.
—Salvador Dali

There exists a close analogy between the trials and tribulations of a chemist and those of an artisan. Just as an artist must balance the sensitive interplay of color, light, and brushstroke to create a timeless painting, so too must the chemist carefully marshal the reagents, solvents, and reactions that occupy chemical space to successfully forge a new molecular entity. Finding reliable, standardized procedures for those routine transformations is often a time-consuming and tedious process, one that subtracts from effort that could be devoted towards more creative and exciting pursuits. Indeed, much of the excitement, fascination, and allure of synthetic organic chemistry resides in the imaginative exploration of new realms in chemical reactivity and complexity, not the search for standard operation procedures.

In *Modern Organic Synthesis in the Laboratory*, Drs. Jie Jack Li, Chris Limberakis, and Derek Pflum provide procedures for the common transformations and protocols that permeate the tapestry of synthetic organic chemistry. From guidelines of proper flash chromatography technique, to procedures for drying solvents, and recipes for staining solutions, all of the basic, yet essential, organic laboratory techniques are described in detail. Nearly all types of common reactions for oxidations, reductions, functional group manipulations, and C–C bond formation are covered. With little question, this book is beautifully suited for practitioners of the field at all levels of study because of the thorough selection of procedures and a layout that is both well designed and comprehensive.

#### viii FOREWORD

We congratulate the authors for filling such an important gap in the chemical literature by carefully designing what will undoubtedly become a survival manual destined for untold numbers of organic chemists.

K. C. Nicolaou and Phil S. Baran The Scripps Research Institute July 12, 2006

## Preface

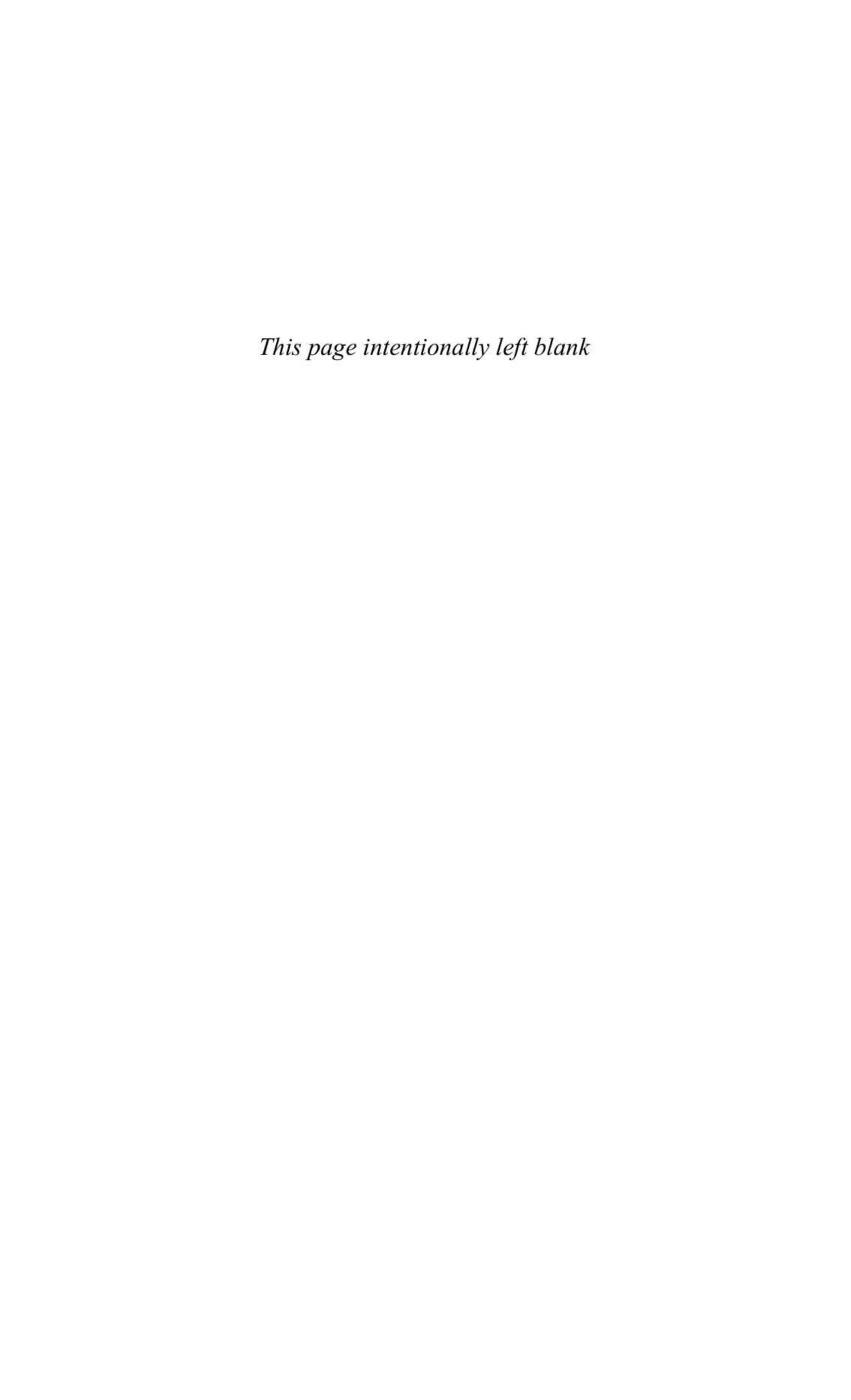
The prevalence of computer databases has, at the present time, made literature searching in organic synthesis an increasingly easier enterprise. However, the dilemma has now become how to select one procedure in an ocean of choices. For novices in the laboratory, in particular, this is a daunting task, deciding what reaction conditions to try first in order to have the best chance of success. This collection intends to serve as an "older and wiser" lab-mate by sharing the authors' own experience through this compilation of the most commonly used experimental procedures from established groups and/or rigorously reviewed journals.

Under the title of each experimental procedure, brief commentaries are often offered which summarize the authors' personal experience, and in many instances review articles are cited. These experimental procedures and commentaries are by no means "carved in stone", as you and your lab-mates may have your own favorite protocol. For the final products, detailed spectral data are not given because they simply take up too much space.

We are indebted to Professor John P. Wolfe at the University of Michigan, and his students, Joshua Ney and Josephine Nakhla; Professor Phil S. Baran at the Scripps Research Institute, and his students, Noah Z. Burns, Mike DeMartino, Tom Maimone, Dan O'Malley, Jeremy Richter, and Ryan Shenvi for proofreading the manuscript.

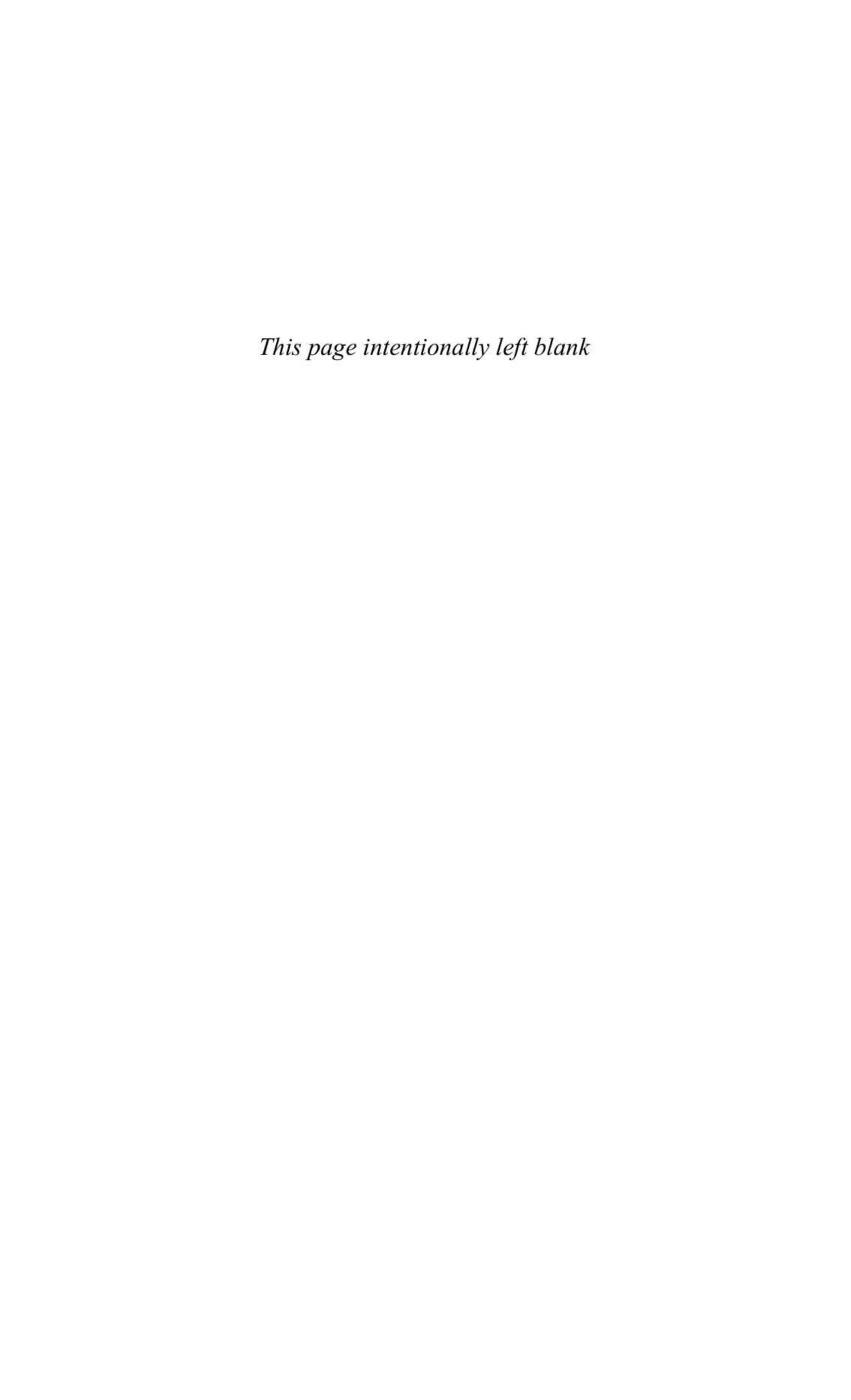
We had a good time putting together these experimental procedures. We have even been using the manuscript ourselves quite often. Hopefully you will find it as useful. We welcome your critique!

> Jack Li, Chris Limberakis, and Derek Pflum Ann Arbor, Michigan September 4, 2006



## Acknowledgments

The authors would like to extend their sincerest gratitude to the original publishers of the experimental procedures for granting permission to reprint these materials that form the core of this book. The publishers include: The American Chemical Society (The Journal of the American Chemical Society, Journal of Medicinal Chemistry, Journal of Organic Chemistry, Organic Letters, Organic Process Research and Development, and Organometallics), Elsevier (Biorganic Medicinal Chemistry, Journal of Organometallic Chemistry, Tetrahedron, and Tetrahedron Asymmetry), John Wiley & Sons (Organic Syntheses), Royal Society of Chemistry (Journal of the Chemical Society Perkin Transactions 1), Wiley-VCH Verlag GmbH & Co KG (Chemistry-A European Journal), Taylor & Francis Group (Synthetic Communications), NRC Research Press (Canadian Journal of Chemistry), Thieme (Synlett), and The Japan Institute of Heterocyclic Chemistry (Heterocycles). In addition, we would like to express much appreciation to the following companies for allowing us to use their product images in Chapter 1: Innovative Technology (Pure-Solv 400 Solvent Purification System), Analogix (Flash 12/40 system<sup>TM</sup>), Teledyne Isco (CombiFlash<sup>®</sup>Companion<sup>®</sup>), and Biotage (Flash 400<sup>TM</sup>).



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# Abbreviations and Acronyms

Ac acac	polymer support acetyl acetylacetonate	BINALH	Lithium 2,2´-dihydroxy- 1,1´-binaphthylethoxy- aluminum hydride
AcOH	acetic acid	BINAP	2,2-bis(diphenylphosphino)-
AE	asymmetric		1,1'-binaphthyl
	epoxidation reaction	BINOL	1,1'-bi-2-naphthol
AIBN	2,2'-azobisisobutyro-	BMS	borane dimethyl
	nitrile		sulfide complex
Alpine-borane®	B-isopinocam-	Bn	benzyl
	phenyl-9-borabi-	Boc	tert-butyloxycarbonyl
	cyclo[3.3.1]-nonane	BOM	benzyloxymethyl
AME	acetyl malonic ester	BOP	benzotriazol-1-yloxy-
AMNT	aminomalononitrile		tris(dimethylamino)-
	<i>p</i> -toluenesulfonate		phosphonium hexafluo-
Ar	aryl		rophosphate
B:	generic base	BPO	benzoyl peroxide
9-BBN	9-borabicyclo-	Bu	butyl
	[3.3.1]nonane	t-Bu	tert-butyl
BFO	benzofurazan oxide	Bz	benzoyl
TBHP	tert-butyl hydrogen	°C	degree Celsius
	peroxide	CAN	ceric ammonium nitrate
BHT	butylated hydroxy		(ammonium cerium(IV)
	toluene		nitrate)

#### xvi ABBREVIATIONS AND ACRONYMS

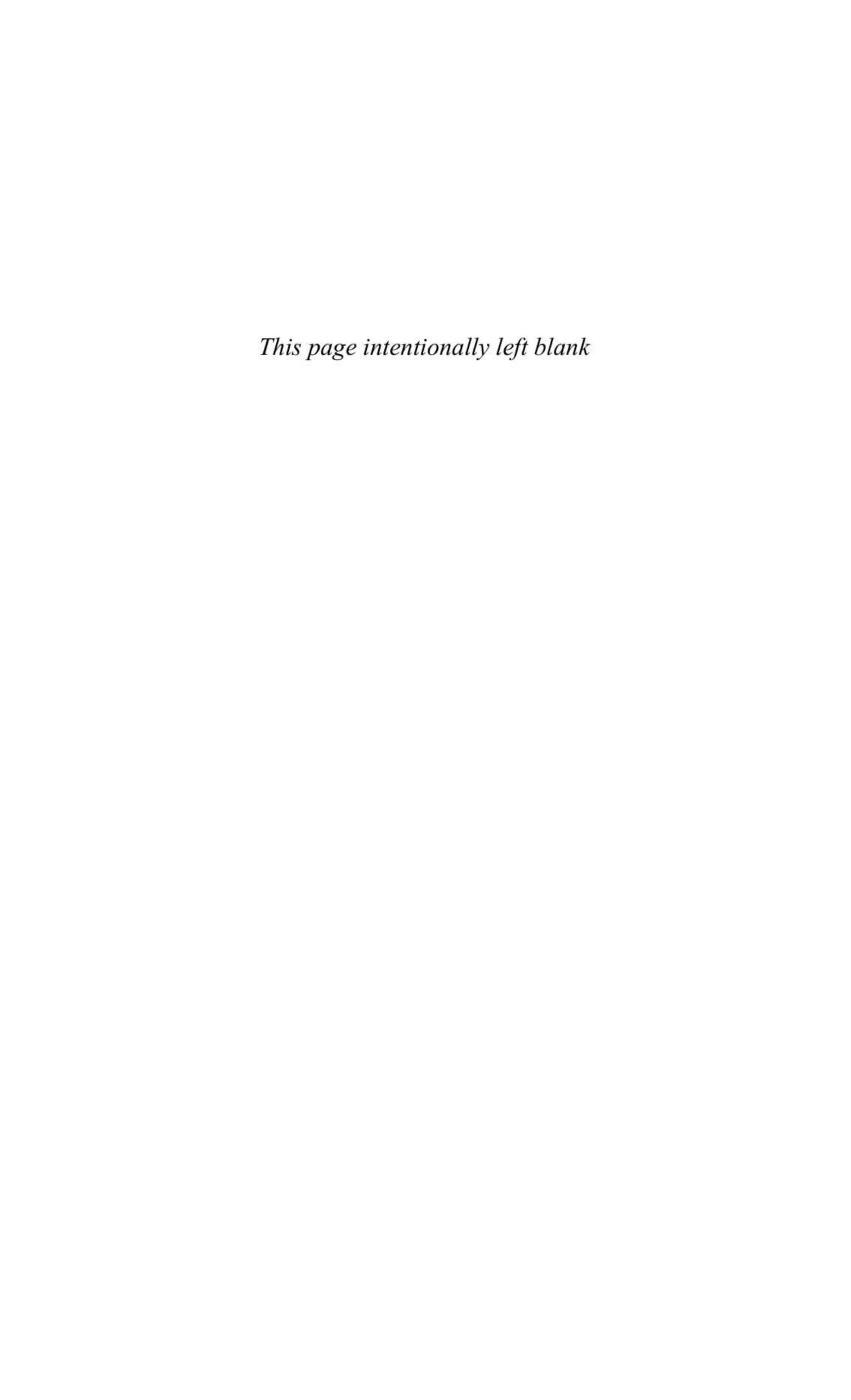
Chirald®	(2 <i>S</i> ,3 <i>R</i> )-(+)-4- dimethylamino-1,2- diphenyl-3-methyl-2-	(DHQD) <sub>2</sub> -PHAL	1,4-bis(9- <i>O</i> -dihydro-quinidine)-phtha-lazine
CTAB	butanol cetyl trimethylammo-	DIAD	diisopropyl azodicarboxylate
CBS	nium bromide Corey-Bakshi-	DIBAL-H	diisobutylaluminum hydride
Cbz	Shibata benzyloxycarbonyl	DIC	diisopropylcarbo- diimide
cp CSA	cyclopentadienyl camphorsulfonic acid	Diglyme	diethylene glycol dimethyl ether
CuTC	copper thiophene-2- carboxylate	Dimsyl	methylsulfinyl- methide
cy DABCO	cyclohexyl 1,4-diazabicyclo-	DIPEA	diisopropylethyl- amine
DAPA	[2.2.2]octane	DMAc	N,N-dimethyl- acetamide
	dipotassium azodicarboxylate	DMA	N,N-dimethylaniline
DAST	(diethylamino)sulfur trifluoride	DMAP	N,N-dimethyl- aminopyridine
dba DBE	dibenzylideneacetone 1,2-dibromoethane	DMDO DME	dimethyldioxirane 1,2-
DBU	1,8-diazabicyclo- [5.4.0]undec-7-ene	DMF	dimethoxyethane dimethylformamide
DBN	1,5-diazabicyclo-	DMFDMA	dimethylamino- formaldehyde
DCB	[4.3.0]non-5-ene dichlorobenzene		dimethyl acetal
DCC	1,3-dicyclohexyl- carbodiimide	DMP	Dess-Martin periodinane
DCM DDQ	dichloromethane 2,3-dichloro-5,6-	DMPU	<i>N</i> , <i>N</i> ′-dimethyl- <i>N</i> , <i>N</i> ′-propylene urea
	dicyano-1,4-benzo- quinone	DMS DMSO	dimethylsulfide dimethylsulfoxide
de	diastereomeric excess	DMSY	dimethylsulf-
DEAD	diethyl azodicarboxy- late	DMT	oxonium methylide dimethoxytrityl
DEPC	diethyl phosphoro- cyanidate	DNP <i>L</i> -DOPA	2,4-dinitrophenyl 3,4-dihydroxy-
DET	diethyl tartrate	DPPA	phenylalanine diphenylphosphoryl
Δ	solvent heated under reflux		azide
DHP DHPM	dihydropyran 3,4-dihydropyrim-	dppb	1,4- bis(diphenylphos-
(DHQ) <sub>2</sub> -PHAL	idin-2(1 <i>H</i> )-one 1,4-bis(9- <i>O</i> -	dppe	phino)butane 1,2-
(-11 X)2 1 11 1L	dihydroquinine)- phthalazine	11	bis(diphenylphos- phino)ethane

dppf	1,1'-bis(diphenylphos-	GC	gas chromatography
	phino)ferrocene	glyme	1,2-dimethoxyethane
dppp	1,3-bis(diphenylphos-	HOBt	1-hydroxybenzotriazole
	phino)propane	h	hour(s)
dr	diastereomeric	hν	irradiation with light
	ratio	His	histidine
E	Entgegen	HMDS	hexamethyldisilazine
	(opposite, trans)	HMPA	hexamethylphospho-
E1	unimolecular		ramide
	elimination	HMPT	hexamethylphospho-
E2	bimolecular		rous triamide
	elimination	HOMO	highest occupied
E1cb	2-step, base-induced β-		molecular orbital
	elimination via carban-	HPLC	high performance
	ion		liquid chromatography
EDA	ethylendiamine	IBCF	isobutylchloroformate
EDCI	1-ethyl-3-[3-(dimethyl-	IBX	1-Hydroxy-l,2-benzio-
	amino)propyl]carbo-		doxol-3(1H)-one
	diimide hydrochloride	Imd	imidazole
EDG	electron donating group	IPA	isopropanol
EDTA	ethylenediamine	i-Pr	isopropyl
	tetraacetic acid	KHMDS	potassium hexamethyl-
ee	enantiomeric		disilazide
	excess	kg	kilogram(s)
EEDQ	2-ethoxy-1-ethoxy-	K-selectride®	potassium tri-sec-butyl-
	carbonyl-1,2-dihydro-		borohydride
	quinoline	L	liter(s)
EMME	ethoxymethylene-	LAH	lithium aluminum
	malonate		hydride
ent	enantiomer	LDA	lithium diisopropyl-
EPP	ethyl polyphosphate		amide
eq	equivalent	LHMDS	lithium hexamethyldi-
Et	ethyl		silazide
EtOAc	ethyl acetate	LiHMDS	lithium hexamethyldi-
EPR (= ESR)	electron paramagnetic		silazide
	resonance spectroscopy	L-selectride®	lithium tri-sec-butyl-
ESR (= EPR)	electronic spin		borohydride
	resonance	LTMP	lithium 2,2,6,6-tetra-
EWG	electron withdrawing		methylpiperidine
	group	LUMO	lowest unoccupied
FMO	frontier molecular		molecular orbital
	orbital	M	metal
Fmoc	9-fluorenylmethoxy-	M	moles per liter (molar)
	carbonyl	MCR	multi-component
FVP	flash vacuum pyrolysis		reaction
g	gram(s)	m-CPBA	m-chloroperoxybenzoic
GABA	γ-aminobutyric acid		acid
Gribri	animobatyric acid		

#### xviii ABBREVIATIONS AND ACRONYMS

Me	methyl	PPA	polyphosphoric acid
MEM	β-methoxyethoxymethyl	PPE	personal protection equip-
Mes	mesitylenyl		ment
MET	methyl ethyl ketone	PPE	polyphosphoric ester
μg	microgram(s)	4-PPNO	4-phenylpyridine- <i>N</i> -oxide
μL	microliter(s)	PPP	3-(3-hydroxyphenyl)-1- <i>n</i> -
μmol	micromole(s)		propylpiperidine
mg	milligram(s)	PPSE	polyphosphoric acid
mL	milliliter(s)	1100	trimethylsilyl ester
mmol	millimole(s)	PPTS	pyridinium <i>p</i> -toluene-
MMPP	magnesium monoperoxy-	1110	sulfonate
1411411 1	phthalate hexahydrate	Pr	propyl
MO	molecular orbital	Pro	proline
mol	mole(s)	psi	pounds per square inch
MOM	methoxymethyl	PTC	phase transfer catalyst
Ms	methanesulfonyl (mesyl)	p-TSA	para-toluenesulfonic
MS	molecular sieves	<i>p</i> -13A	acid
MSDS	material safety data sheet	Py or Pyr	pyridine
MTBE	methyl <i>tert</i> -butyl ether	Ra-Ni	Raney nickel
MTPA	T	RCM	ring-closing metathesis
WITA	α-methoxy-α-trifluoro-		9
MVIII	methylphenylacetic acid	Redal-H	sodium bis(2-
MVK	methyl vinyl ketone		methoxyethoxy(aluminum
MWI (μν)	microwave irradiation	DOM	hydride)
NBS	N-bromosuccinimide	ROM	ring-opening metathesis
NCS	N-chlorosuccinimide	rt	room temperature
NIS	N-iodosuccinimide	Salen	N,N'-disalicylidene-
NMDA	N-methyl-D-aspartate	CEL (	ethylenediamine
NMM	N-methylmorpholine	SEM	2-(trimethylsilyl)
NMO	N-methylmorpholine-N-	arm.	ethoxymethyl
	oxide	SET	single electron
NMP	1-methyl-2-pyrrolidinone		transfer
NMR	nuclear magnetic reso-	$S_NAr$	nucleophilic aromatic
	nance	2000 - 20	substitution
Nu	nucleophile	$S_N 1$	unimolecular nucleophilic
PCC	pyridinium chloro-		substitution
	chromate	$S_N 2$	bimolecular nucleophilic
PDC	pyridinium dichromate		substitution
PDE	phosphodiesterase	t-Bu	<i>tert</i> -butyl
PEG	polyethylene glycol	TADDOL	$\alpha,\alpha,\alpha',\alpha'$ -tetraaryl-4,5-
pGlu	pyroglutamic acid		dimethoxy-1,3-
Ph	phenyl		dioxalane
PhFl	9-phenylfluoren-9-yl	TASF	$(Et_2N)_3S^+(Me_3SiF_2)^-$
phth	phthaloyl	TBAF	tetrabutylammonium
pKa	log acidity constant		fluoride
PMA	phosphomolybdic acid	TBD	1,5,7-triazabicyclo
PMB	para-methoxybenzyl		[4.4.0]dec-5-ene

TBDMS or TBS	tert-butyldimethyl-	THP	tetrahydropyranyl
	silyl	TIPS	triisopropylsilyl
TBDPS	tert-butyldiphenyl-	TLC	thin layer
	silyl		chromatography
TBHP	tert-butylhydro-	<b>TMEDA</b>	N, N, N', N'-tetram-
	peroxide		ethylethylenediamine
TCCA	trichlorocyanuric	TMG	tetramethylguanidine
	acid	TMP	tetramethylpiperidine
TCT	2,4,6-trichloro-	TMS	trimethylsilyl
	[1,3,5]-triazine	TMSC1	trimethylsilyl
TEA	triethylamine		chloride
TEMPO	2,2,6,6-tetramethyl-	<b>TMSCN</b>	trimethylsilyl cyanide
	1-piperidinyloxy,	TMSI	trimethylsilyl
	free radical		iodide
TES	triethylsilyl	<b>TMSOTf</b>	trimethylsilyl triflate
Tf	trifluoromethane-	Tol	toluene or tolyl
	sulfonyl (triflic)	Tol-BINAP	2,2'-bis(di-p-tolylphos-
TFA	trifluoroacetic acid		phino)-1,1'-binaphthyl
TFAA	trifluoroacetic	TosMIC	(p-tolylsulfonyl)methyl
	anhydride		isocyanide
TFE	trifluoroethanol	TPAP	tetra-n-propylammonium
TfOH	triflic acid		perruthenate
TFP	tri-o-furylphosphine	Tr	trityl
TFPAA	trifluoro peracetic acid	TRIS	tris(hydroxymethyl) aminomethane
TFSA	trifluorosulfonic acid	Ts(Tos)	p-toluenesulfonyl (tosyl)
THF	tetrahydrofuran	TSA	p-toluenesulfonic acid
THP	tetrahydropyran	TsO	tosylate
THIP	4,5,6,7-tetrahy-	$X_{\rm c}$	Chiral auxillary
	droisoxazolo[5,4-c]- pyridin-3-ol	Z	Zusammen (together, cis)



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